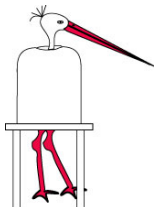


NMR developpments



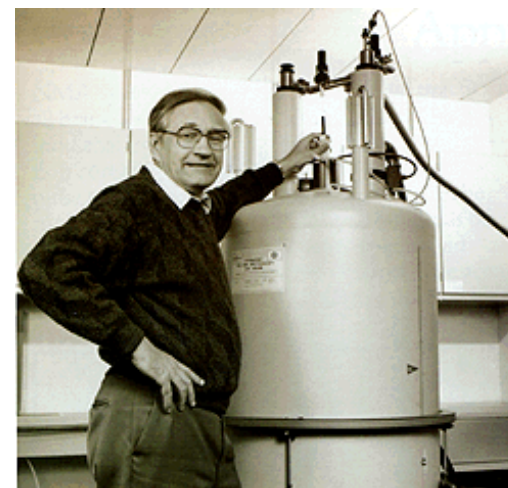
Bruno Kieffer (kieffer@igbmc.fr)
Groupe de RMN biologique
ESBS-IGBMC
Illkirch



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NMR developments: a tribute to R.R. ERNST



Why just NMR ?

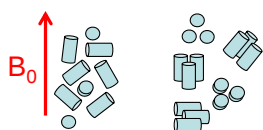
- « Because there is hardly another technique that is so informative for so many different types of applications, and because there is no other technique that provides so much fun ».

Richard Robert
ERNST

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Three States of living matter

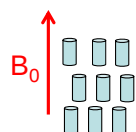
Solid state



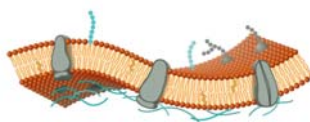
Static disorder



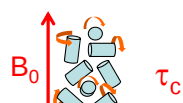
Solid state Oriented



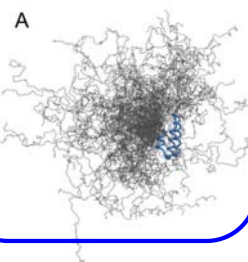
Static Order



Solution



Dynamic disorder

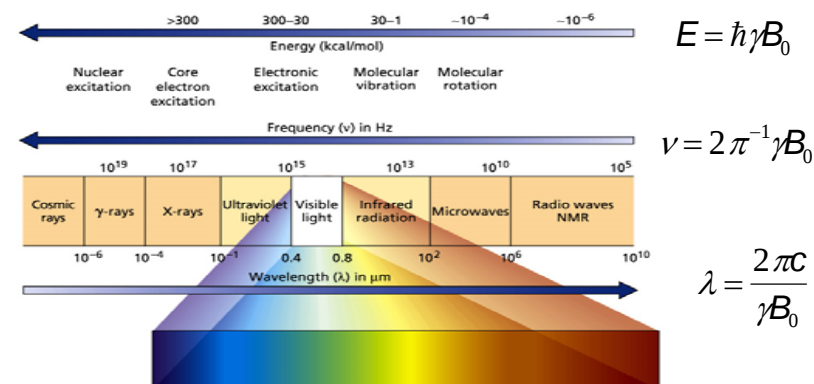


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Few introductory concepts about NMR

- NMR is a very very low energy spectroscopy

Thermal energy
0,5 kcal/mol



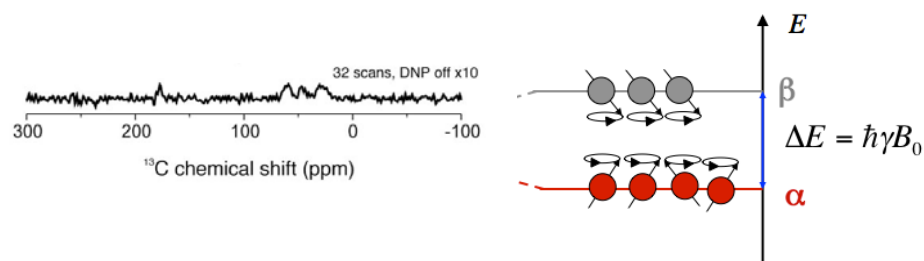
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Low energy means...

- **Intrinsically weak signal** due to low population difference

Boltzman statistic:

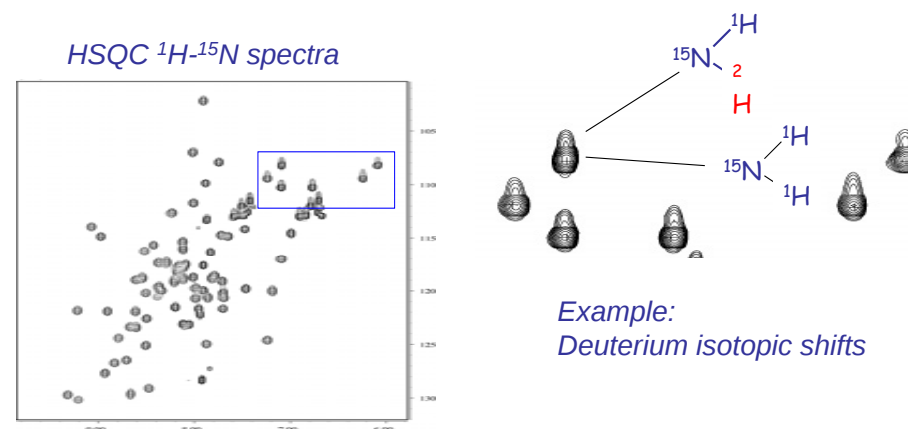
$$\frac{P_{\beta}}{P_{\alpha}} = e^{\frac{-\Delta E}{kT}} = e^{\frac{-\gamma\hbar B_0}{kT}} \approx 1 - \underbrace{\frac{\gamma\hbar B_0}{kT}}_{10^{-4}}$$



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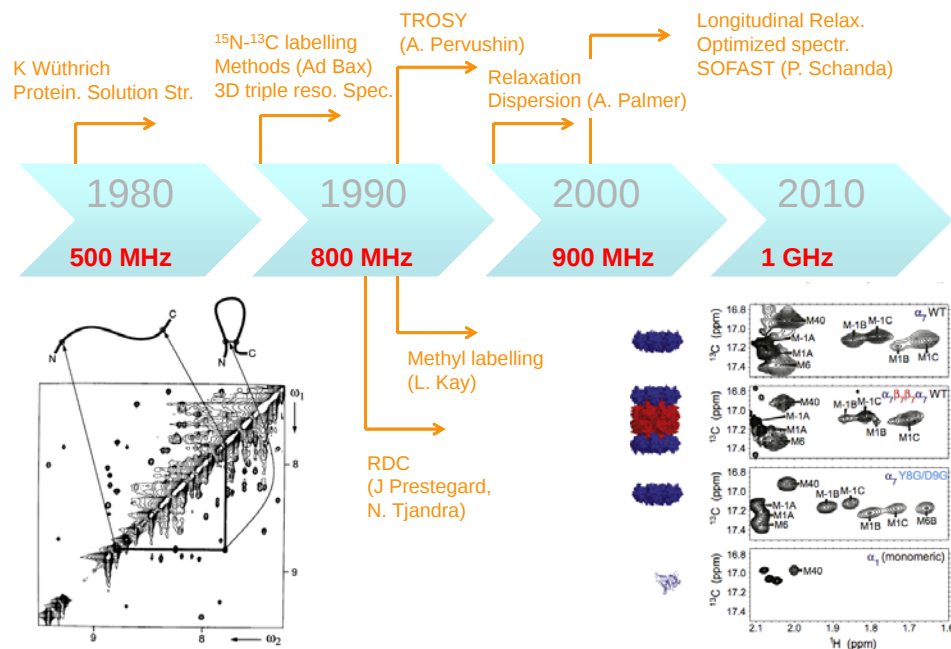
Low energy means...

- Very long live time of excited state...
- Subsequently an **extreme precision** on measurements

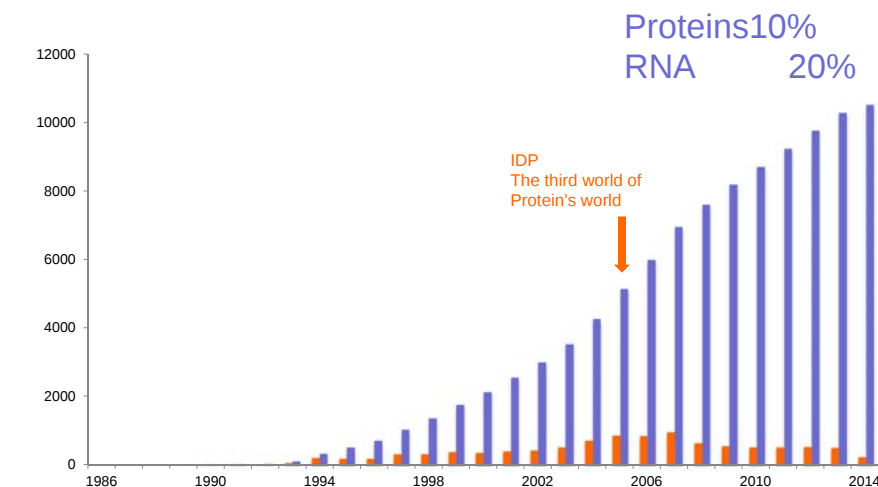


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Biological NMR: 35 Years of innovations

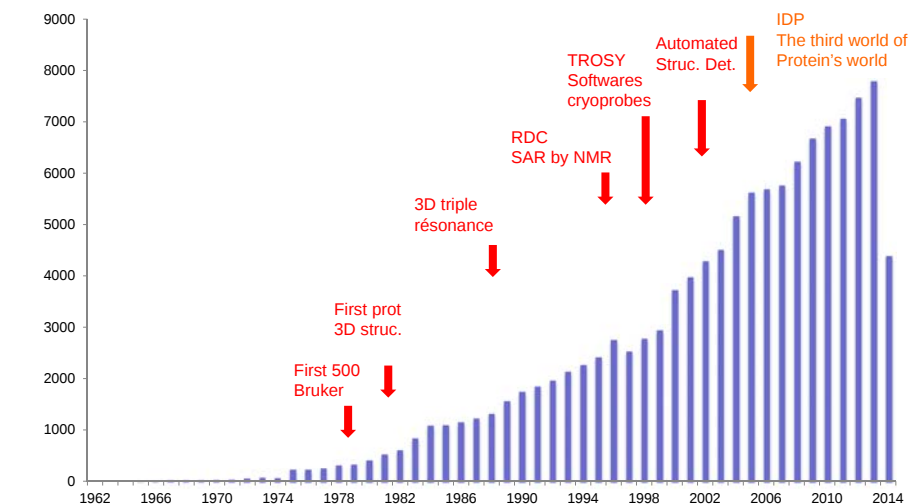


Nb of structures determined by NMR in the PDB



ReNaFobis 2016

PUBMED publications with NMR in the title



Jan 2013

The Quiet Renaissance of Protein Nuclear Magnetic Resonance

Paul J. Barrett,[†] Jiang Chen,[†] Min-Kyu Cho,[†] Ji-Hun Kim,[†] Zhenwei Lu,[†] Sijo Mathew,[†] Dungeng Peng,[†] Yuanli Song,[†] Wade D. Van Horn,^{†,§} Tiandi Zhuang,[‡] Frank D. Sönnichsen,^{||} and Charles R. Sanders^{*,†}

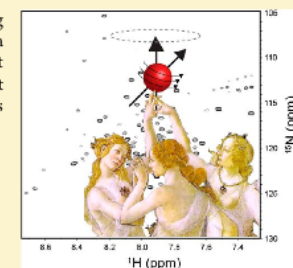
[†]Department of Biochemistry and Center for Structural Biology, Vanderbilt University, Nashville, Tennessee 37232-8725, United States

[‡]Department of Molecular Physiology and Biological Physics, University of Virginia, Charlottesville, Virginia 22908, United States

[§]Department of Chemistry and Biochemistry, Arizona State University, Tempe, Arizona 85287-1604, United States

^{||}Institute for Organic Chemistry, Christian-Albrechts University of Kiel, D-24118 Kiel, Germany

ABSTRACT: From roughly 1985 through the start of the new millennium, the cutting edge of solution protein nuclear magnetic resonance (NMR) spectroscopy was to a significant extent driven by the aspiration to determine structures. Here we survey recent advances in protein NMR that herald a renaissance in which a number of its most important applications reflect the broad problem-solving capability displayed by this method during its classical era during the 1970s and early 1980s.



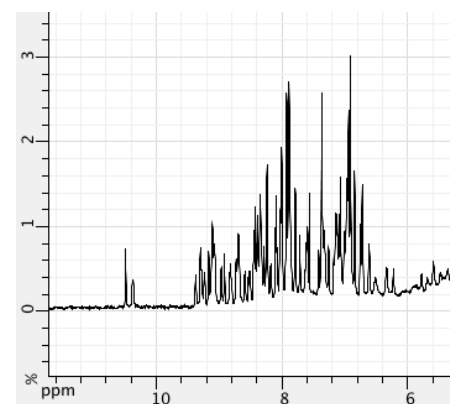
2005-2015

NMR: a tool for integrative structural biology

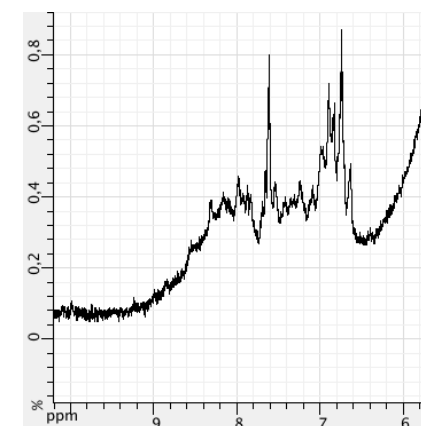
- Study of Intrinsically Disordered Proteins (IDP)
- Study of Molecular Recognition fundamental mechanisms
- Description of Protein and Nucleic Acid excited states
- Visualizing Large complexe's motions
- Monitoring protein's states within the cell

Recent methodological developments aim at:

- solving resolution problem (size limitation)
- sensitivity issues

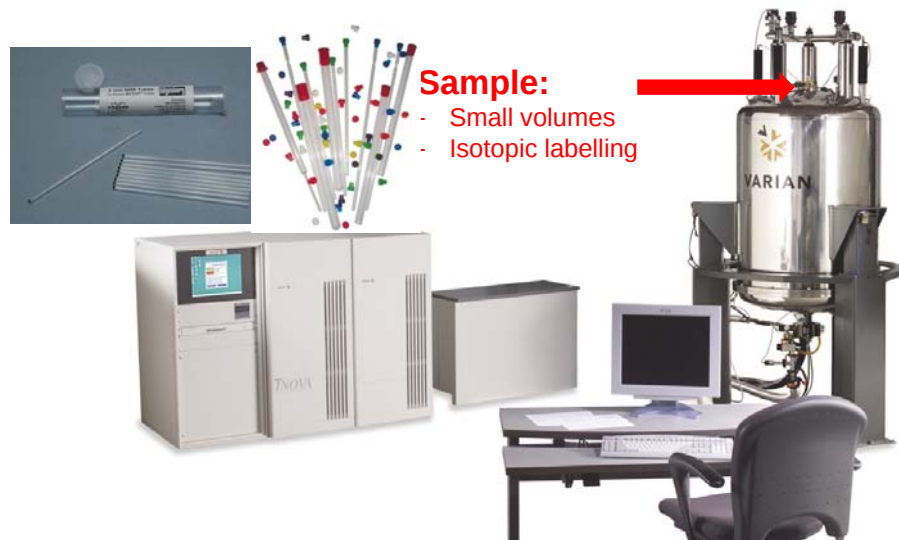


GB1 (~6 kDa)



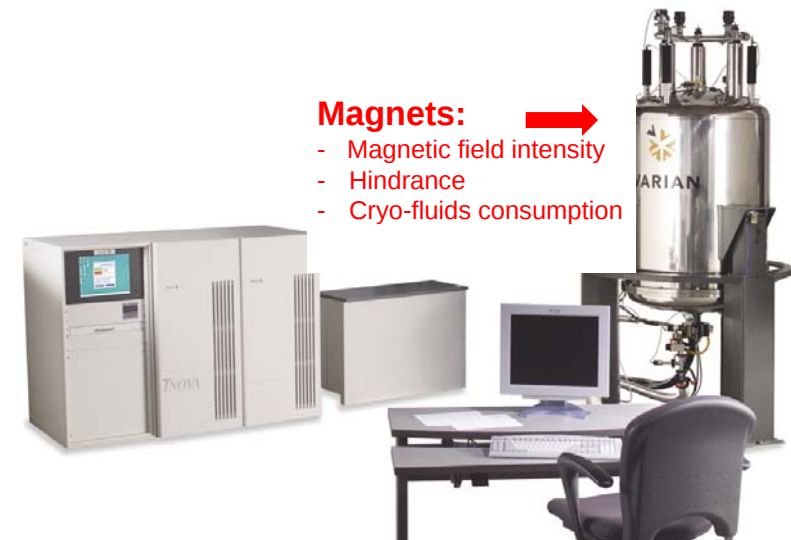
Dimère LBD RXR
(~50 kDa)

NMR Methodological Innovations



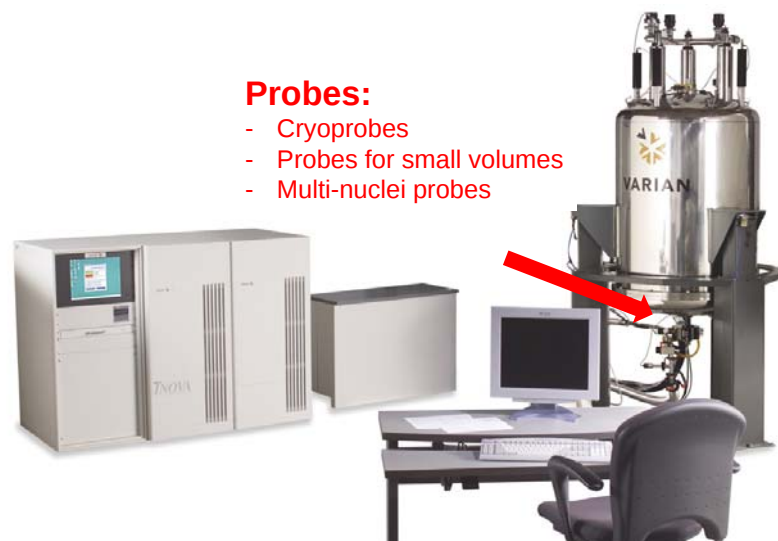
ReNaFobis 2016

NMR Methodological Innovations



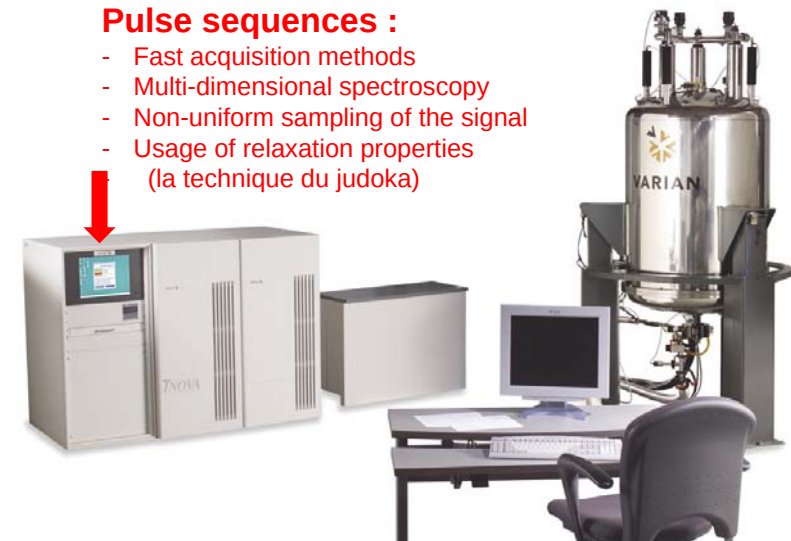
ReNaFobis 2016

NMR Methodological Innovations



ReNaFobis 2016

NMR Methodological Innovations



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NMR Methodological Innovations

Numerical tricks:

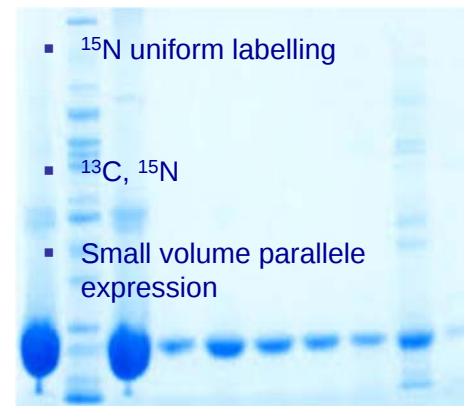
- Filtering
- Data processing and handling
- Modelling softwares



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NMR: co-evolution of biochemical and spectroscopic methods

- Unlabelled proteins
- ^{15}N uniform labelling
- ^{13}C , ^{15}N
- Small volume parallel expression
- 2D spectroscopy
- Heteronuclear Single Quantum Correlation (HSQC)
- HNCA, HNCO
- micro-cryoprobe



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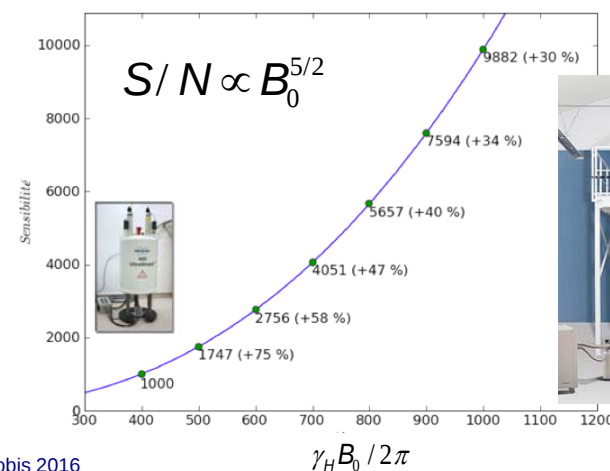
1. Recent advance in NMR instruments



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Recent advance in NMR instruments

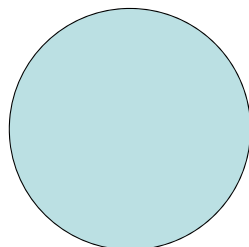
- The Signal/Noise ratio depends on the applied magnetic field



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Recent advance in NMR instruments

- Compactness of magnets



Non - shielded magnet



Shielded magnet

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Cryoprobes



- Provided that the sample is properly conditioned (tubing) the sensitivity can be optimized (mass of matter needed for the experiment)

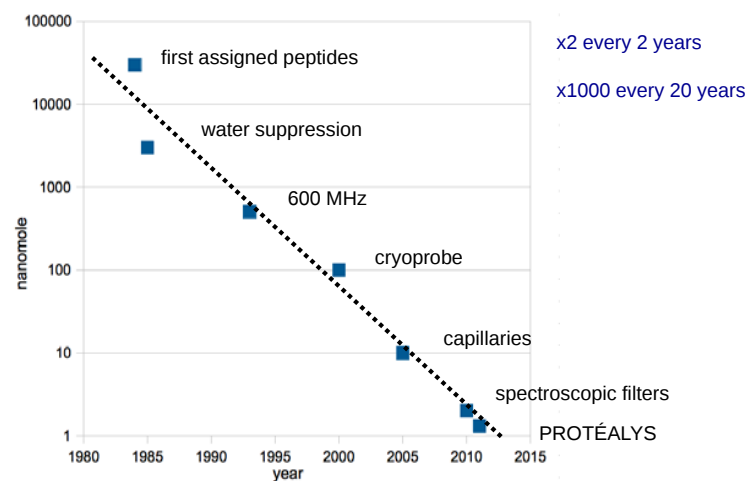
- Hoult law: $S/N \propto Q\eta M_0$

Q : facteur de qualité
 η : facteur de remplissage

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Evolution of Detection Sensitivity

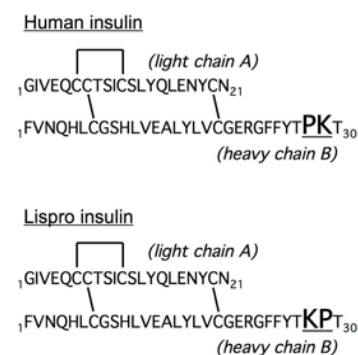
Quantity of measured protein



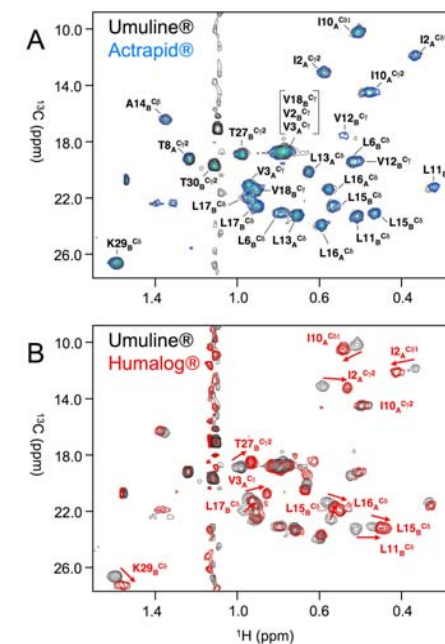
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From MA Delsuc

Bio-drugs quality control by NMR



Brevet Kieffer B. (2011) WO 2011/080492

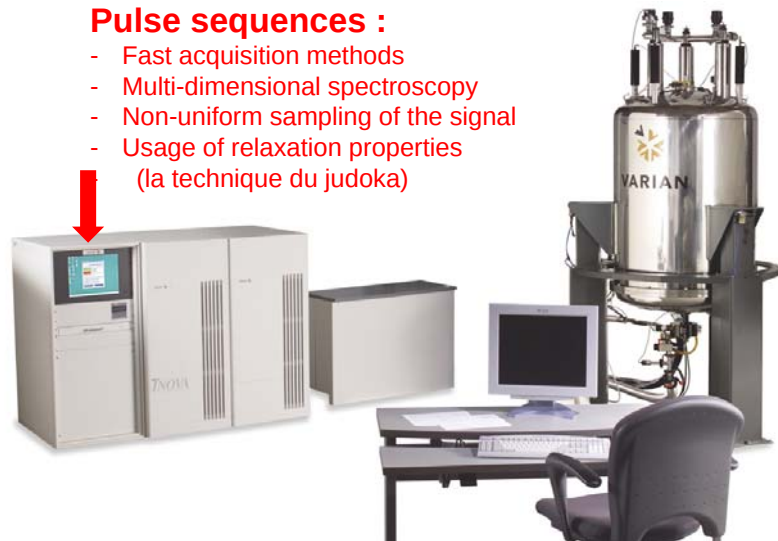


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2. Recent advance in NMR methodologies

Pulse sequences :

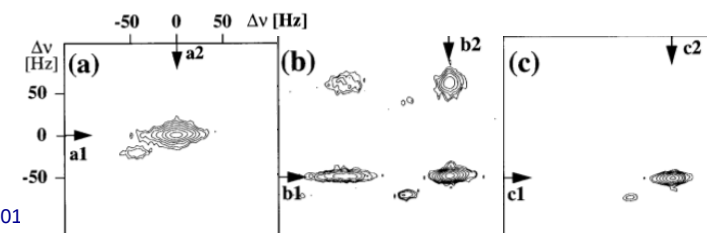
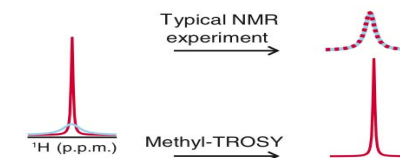
- Fast acquisition methods
- Multi-dimensional spectroscopy
- Non-uniform sampling of the signal
- Usage of relaxation properties (la technique du judoka)



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Taking advantage of relaxation properties

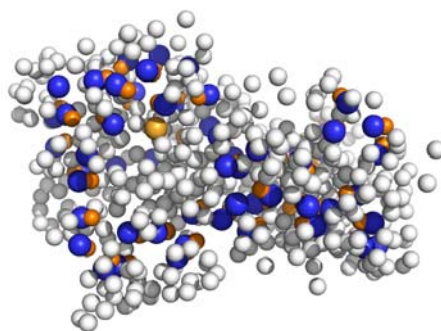
- 1. Transverse relaxation
 - Exploit cross-correlation phenomena between several relaxation mechanisms (CSA-DD) => TROSY



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Taking advantage of relaxation properties

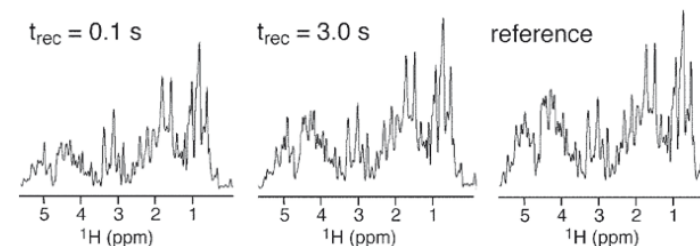
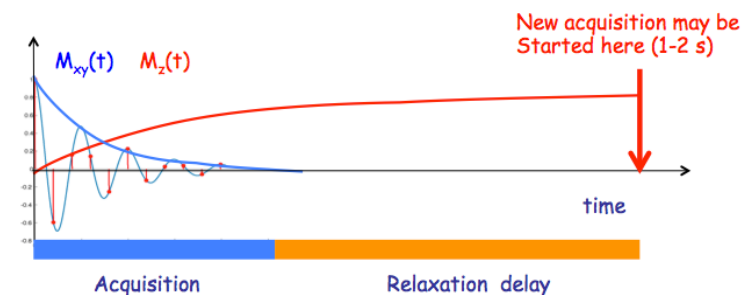
- 2. Longitudinal relaxation:
 - Accelerate the return to thermal equilibriumstate before the next acquisition => SOFAST



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P Schanda et al. 2005

Taking advantage of relaxation properties

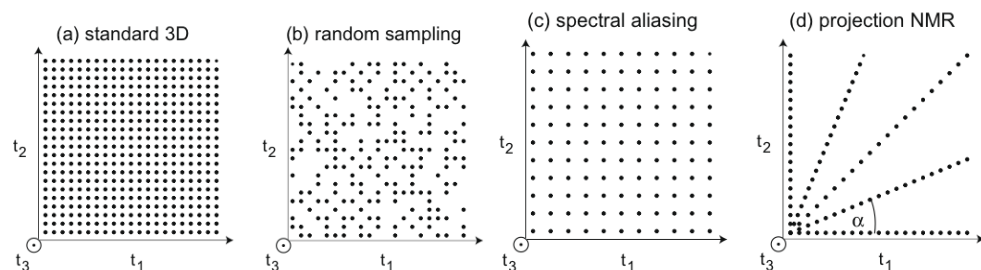


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P Schanda et al. 2005

Working on sampling methods

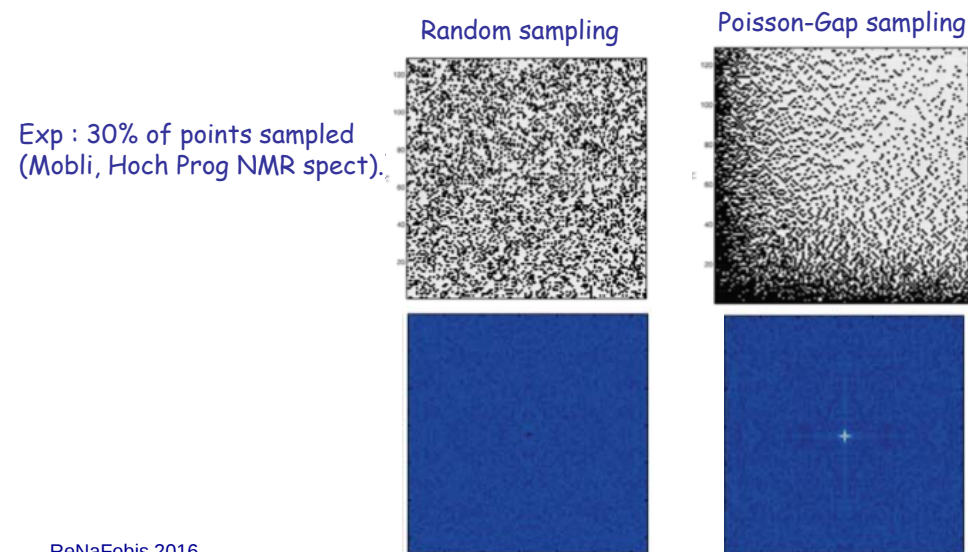
- The use of FFT imposes a linear sampling
- Alternate methods (NUS) are currently being developed allowing considerable gain of time



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P Schanda Progress NMR Spect. 2009

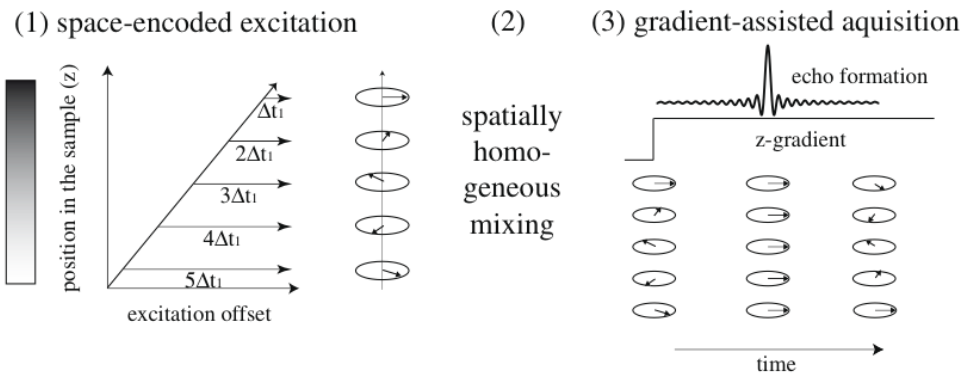
Sampling 3D spaces



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Ultra-Fast NMR

- Principle: Use field gradients to establish a relationship between space and time domain
 $\Rightarrow$ A 2D can be recorded in few seconds (provided that you have enough S/N) (L. Friedman et al. PNAS 2002)

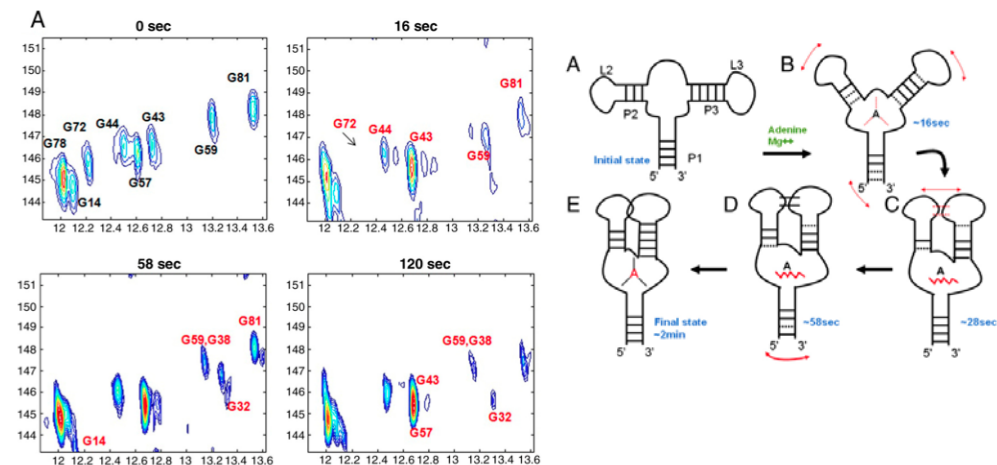


P Schanda Progress NMR Spect. 2009

Application: Real-time follow-up of a folding process

Real-time multidimensional NMR follows RNA folding with second resolution

Mi-Kyung Lee¹, Maayan Gal¹, Lucio Frydman^{1,2}, and Gabriele Varani^{1,2} PNAS 2010



3. Sample preparation

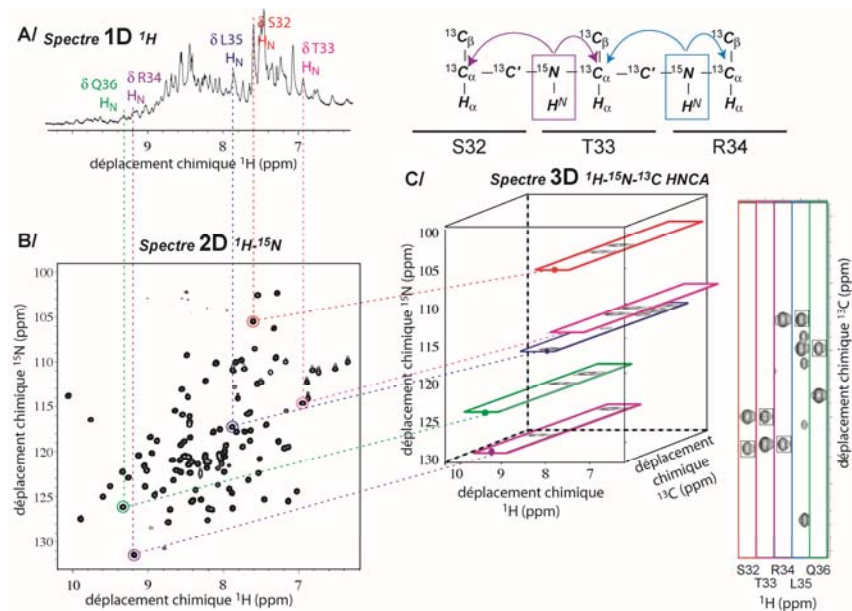


Sample preparation

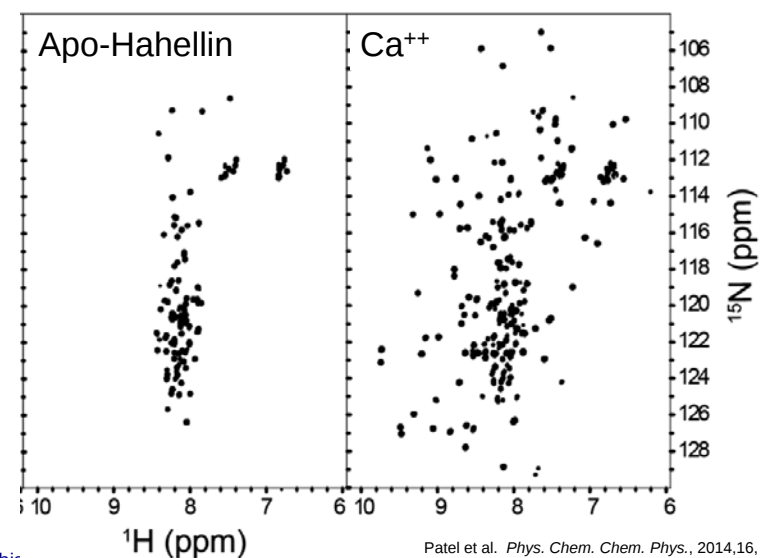
- Isotopic Labelling:
 - Spectral Simplification (reduction of the number of frequencies present in the spectrum)
 - Reduce the relaxation sources (Partial deuteration)
- Introduction of additional probes
 - Paramagnetic Relaxation
 - Fluorine NMR
- The complexity of observed medium
 - Use of anisotropic media (Residual Dipolar Couplings)
 - In-cell NMR

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Standard approach: Uniform ^{15}N - ^{13}C associated to multi-dimensional triple resonance spectroscopy



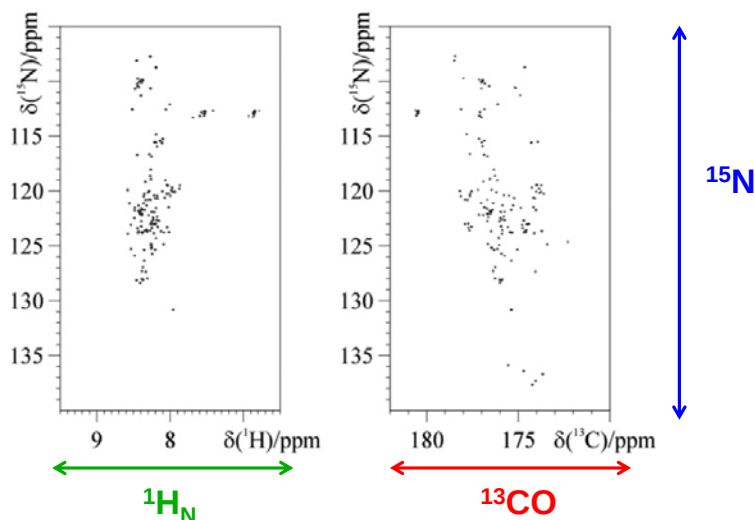
Development of specific methods for Intrinsically disordered proteins (IDP)



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Patel et al. *Phys. Chem. Chem. Phys.*, 2014,16, 12703-12718

^{13}C , ^{15}N detection



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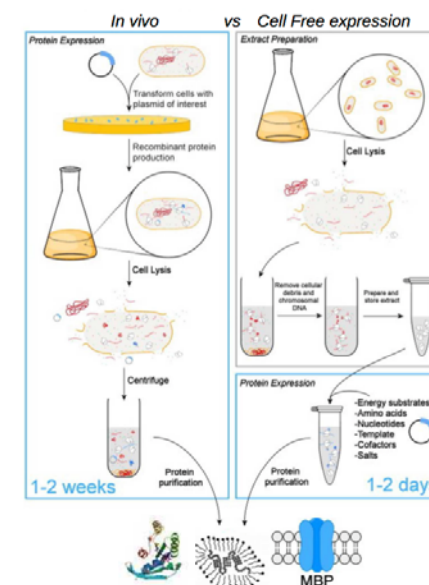
I. Felli et al. IUBMB Life, 64(6): 473–481, June 2012

Cell free expression and combinatorial labelling



Lionel Imbert (IBS) Cell-Free expression platform
Adapted from Carlson et al. 2012 Kigawa et al. 2001
Schwartz et al. 2007

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Overview of cell free isotopic labeling



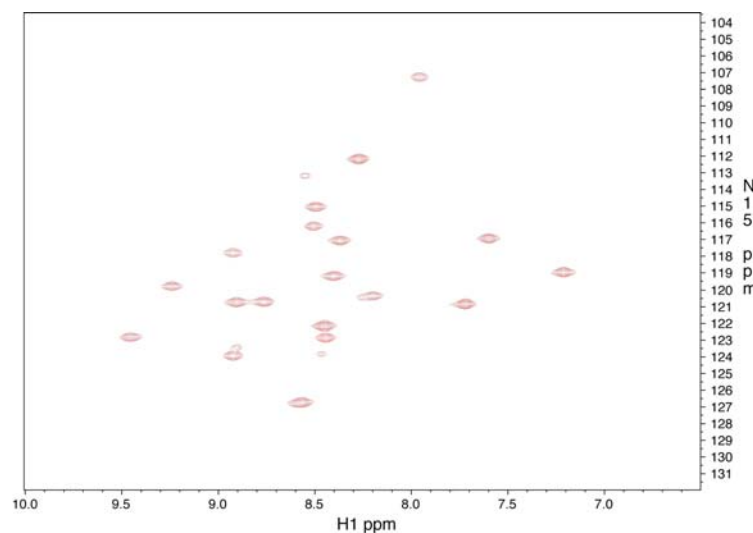
Specific amino acids labeling

CECF
Val, Thr ^{15}N , ^{13}C
+ ^{18}O aa unlabeled

Expected peaks: 21
13 Val
8 Thr

Observed: 22

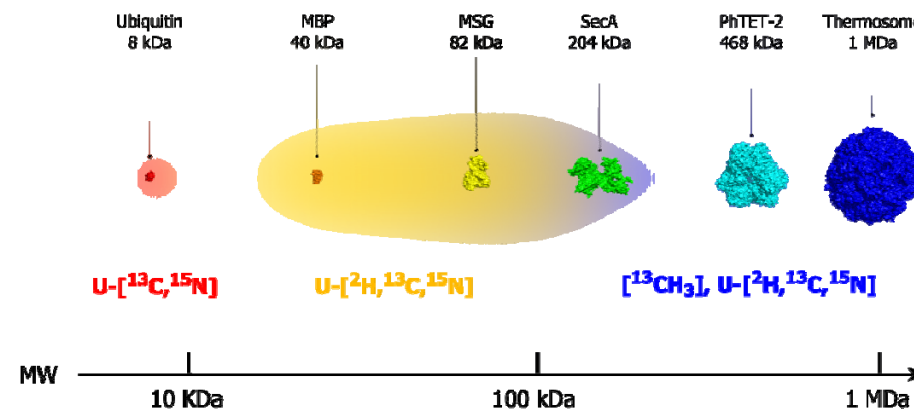
Yield: 4 mg/ml



^1H SQC 600 MHz ^1H 23 (16 kDa)

Lionel Imbert (IBS) Cell-Free expression platform

Is NMR spectroscopy only limited to small molecules?

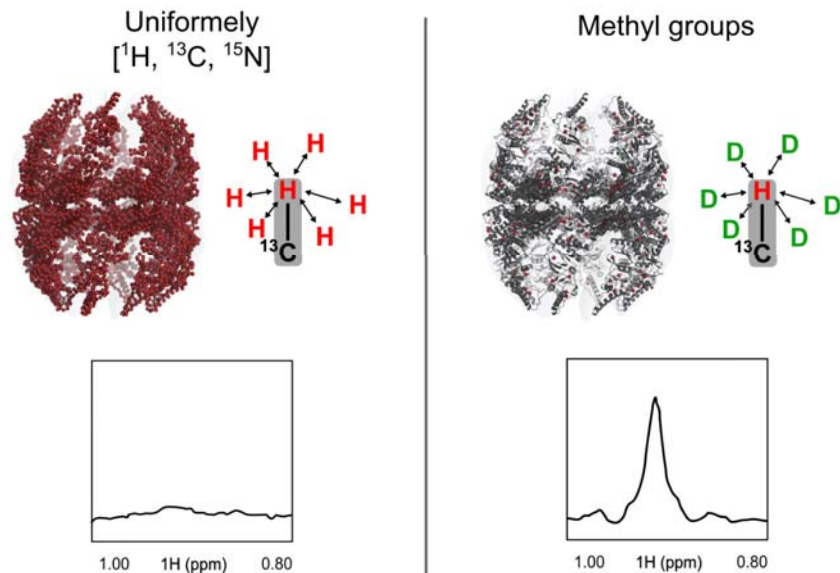


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Courtesy of J Boisbouvier et al.

Solution NMR Spectroscopy

Specific protonation of methyl groups

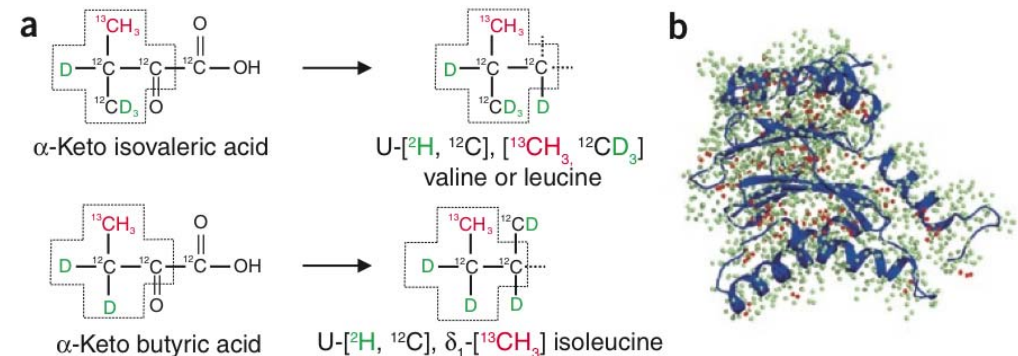


Gardner *et al*, J. Am. Chem. Soc. (1997)

Solution NMR of supramolecular complexes: providing new insights into function

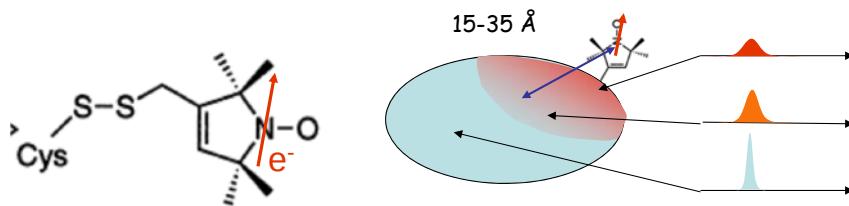
Remco Sprangers, Algirdas Velyvis & Lewis E Kay

Nature Methods 2007



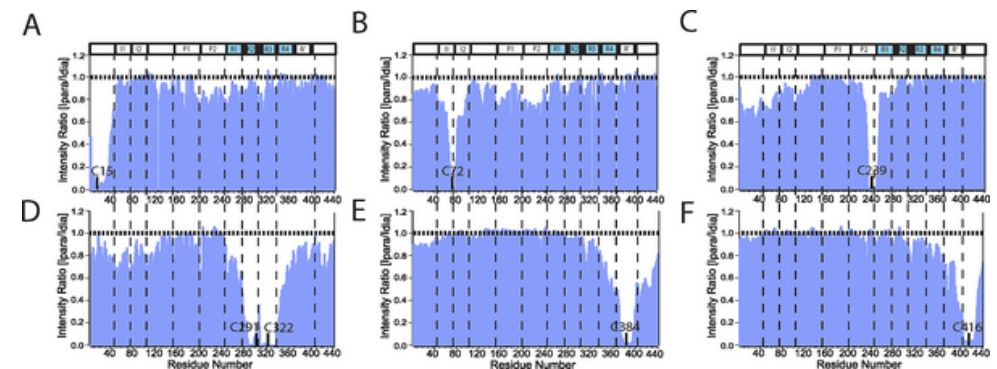
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Introducing spin label (PRE)



- The electronic spin induces an efficient relaxation of nuclear spins at long range distances
- This effect can be used to get structural restraints in large complexes
- This method prove to be invaluable to detect transient molecular events (encounter complexes, excited states, protin sliding on DNA,...)

Application to disordered proteins (Tau)

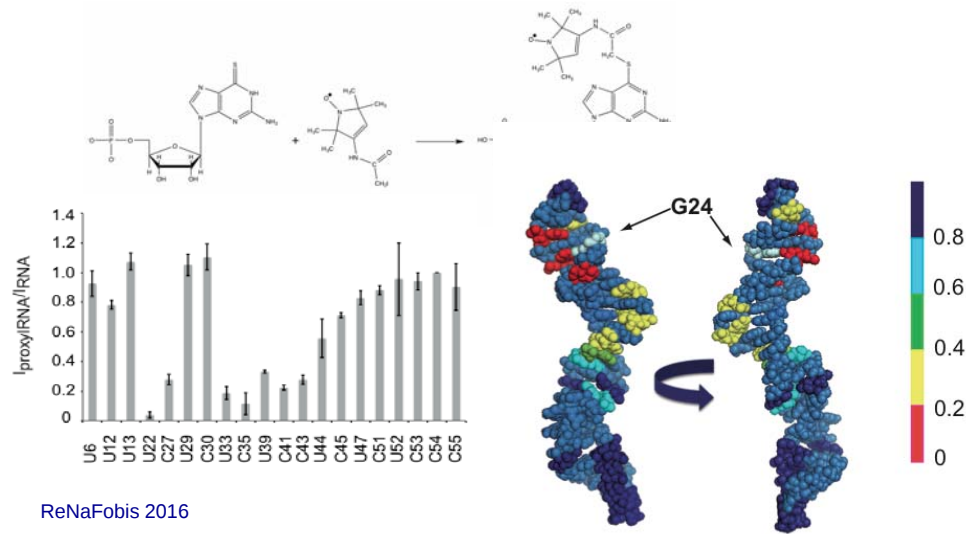


Mukrasch MD, Bibow S, Korukottu J, Jegannathan S, et al. (2009) Structural Polymorphism of 441-Residue Tau at Single Residue Resolution. PLoS Biol 7(2): e1000034. doi:10.1371/journal.pbio.1000034
<http://www.plosbiology.org/article/info:doi/10.1371/journal.pbio.1000034>

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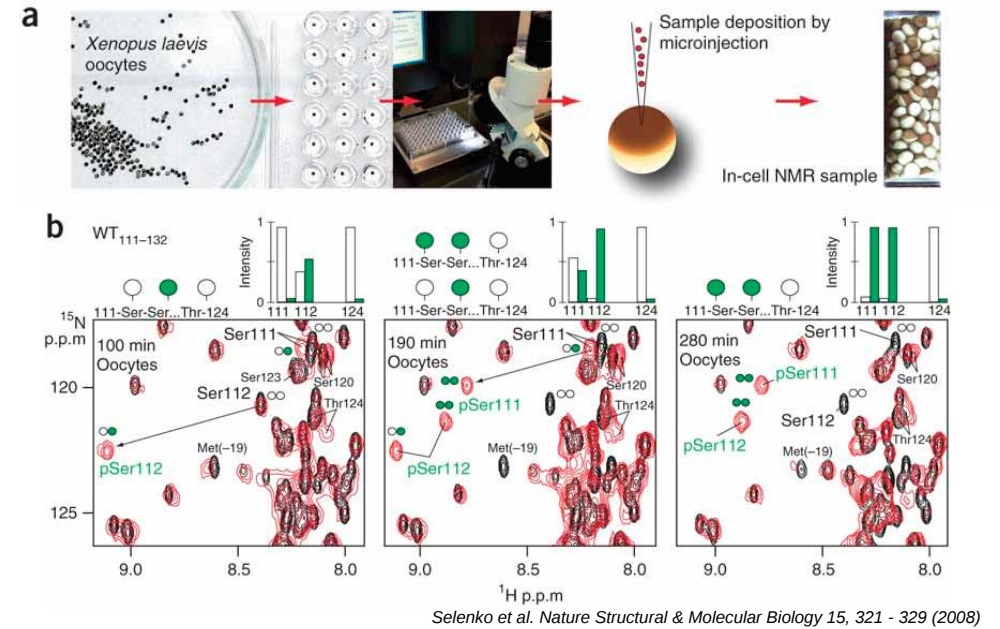
RNA spin labelling

Lebars I.*, Vilen B., Bourbigot S., Turek P., Wolff P. & Kieffer B. (2014)
 "A fully enzymatic method for site-directed spin-labeling of long RNA", *Nucleic Acids Res.*, 42(15), e117,



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In-cell NMR



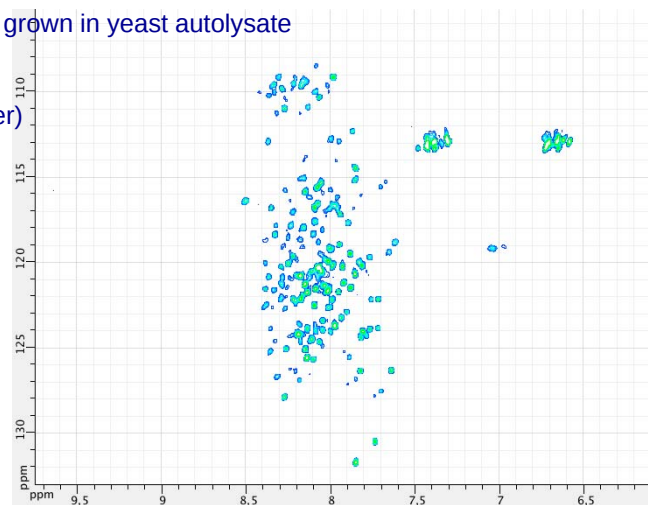
Selenko et al. *Nature Structural & Molecular Biology* 15, 321 - 329 (2008)

Expression of ^{13}C , ^{15}N labelling proteins in eukaryotic cells

Full length RXR nuclear receptor uniformly labelled with ^{15}N
 The protein forms an homodimer with 100 kDa molecular weight.

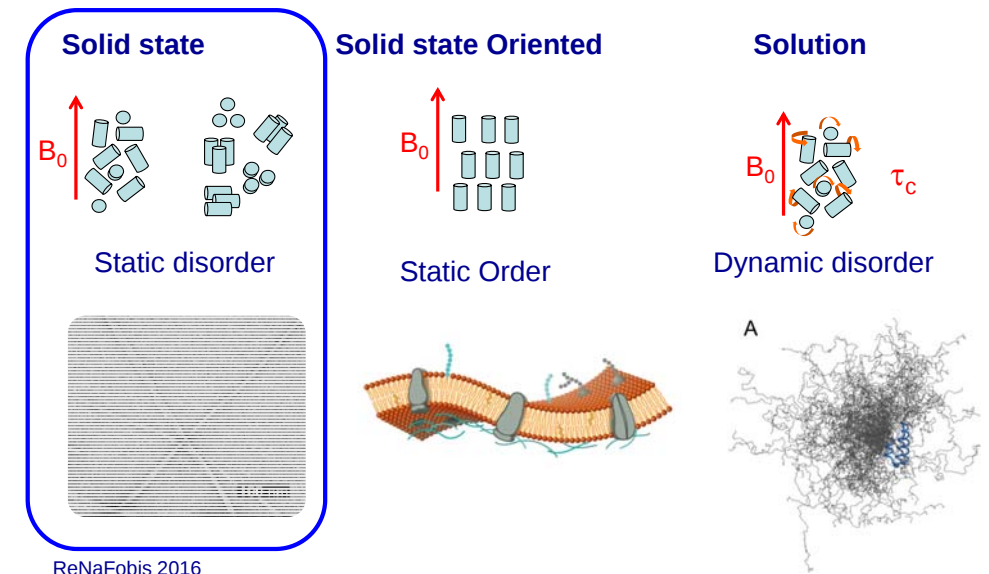
Expression in S2 insect cells grown in yeast autolysate

➤ Coll. François Bontems
 (Pasteur, ICSN, BioSpinger)



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Three States of living matter



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