# What can be seen with a transmission electron microscope

Ottilie von Loeffelholz Klaholz Group, IGBMC Strasbourg

#### The transmission electron microscope

Thungsten filament







apertures

- vacuum: ~10<sup>-6</sup> Pa
- potentially high electron dose
- potentially high resolution
  (λ≈ 0.027Å at 200kV)
- i.e. <u>not limited by the</u>

<u>wavelength</u> or the optical system



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Field emission gun (FEG) electron microscope (Tecnai20, IGBMC)

# Image formation with an electron microscope



Orlova and Saibil, Chem. Rev., 2011

## Electron microscopy techniques Negative staining

1) adsorption



2) wash with 2% uranyl acetate



3) air-dry



Heavy metal stains: ammonium molybdate, uranyl acetate, uranyl formate, phosphotungstic acid (PTA), auroglucothionate and others ...

> Drawn by J-F. Ménétret Slide from Bruno Klaholz

### Electron microscopy techniques Negative staining



- Fast
- Technique for low resolution information
- 3D reconstruction is a "shell"
- Good for screening

Joachim Frank, Quaterly Reviews of Biophysics, 1990

## Electron microscopy techniques Shadowing technique of supercoiled DNA



Patrick Schultz

- Heavy atoms evaporated on a sample under an angle

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#### Electron microscopy techniques Cryo electron microscopy Purified human ribosomes



Khatter *et al.*, *Nucl. Ac. Res.*, 2014 - Full preservation of sample in vitreous ice, potential to reach high resolution *Slide from Bruno Klaholz* 

#### Electron microscopy techniques electron diffraction – 2D



Plisson *et al.* (2003); *JBC*, **278**, 23753–23761.

Ruprecht et al. (2004); EMBO, 23, 3609-3620

Slide from → e.g. transmembrane helices visible Bruno Klaholz Limitation of 2D crystals: usually <u>anisotropic</u> resolution

#### Electron microscopy techniques electron diffraction – 3D



Lysozyme nanocrystal seen in TEM

-May be alternative to X-ray crystallography



Diffraction of lysozyme

Clabbers et al., Acta Cryst D, 2017

# Electron microscopy techniques Tomography – plastic embedding



- Fast freezing that prevents ice formation in thicker samples
- Limited resolution, artifacts from sectioning?

Leica Microsystems, MiTeGen

## Electron microscopy techniques cryo electron tomography in cells



- Great potential of the technique, but very tidious

Rigort, PNAS, 2012

### Changing focus in the microsocope





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#### The contrast transfer function (CTF)



Contrast transfer function (CTF)

The defocus image is <u>convoluted</u> by the CTF Result:

- spreading of each pixel over a bigger surface
- Inversion of contrast of some pixels

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# Deconvolution of the CTF: Fourier transform in image processing



Images taken from: https://spider.wadsworth.org/spider\_doc/spider/docs/exa/pw.html

#### Power spectrum = Amplitude spectrum



Real image

Power spectrum

#### Orlova and Saibil, Chem. Rev., 2011

## Changing focus in the microsocope



#### **Contrast generation by Defocussing**



-1.5 µm







Dubouchet, Quaterly review of Biophysics, 1988



#### **Objects seen in Fourier space**



Layer lines: Tubulin is 40 Å big and Kinesin binds every 80 Å on the MT lattice

von Loeffelholz et al., Nature Comm., 2017

#### Image contrast in cryo-EM

**Amplitude contrast (inelastic scattering, absorption)** 



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#### Contrast generation with phase plates



von Loeffelholz and Klaholz, 3rd Edition Meth. in Mol Biol. –Structural Proteomics, submitted; von Loeffelholz, JSB, 2017

#### Contrast generation with phase plates



With phase plate in focus

Without phase plate -1.6 µm defocus

Danev et al, Elife, 2016

#### Cryo electron microscopy

#### $\downarrow \downarrow \downarrow$



Joachim Frank, Annu. Rev. Biophys. Biomol. Struct. 2002

### Electron micrograph of particles



von Loeffelholz et al, PNAS, 2015

### **Class averages of particles**



representative views of the 70S / RF2 complex; Klaholz et al., Nature 2003.

# Angle assignment to the class averages or individual particles



Slide adapted from Bruno Klaholz

#### Backprojection – 3D Reconstruction





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