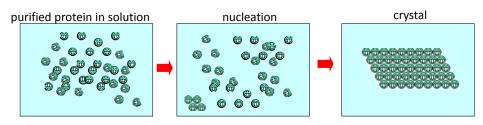
# Xray diffraction by molecular crystals: a (short?) introduction

Laurent Maveyraud, Oléron 2016

# Crystallogenesis of proteins



Many crystallization assays (>>1000) are required to obtain suitable protein crystals.

Crystallization usually performed by slow evaporation of water (various pH, precipitating agents...).

#### Methodology:

Preliminary screens (96-solutions kits, robotized)

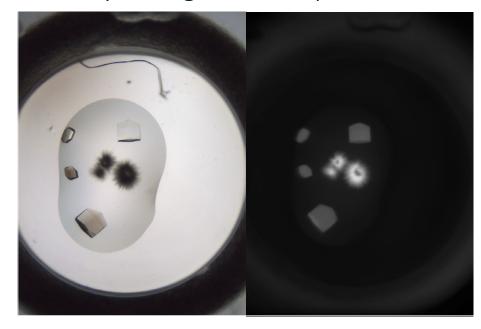
Crystal optimization of (24-wells, manually set up)

Further readings: McPherson, 2014, Acta Cryst F70:1445 McPherson, 2004, Methods, 34:254

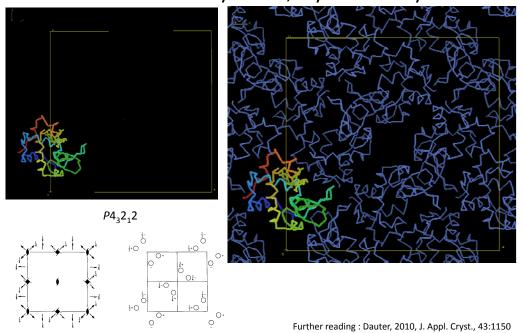
#### overview

- Crystals: How to get them? What do they look like?
- Some theory about diffraction: structure factors, reciprocal lattice, Ewald's sphere
- Data collection: crystal conditionning, practical aspects
- Data processing: XDS, mosflm, assessing data quality
- "Stéphane, how do we solve a structure with these data?"

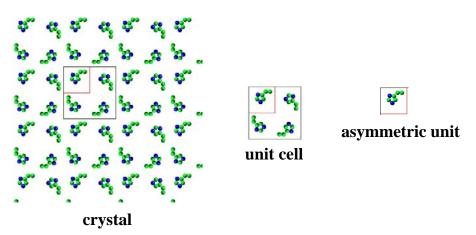
#### Crystallogenesis of proteins



#### Protein crystals, symmetry



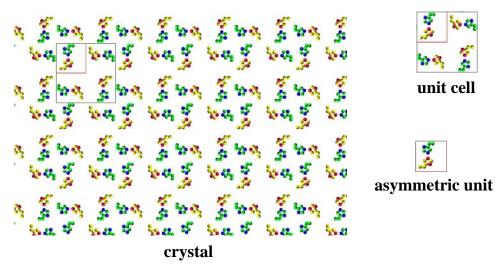
#### Protein crystals and symmetry



As proteins are chiral, only rotation and translations allowed in protein crystals: 65 possible space groups.

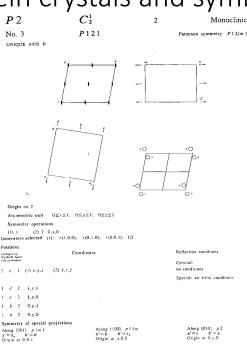
Symmetry results in equivalent positions.

#### Protein crystals and symmetry

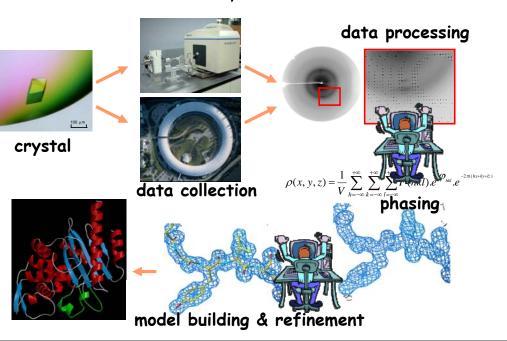


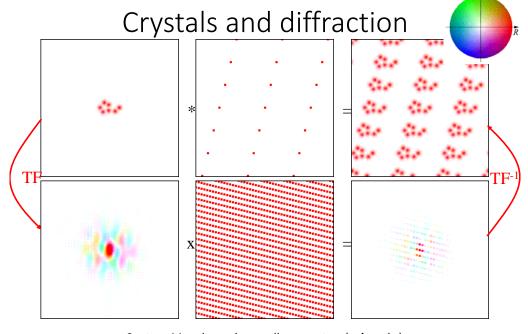
# You can have more than one copy of the protein in the asymmetric unit (Non Crystallographic Symetry)

#### Protein crystals and symmetry



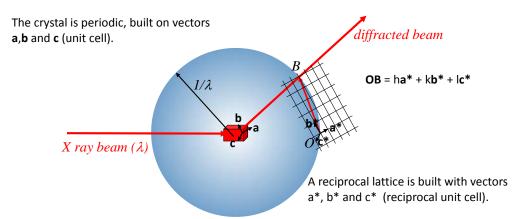
### You have a crystal? So what?





Spot position depends on cell parameters (**a**, **b** and **c**) Spot intensity depends on the structure of the molecule

#### Crystals and diffraction: Ewald's sphere



A wave is scattered when a node of the reciprocal lattice (indices h k l) touches the Ewald's sphere. The structure factor (amplitude F and phase  $\phi$ ) of the diffracted wave is :

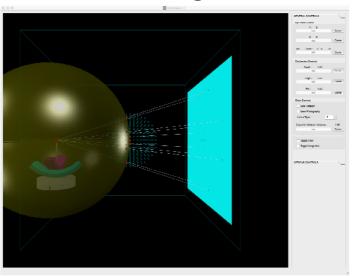
$$\mathbf{F}(hkl) = N_{cell} \cdot \Sigma f_{j} \cdot \exp(-2\pi(hxj + kyj + lzj))$$

## Getting ready for data collection

• Xrays can fry your crystals: better cool them!



#### Collecting data

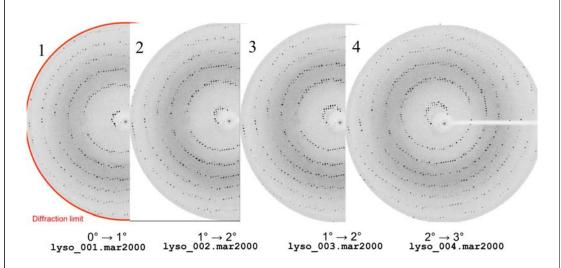


You want to be sure to collect every diffracted beam! That is, all nodes of the reciprocal lattice should hit the Ewamd's sphere: rotate the crystal while exposing it to Xray

#### Collecting data: the oscillation method

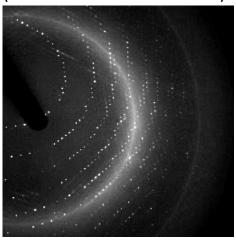
- How many images to collect ?
  - Crystal symmetry, phasing method
- Which oscillation angle ?
  - Cell parameters, type of detector, type of processing
- Which crystal to detector distance?
  - Resolution limit of the crystal, cell parameters
- Which exposure time?
  - Type of detector, no saturated spots

#### Collecting data



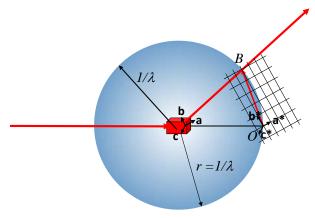
#### Collecting data: the oscillation method

With recent detectors (Pilatus) the crystal is rotated continuously (shutterless data collection).



Further readings: Evans, 1999, Acta Cryst, D55:1771 Dauter, 1999, Acta Cryst, D55:1703

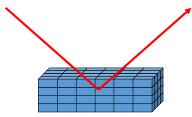
#### Collecting data: the oscillation method



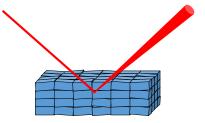
#### In theory:

- perfect crystal: reciprocal lattice is built of points
- perfect beam (no wavelength dispersion, no divergence...)

#### Collecting data: let's face reality



- perfect crystal: reciprocal lattice is built of points
- perfect beam (no wavelength dispersion, no divergence...)

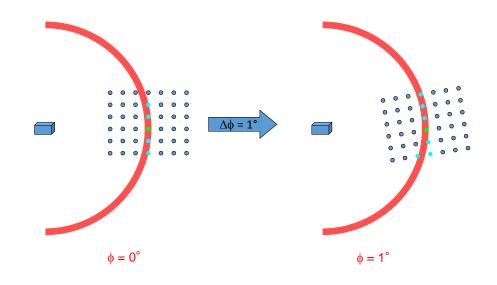


#### Real life:

- mosaic crystal
- real beam (wavelength dispersion, divergence...)

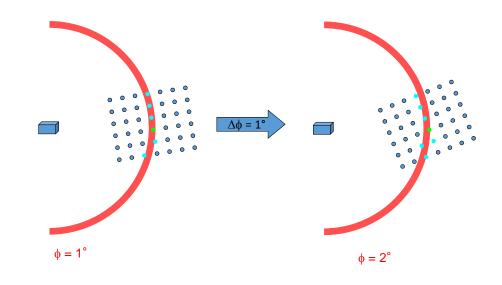
#### Collecting data: let's face reality

Consequences for the Ewald's construction



#### Collecting data: let's face reality

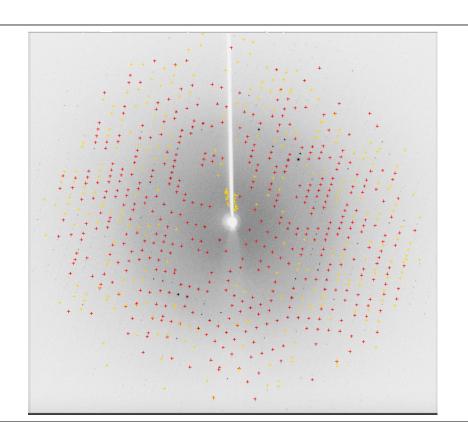
Consequences for the Ewald's constuction

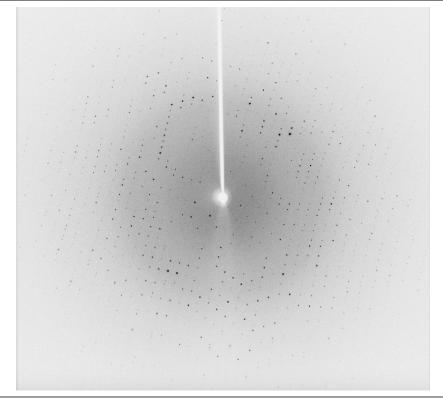


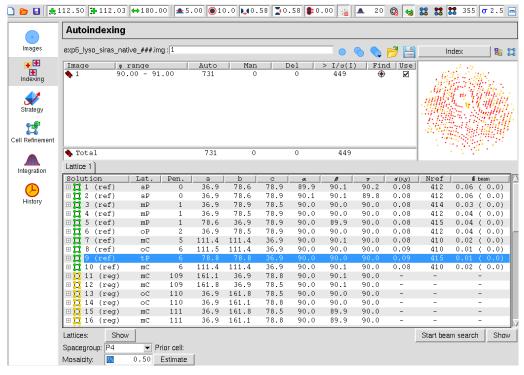
#### Processing data: XDS, iMosflm)

Three steps for data processing:

- Indexing data: find possible cell parameters, crystal orientation, guestimate symmetry
  - For each diffraction spot, you know Miller indices
  - Symmetry derived from cell parameters: it's only a hypothesis !!!!







#### Processing data: XDS, iMosflm)

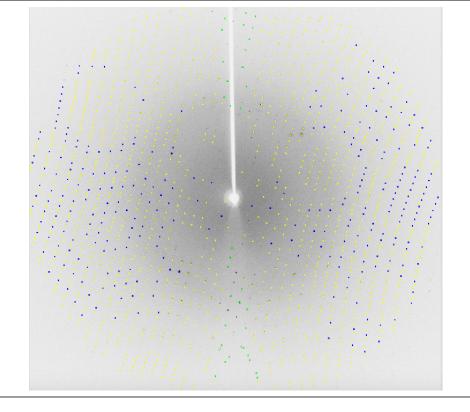
Three steps for data processing:

- Indexing data: find possible cell parameters, crystal orientation, guestimate symmetry
  - For each diffraction spot, you know Miller indices
  - Symmetry derived from cell parameters: it's only a hypothesis !!!! If the cells seems to obey to some symmetry constraints, it's likely because this symmetry is present in the crystal.
- Now that we have a unit cell and an orientation, we can predict spot position on any frames

#### Processing data: XDS, iMosflm

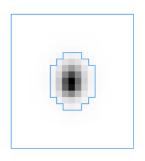
Three steps for data processing:

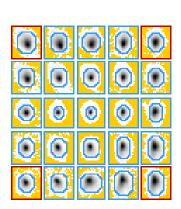
- Indexing data: find possible cell parameters, crystal orientation, guestimate symmetry
  - For each diffraction spot, you know Miller indices
  - Symmetry derived from cell parameters: it's only a hypothesis !!!!
- Integration: for each spot on each frames, measure the intensity
  - Locate spot, assign pixel to « background » or to « spot »
  - Sum the intensity for « spot » pixels
  - Profile fitting (2D iMosflm, 3D XDS)



#### Processing data: iMosflm

2D profile fitting: wide slicing

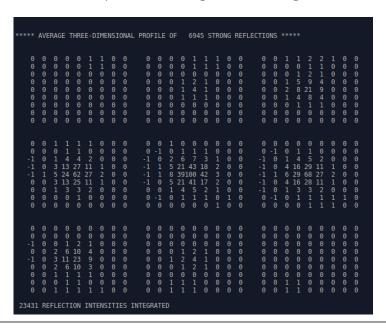




Detector surface splitted in 9 or 25 regions. Profiles are learned for intense well defined spots.

#### Processing data: XDS

3D profile fitting: fine slicing

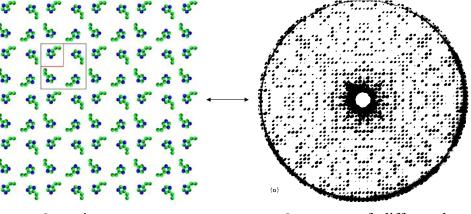


#### Processing data: XDS, iMosflm

#### Three steps for data processing:

- Indexing data: find possible cell parameters, crystal orientation, guestimate symmetry
  - For each diffraction spot, you know Miller indices
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- Integration: for each spot on each frames, measure the intensity
  - Locate spot, assign pixel to « background » or to « spot »
  - Sum the intensity for « spot » pixels
  - Profile fitting (2D iMosflm, 3D XDS)
- Scaling of data: correct for variation in diffracting volume, beam intensity variations,...
  - Use equivalent reflections to place all images: uses the symmetry of the crystal!

#### Symmetry of reciprocal space



#### Crystal symmetry: equivalent positions

x, y, z y, -x, z

-x, -y, z -y, x, z

Symmetry of diffracted intensities: equivalent reflections

> h, k ,l k, -h, I

-h, -k, l -k, h l

-k, h, -l

-h, -k, -l

k, -h, -

## Processing data: XDS, iMosflm

#### Three steps for data processing:

- Indexing data: find possible cell parameters, crystal orientation, guestimate
  - For each diffraction spot, you know Miller indices
  - Symmetry derived from cell parameters: it's only a hypothesis !!!!
- Integration: for each spot on each frames, measure the intensity
  - Locate spot, assign pixel to « background » or to « spot »
  - Sum the intensity for « spot » pixels
  - Profile fitting (2D iMosflm, 3D XDS)
- Scaling/merging of data:
  - Scaling: correct for variation in diffracting volume, beam intensity variation,. Use the symmetry of the crystal (validate, or not, the symmetry hypothesis from the indexing step)
  - Merging: average different observations of equivalent reflections, compute data processing statistics

#### Checking the quality of your data

SUBSET OF DESCRIPTION LIMIT		ATA WITH OF REFL UNIQUE		ISE >= -3.0 AS COMPLETENESS OF DATA			UTION COMPARED	I/SIGMA	R-meas	CC(1/2)	Anomal Corr
5.35	6059	778	779	99.9%	2.1%	2.7%	6059	67.08	2.3%	100.0*	54*
3.80	10814	1395	1395	100.0%	2.7%	2.7%	10814	67.86	2.9%	99.9*	26*
3.11	13860	1797	1797	100.0%	2.9%	2.8%	13860	63.55	3.1%	99.9*	16*
2.69	16578	2139	2139	100.0%	3.4%	3.4%	16578	49.96	3.7%	99.9*	6
2.41	18603	2406	2406	100.0%	4.2%	4.1%	18603	42.43	4.5%	99.9*	5
2.20	20632	2675	2675	100.0%	4.9%	4.9%	20632	35.82	5.2%	99.9*	8
2.04	22300	2899	2899	100.0%	6.0%	6.1%	22300	29.20	6.4%	99.8*	2
1.91	23848	3113	3113	100.0%	8.4%	8.7%	23848	21.33	9.0%	99.7*	5
1.80	24479	3304	3312	99.8%	12.2%	13.0%	24467	14.55	13.1%	99.4*	1
total	157173	20506	20515	100.0%	3.9%	3.9%	157161	37.30	4.2%	99.9*	7

# Checking the quality of your data

Table 1

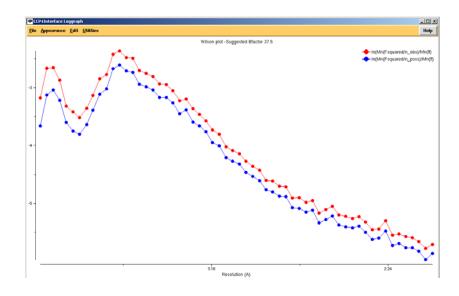
	55.70 – 1.80 Å	1.84 – 1.80 Å		
N observations	156,728	8,565		
N unique	11,204	646		
Multiplicity	14.0	13.3		
Completeness (%)	100.0	100.0		
Rsym or Rmerge	0.053	0.145		
Ι/σ	34.8	15.2		

Is Rsym/Rmerge a good indicator of data quality?

### Checking the quality of your data

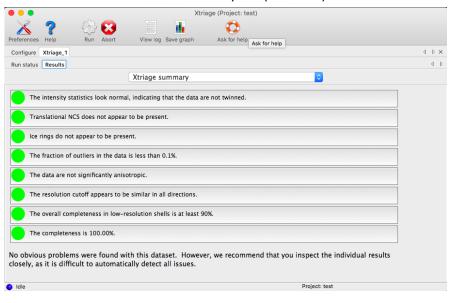
	55.70 – 1.80 Å	1.84 – 1.80 Å		
N observations	156,728	8,565		
N unique	11,204	646		
Multiplicity	14.0	13.3		
Completeness (%)	100.0	100.0		
Rsym or Rmerge	0.053	0.145		
Rmeas	0.057	0.155		
CC1/2	0.999	0.995		
Ι/σ	34.8	15.2		

# Checking the quality of your data Wilson Plot



## Crystal/dataset pathologies

XTRIAGE analysis (Phenix)



What can we do with these data?

Stéphane... tell us about phases

h	k	1	F	SIGF	DANO	SIGDANO	F(+)	SIGF(+)	F(-)	SIGF(-)
0			0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.0
0	0	2	-1.00	0.00	-1.00	0.00	-1.00	0.00	0.00	0.0
0	0	3	-1.00	0.00	-1.00	0.00	-1.00	0.00	0.00	0.0
0	0	4	101.12	6.29	0.00	0.00	100.92	9.00	100.05	9.1
0	0	5	5087.18	868.91	5087.18	868.91	5087.18	868.91	5004.75	871.4
0	0	6	-1.00	868.91	-1.00	868.91	-1.00	868.91	5004.75	871.4
0	0	7	-1.00	868.91	-1.00	868.91	-1.00	868.91	5004.75	871.4
0	0	8	712.77	26.26	0.00	0.00	713.90	35.18	706.38	40.0
0	0	9	251303.12	24365.59	251303.12	24365.59	251303.12	24365.59	246856.75	27390.6
0	0	10	-1.00	24365.59	-1.00	24365.59	-1.00	24365.59	246856.75	27390.6
0	0	11	-1.00	24365.59	-1.00	24365.59	-1.00	24365.59	246856.75	27390.6
0	0	12	374.42	11.63	0.00	0.00	377.39	14.45	367.19	19.8
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36	20	1	239.06	4.01	-32.37	8.15	221.41	6.19	253.78	5.3
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