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# **Traitement d'image et reconstruction tri-dimensionnelle**

**ReNaFoBis Ile d'Oléron 5th June 2014**

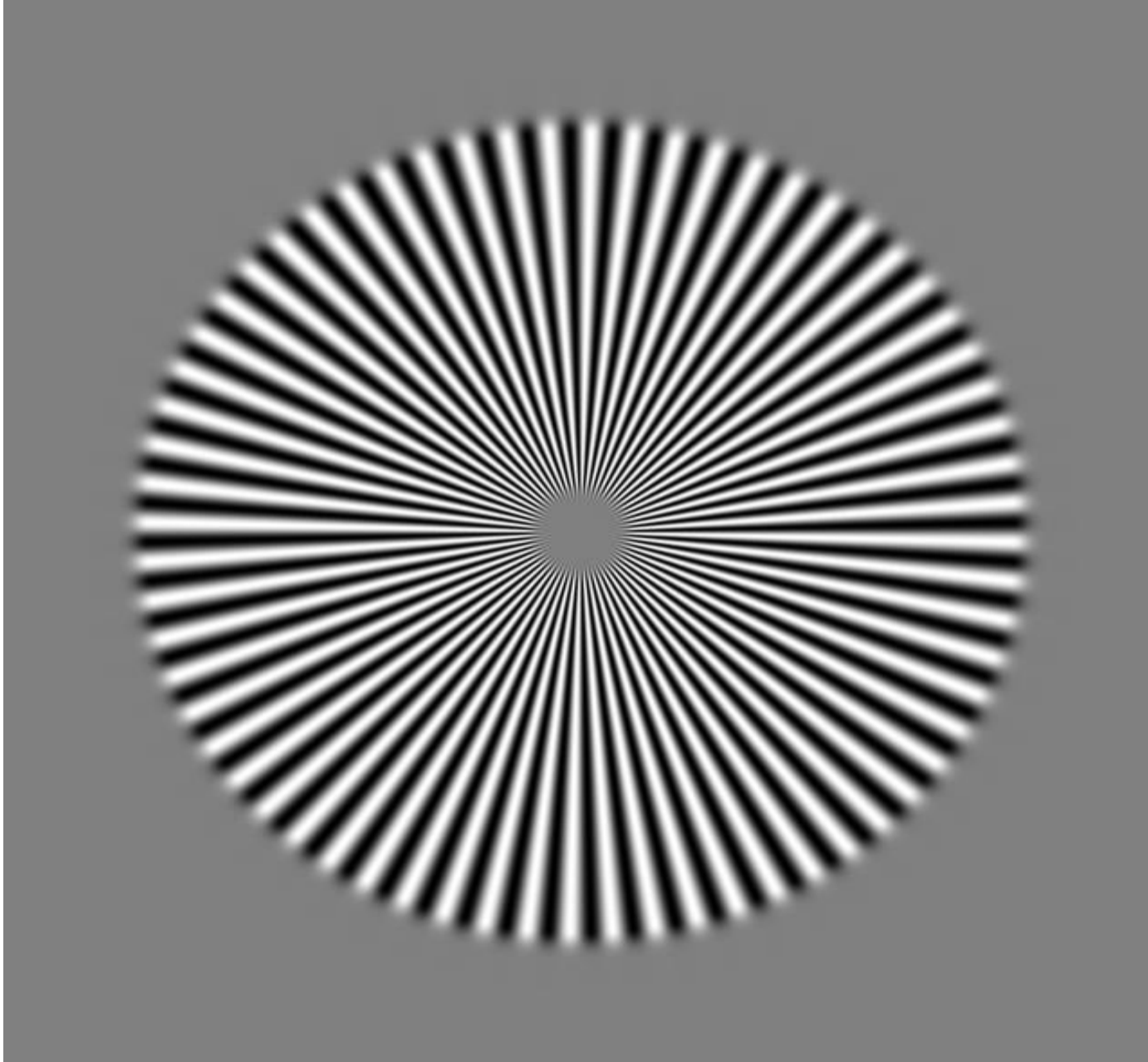
**Jean-François Ménéret**

**for Bruno Klaholz**

[\*\*http://www.igbmc.fr\*\*](http://www.igbmc.fr)

[\*\*http://igbmc.fr/Klaholz\*\*](http://igbmc.fr/Klaholz)

# Siemens stars: a whole range of spacings / frequencies in a single image

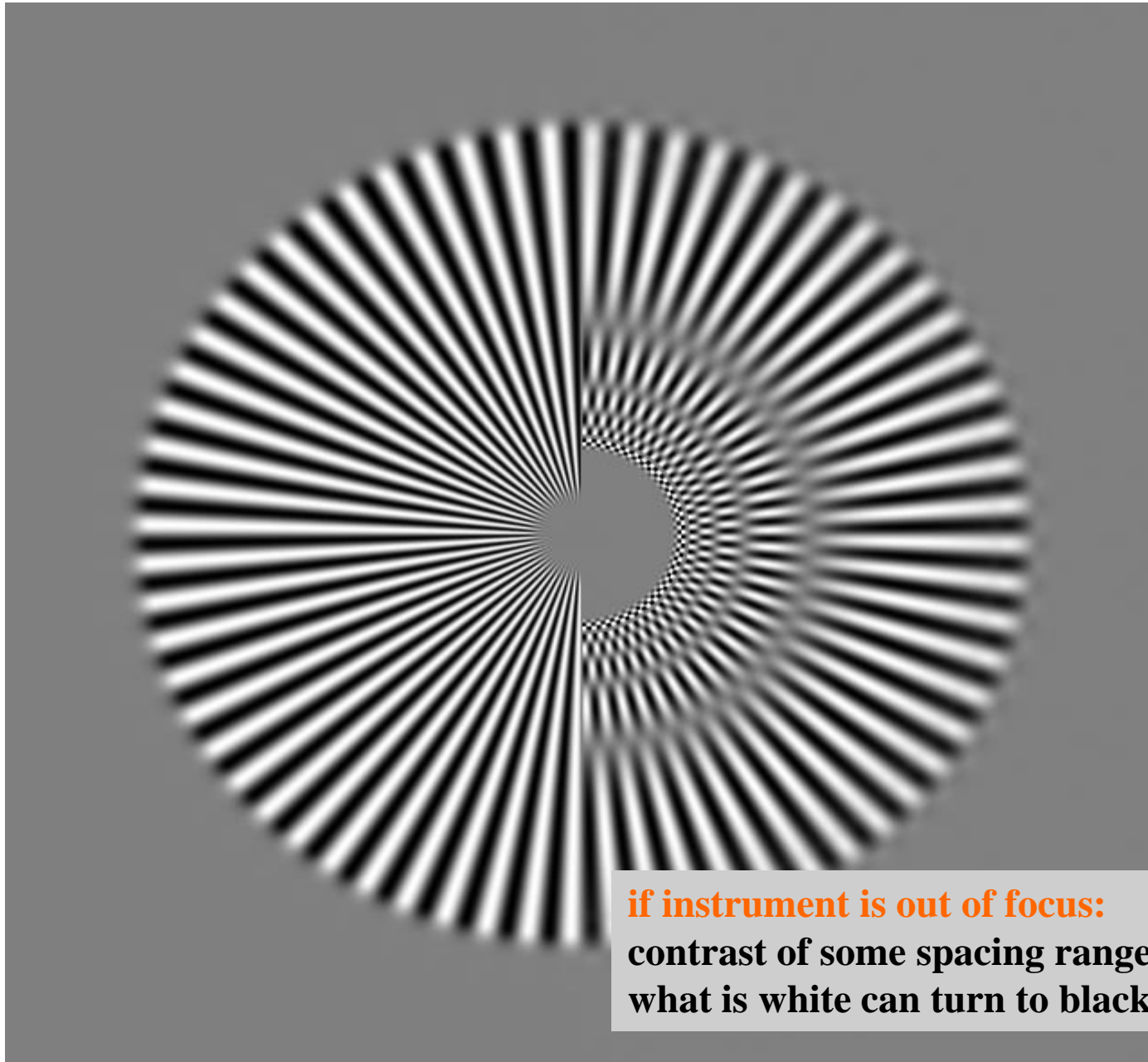


**One wedge  
every 1.5  
degree**

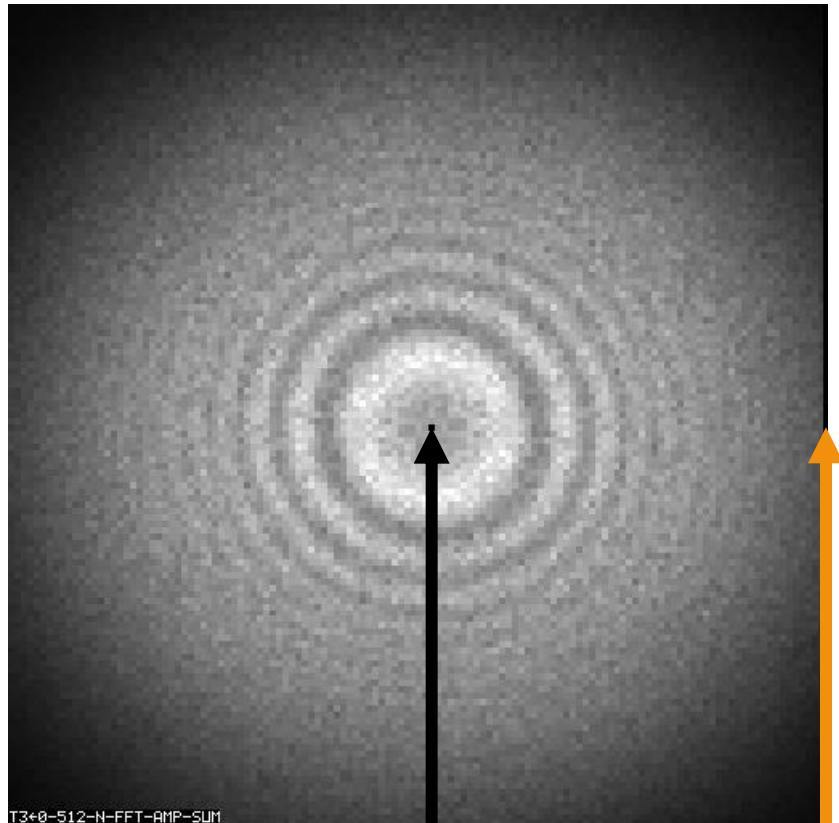
**It is used to test  
the resolution  
of optical  
instruments.**

*drawn by  
M. van Heel*

# Calculated effect of an electron microscopical PhCTF on the image of a Siemens star



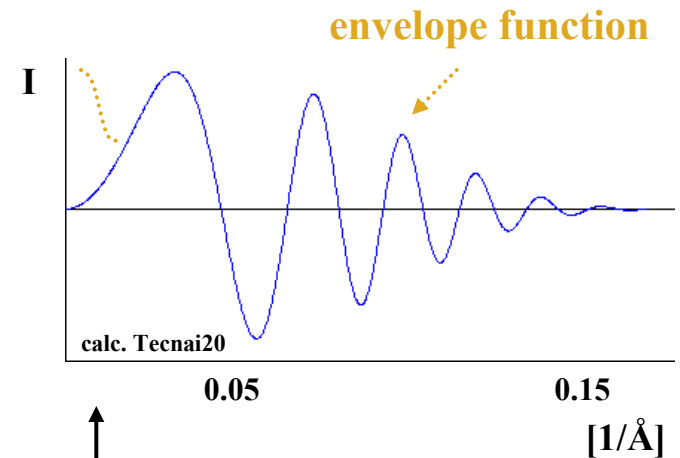
# Powerspectrum = Fourier Transformation of the image



low resolution  
low frequency

high resolution  
high frequency

## Profile of the intensity distribution



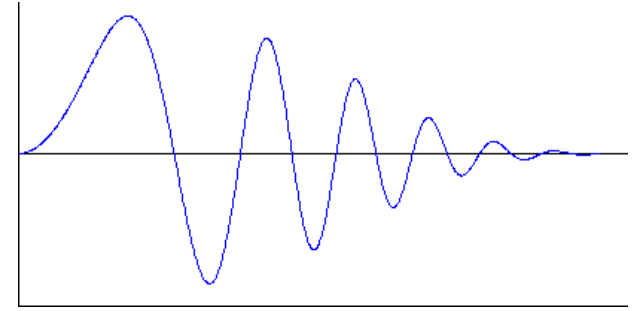
very low resolution difficult to measure

→ averaging techniques

# I. Pre-processing

## - correction of the Contrast Transfer Function

Weak-phase biological specimen → weak-contrast image



$$CTF(f) = A(\sin(\pi\lambda f^2(\Delta z - 0.5\lambda^2 f^2 c_s))) + B \cos(\pi\lambda f^2(\Delta z - 0.5\lambda^2 f^2 c_s))$$

**Phase contrast                      and                      amplitude contrast**

$\lambda$  wavelength defined by accelerating voltage (e.g. high tension 200 kV -->  $\lambda = 0.025 \text{ \AA}$ )

$C_s$  spherical aberration coefficient, determines the quality of objective lens

$B$  fraction of amplitude contrast

$A$  defocus-dependent envelope function

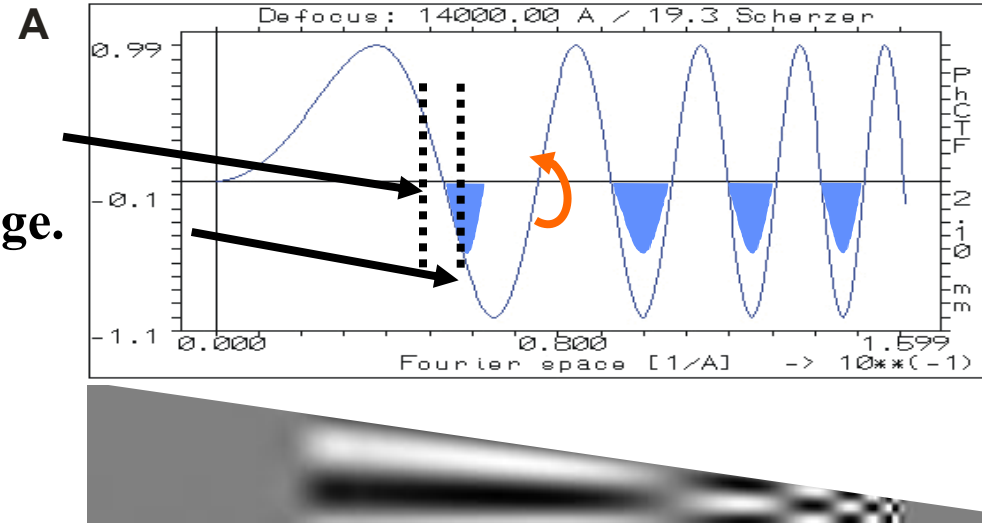
$\Delta z$ : the **defocus value**, (offset from focus, "Scherzer" focus, underfocus: **focal point is below the sample**)

$f$ , spatial frequency

# I. Pre-processing

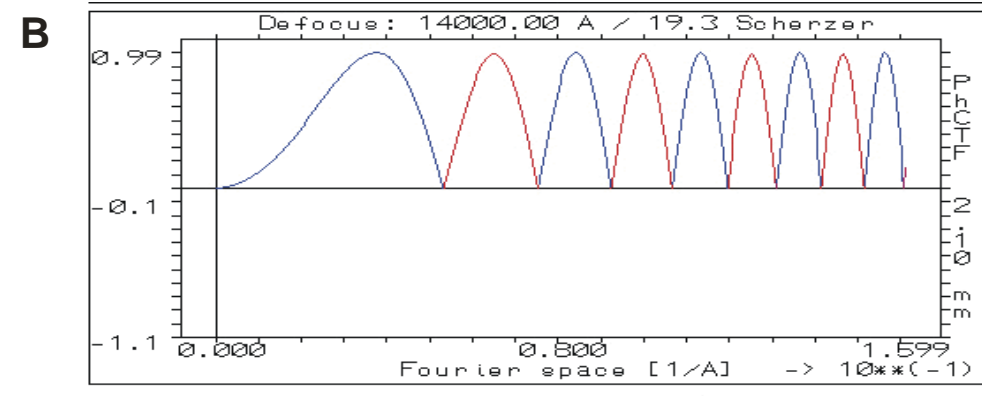
- correction of the contrast transfer function (CTF)

No information transfer at the zero crossing;  
CTF leads to **contrast inversion** within the image.



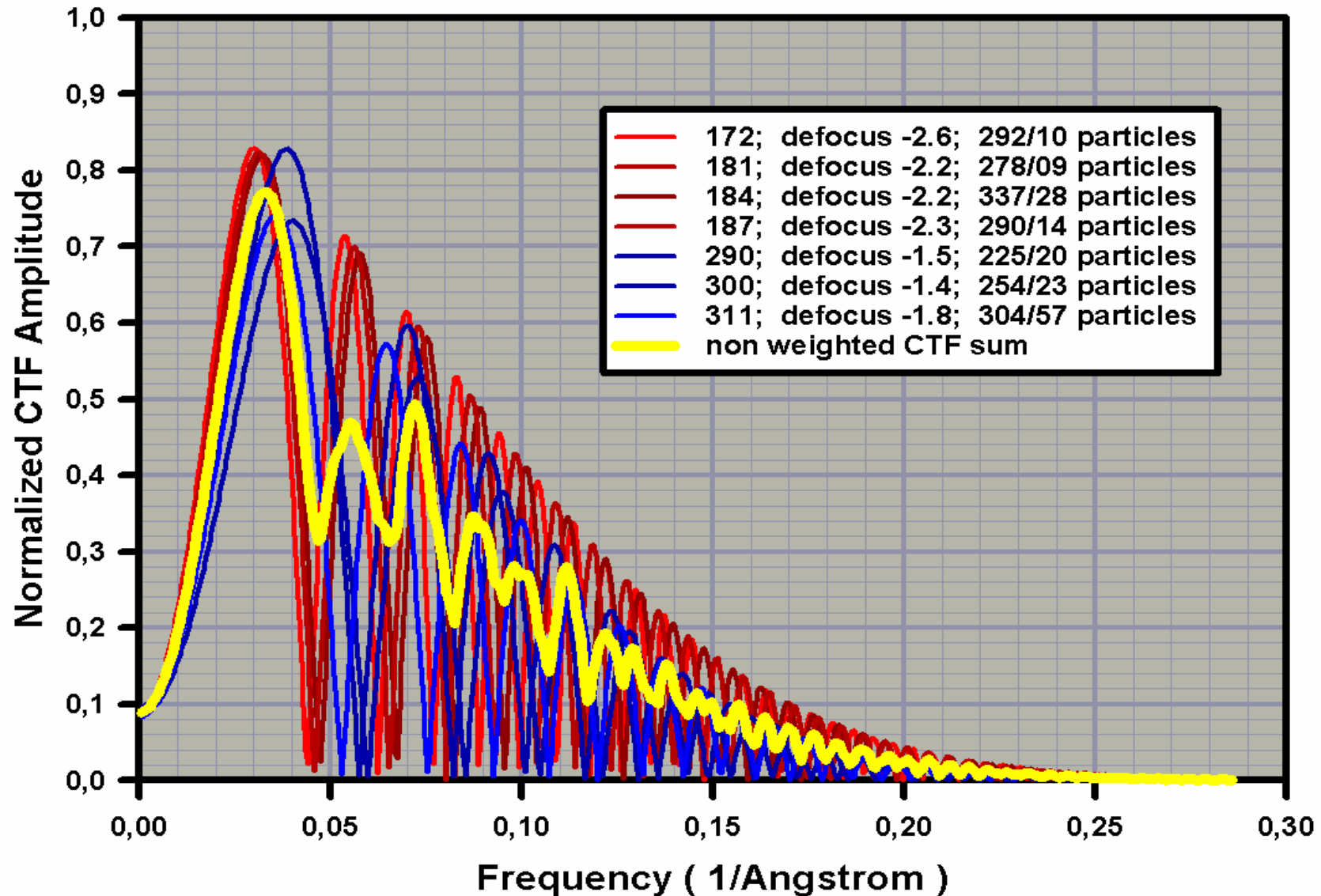
Radial segment of Siemens star

CTF-correction? By "phase flipping":



Therefore: data collection over range of defocus values,  
e.g. under-focus -1.0 – 3.0  $\mu\text{m}$

# Combination of powerspectra from different defocus images



*(spectra also take the envelope function into account)*

### **In EMAN2 on Virtual Machine**

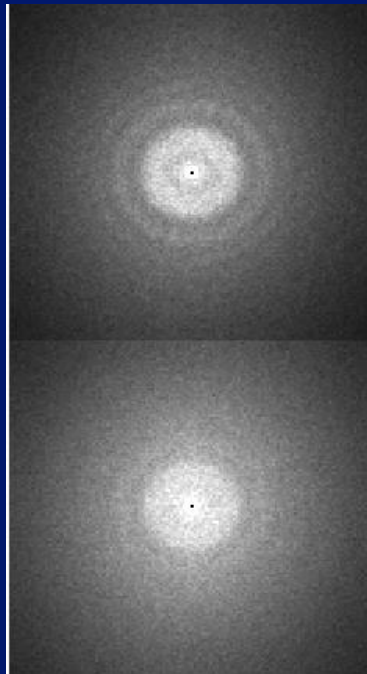
- 1) display images**
- 2) calculate their FFT**
- 3) to see the CTF + noise + structure factor**

### **In EMAN1 on Virtual Machine**

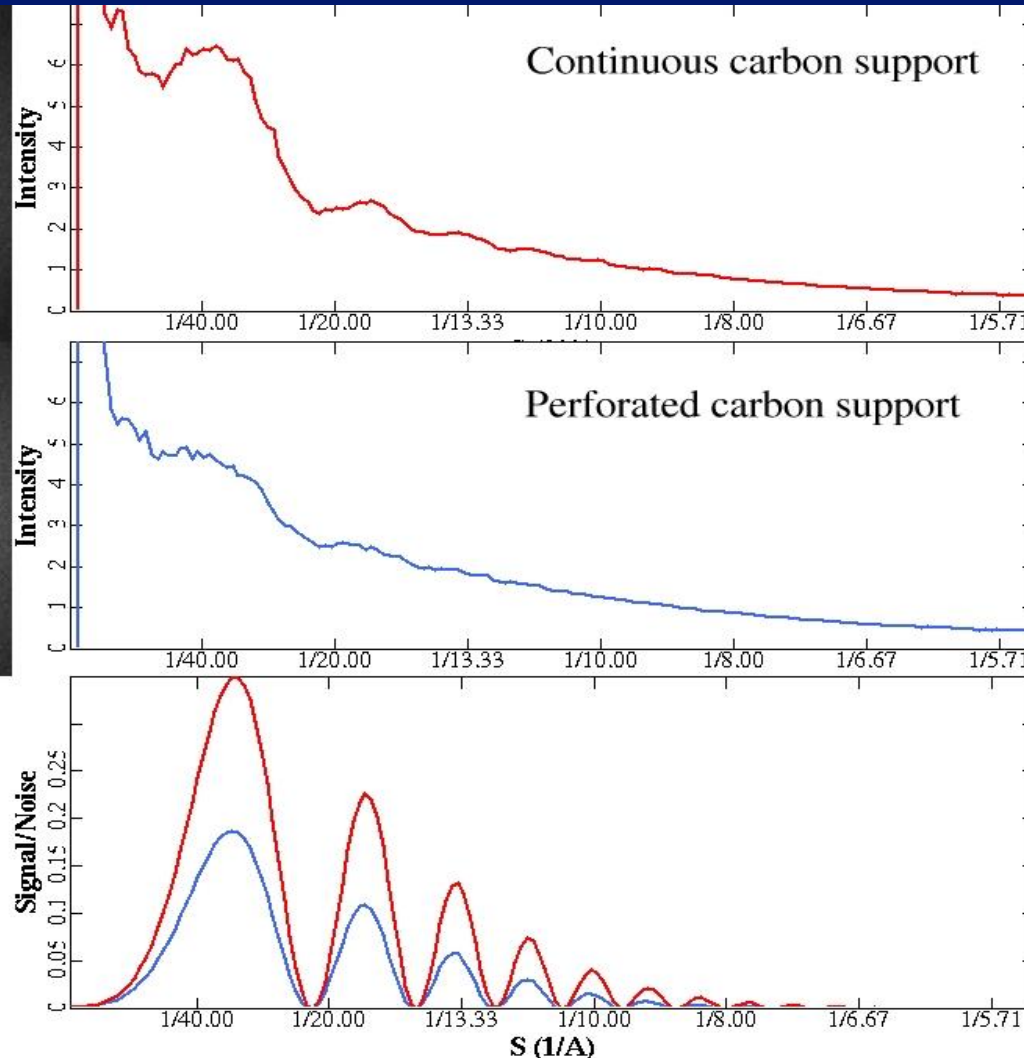
- 1) simulate the CTF, envelope function, Nyquist limit**
- 2) add several CTF curves**
- 3) See complementarity between curves at different defocus values  $\Delta f$**



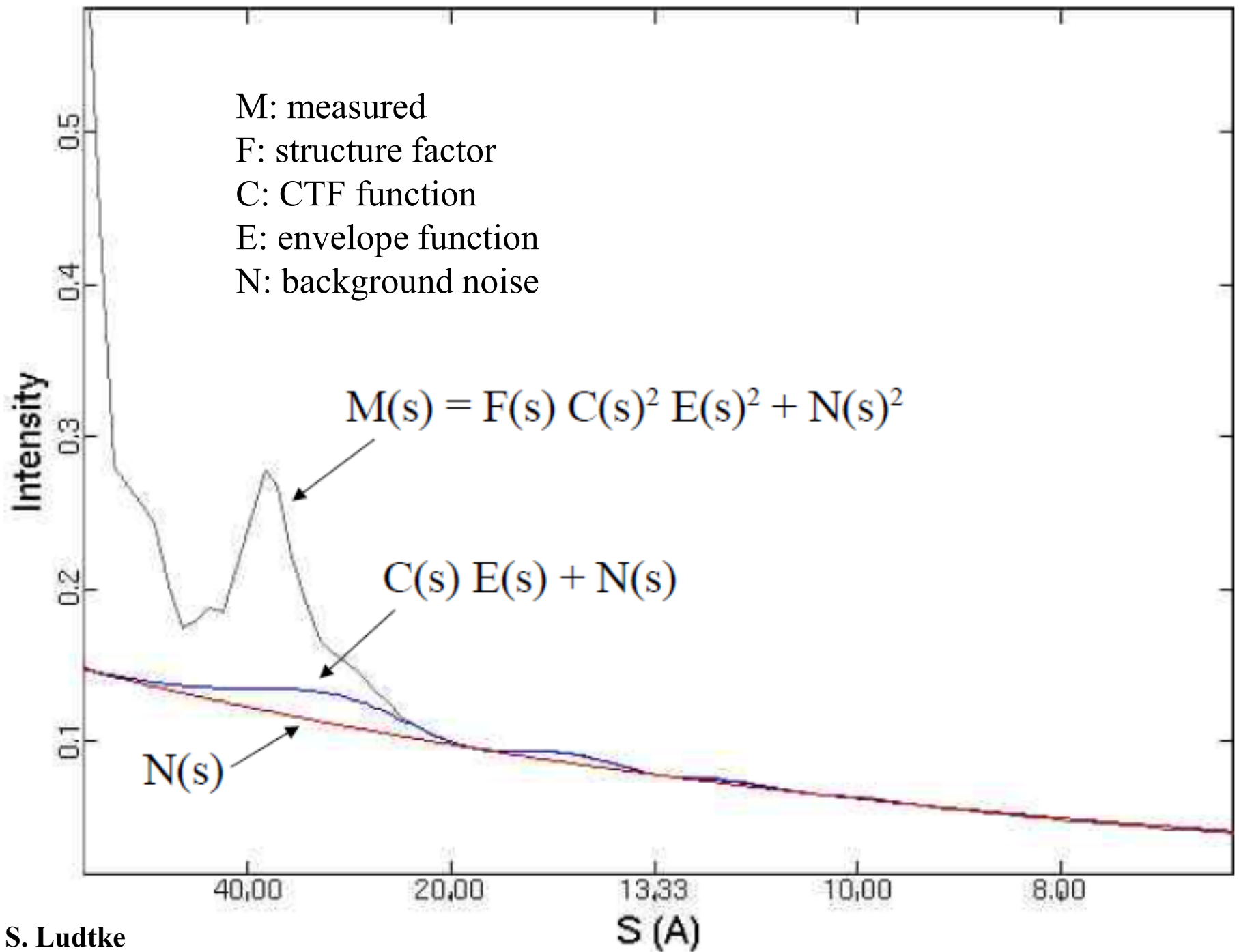
# Cryo-EM; Contrast Transfer Function

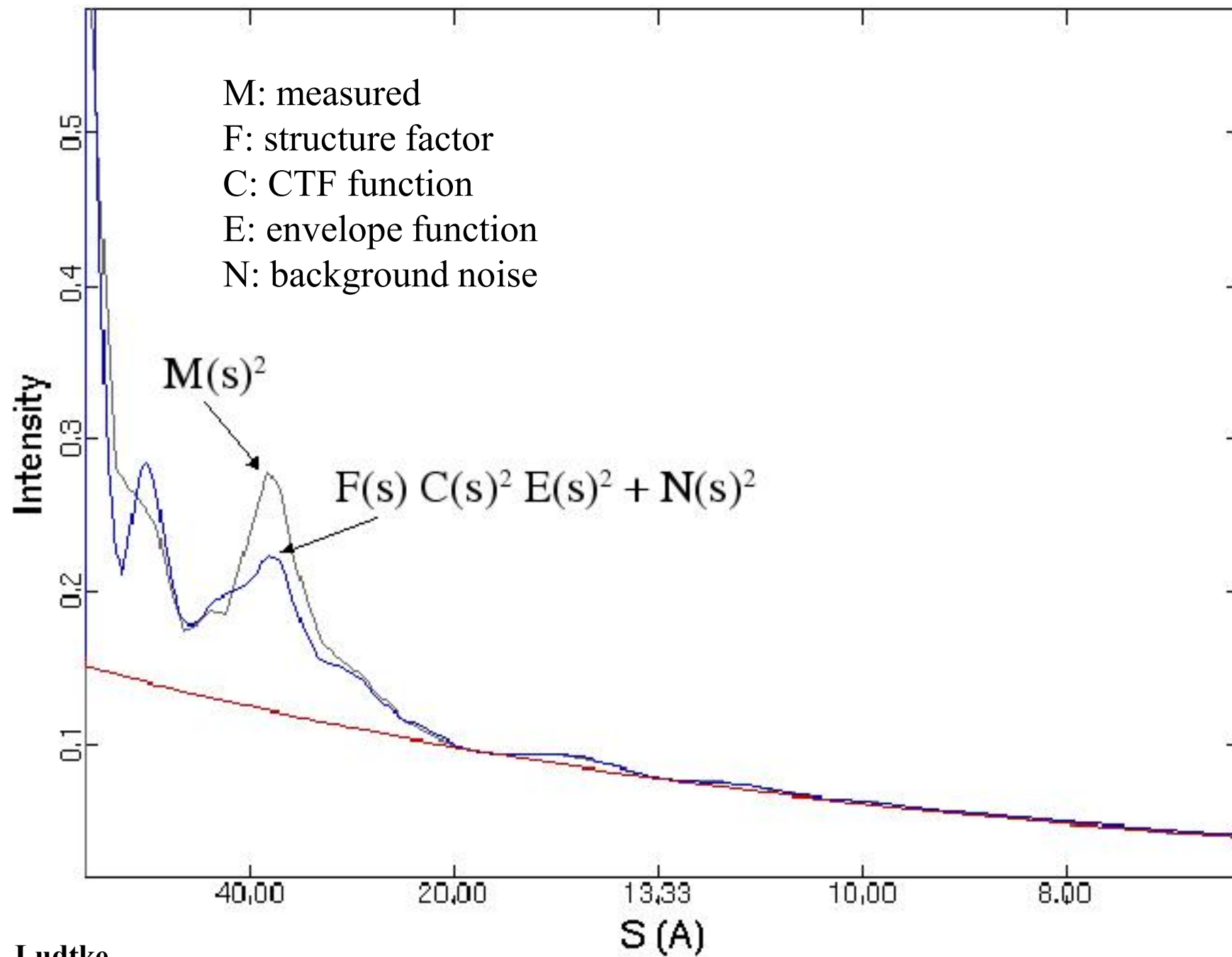


100  
particles  
in each case



Curves include:  
CTF,  
background noise,  
structure factor,  
envelope function



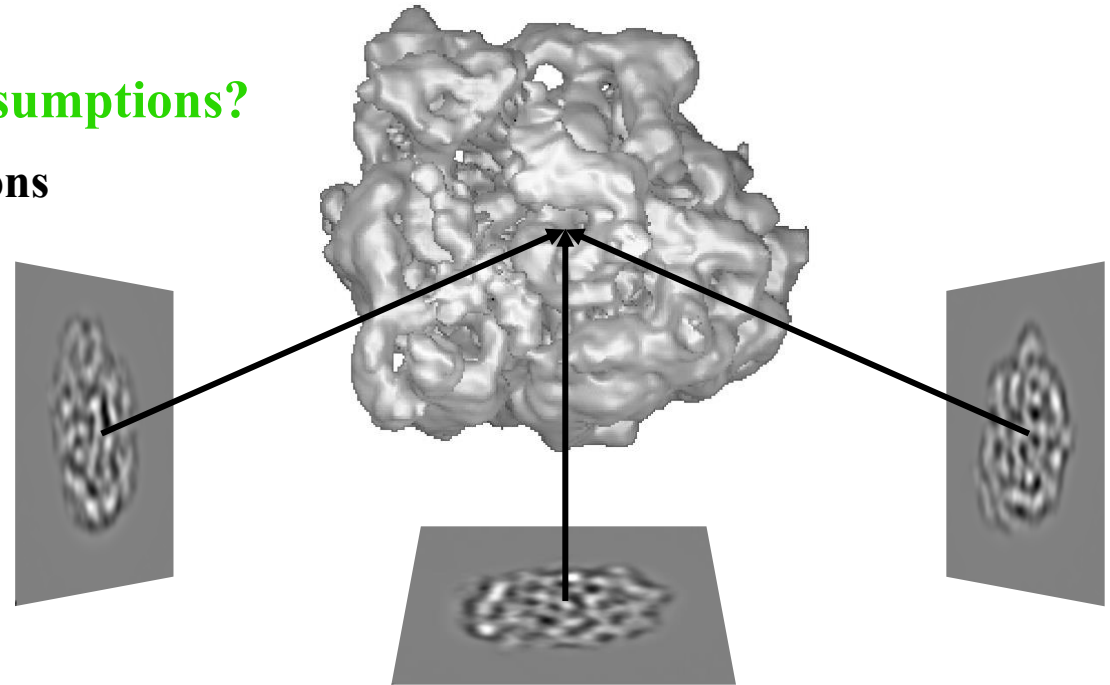
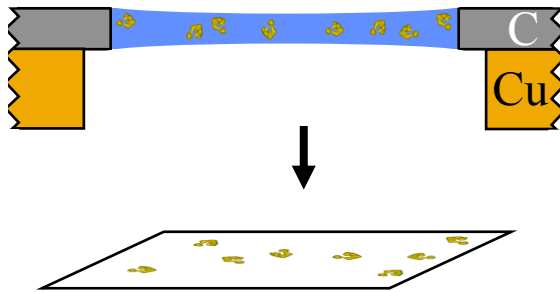


## II. Structure determination

- angle assignment
  - angular reconstitution (in early stage of structure determination)
  - projection matching (if structure already well refined): find best correlation between input image and reference images from 3D re-projections)

3D reconstruction of single particles: **assumptions?**

**unique** particle type in **random** orientations



## *Real space*

## *Fourier space*

*map*

3d density distribution

3D FFT

3D Fourier transform

back-project  $\uparrow$   $\downarrow$  project

$\uparrow$   $\downarrow$  extract central section  
insert

*class average*

2D projection

2D FFT

2D central section

back-project  $\uparrow$   $\downarrow$  project to line

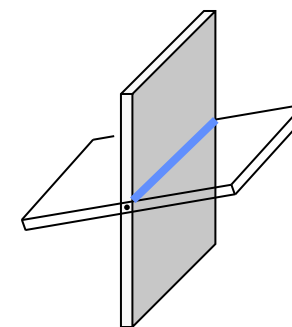
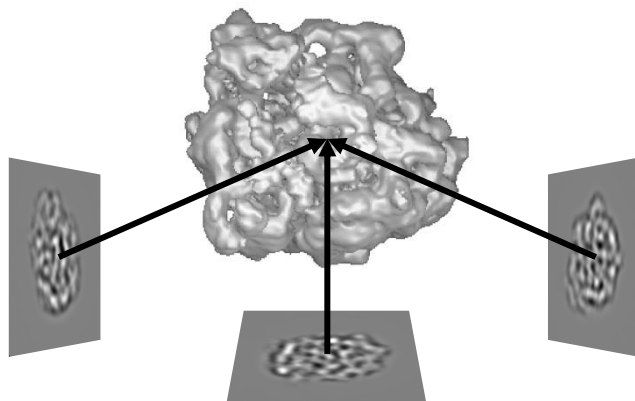
$\uparrow$   $\downarrow$  extract central line  
insert

*sinogram line*

1D projection

1D FFT

1D central line



*common line*

common line projections theorem

## II. Structure determination

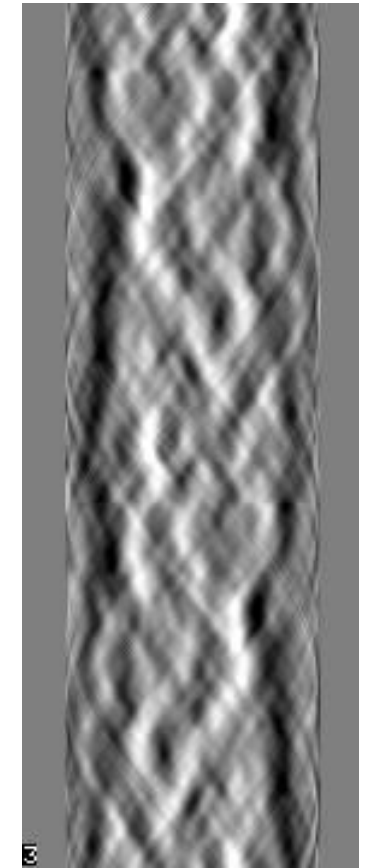
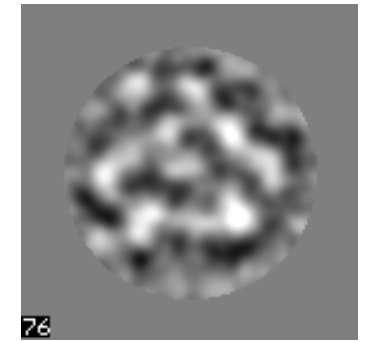
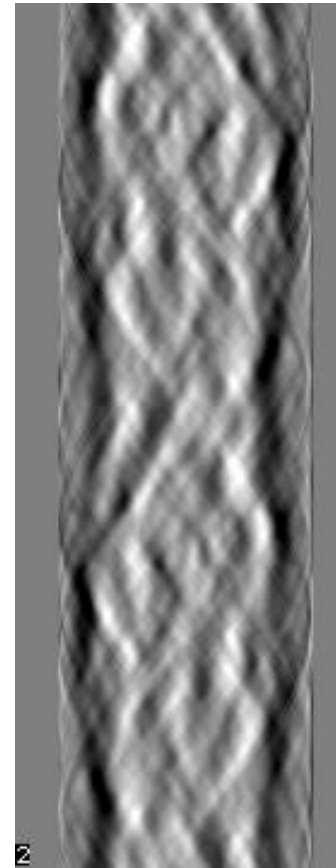
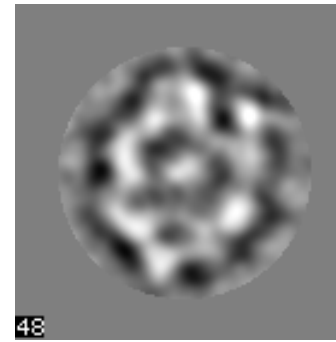
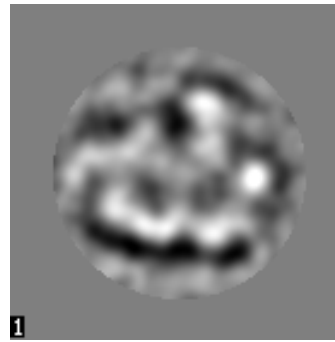
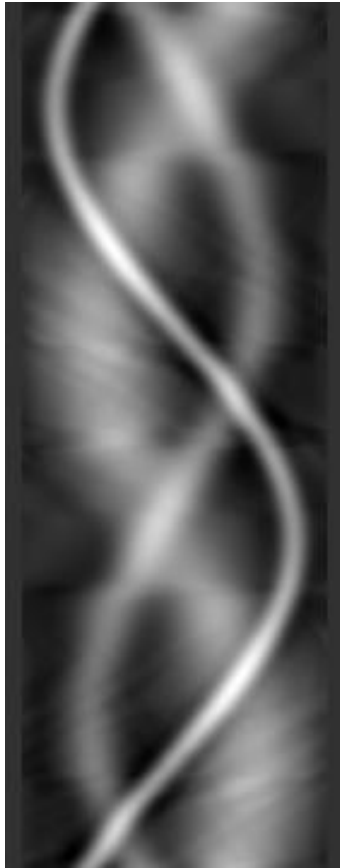
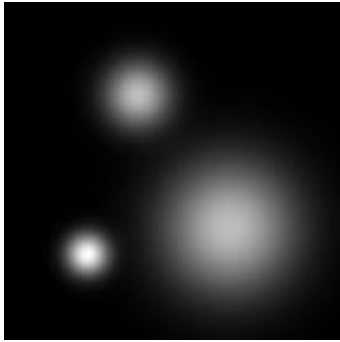
- angle assignment
- angular reconstitution



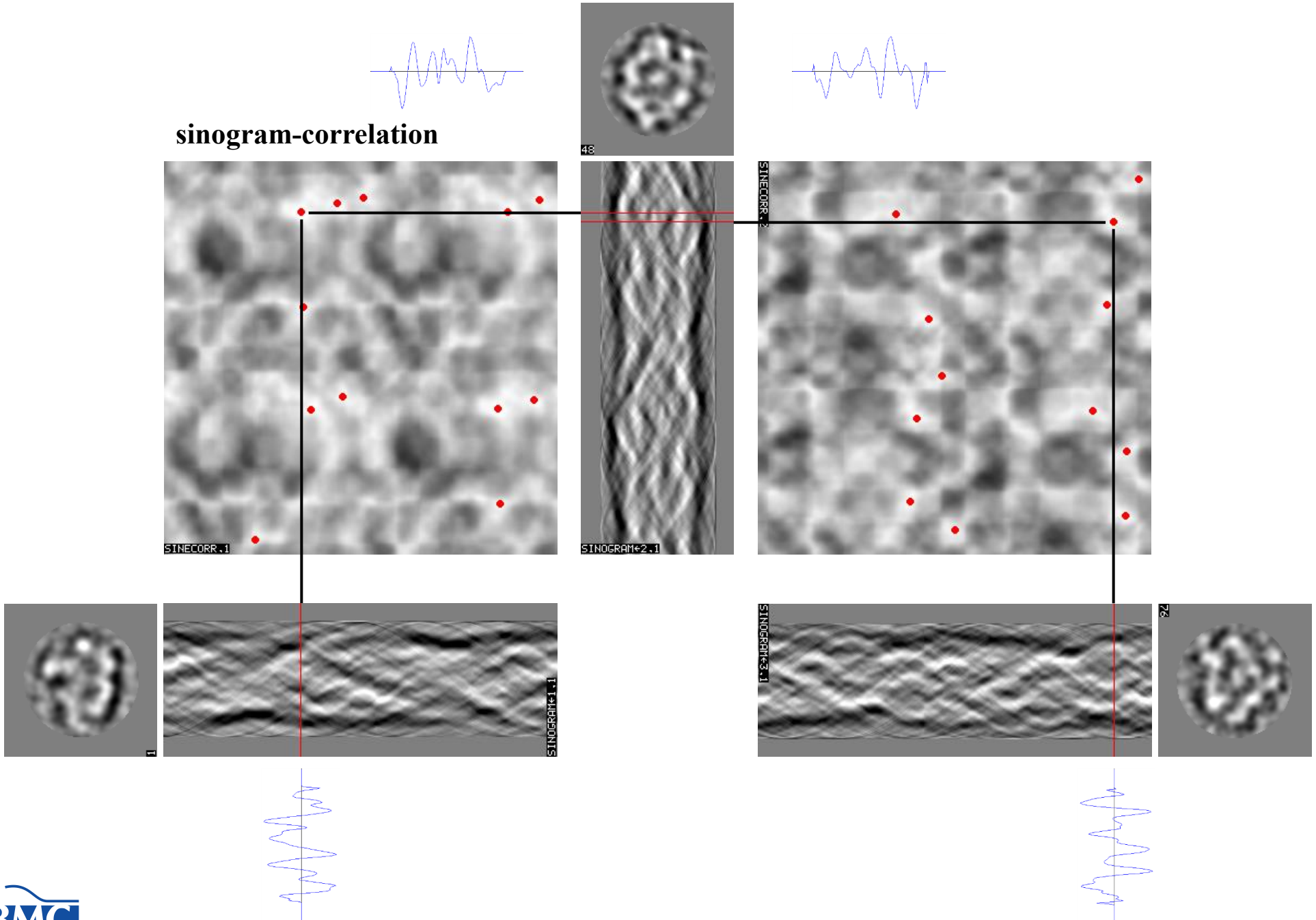
**sinogram** = **line-projection** of the 2D image  
(also called Radon transform)

*amplitude-square-root filtered*

Select 3 clearly different views (here: class average numbers 1,48,76):



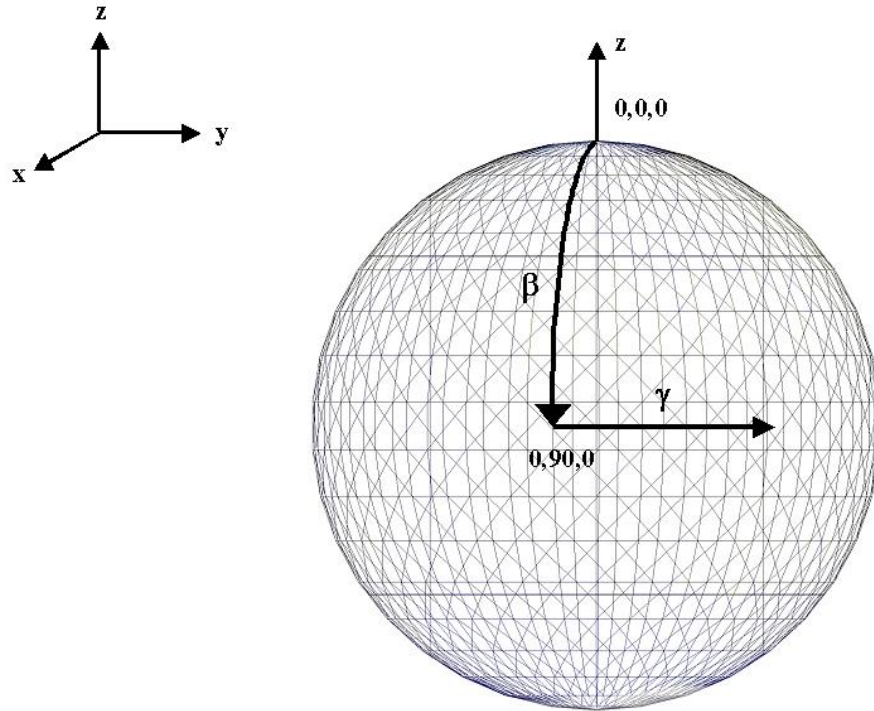
# sinogram-correlation



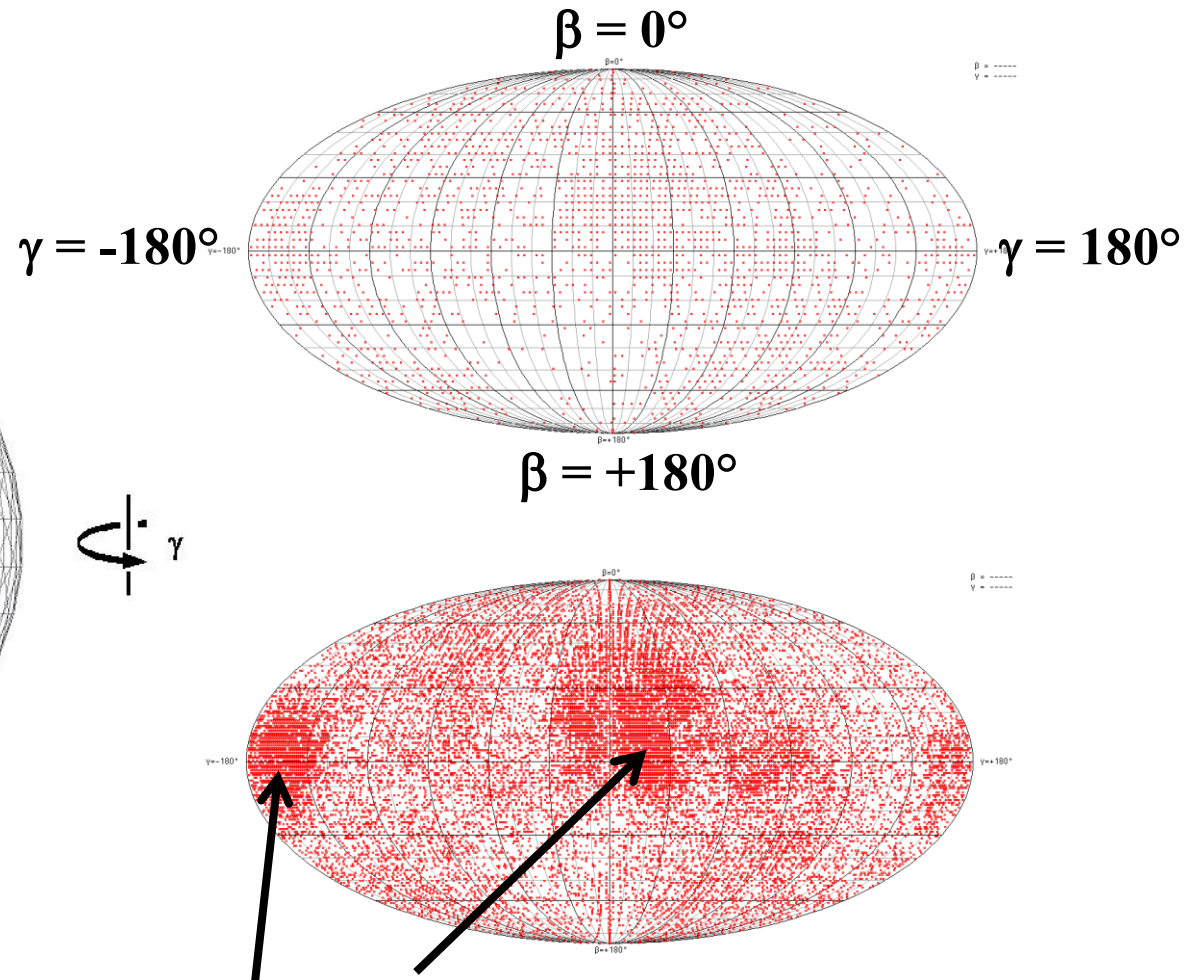


## II. Structure determination

- angle assignment
- angular reconstitution



Particle angles plotted on sphere:



Preferential views

## II. Structure determination

- angle assignment
  - angular reconstitution

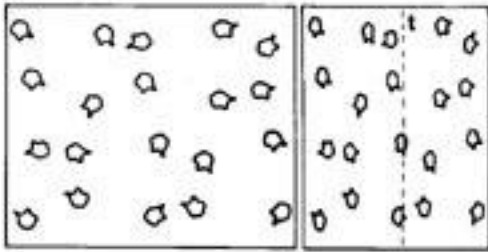
In case of *ab initio* structure determination by  
reference-free alignment and angular reconstitution:

Does not allow to determine **handedness**, requires either:

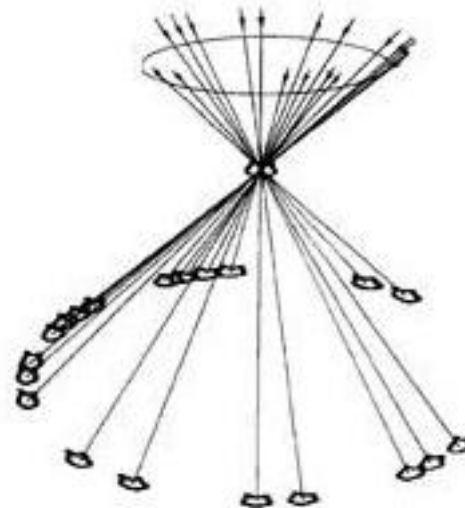
- random conical tilt (Radermacher *et al.*, J. Microsc. 1987)
- tomography
- phase residual error using a tilt pair (Rosenthal & Henderson, JMB 2003)
- fitting of crystal structures

## II. Structure determination

### Random Conical Tilt method (Radermacher 1987)



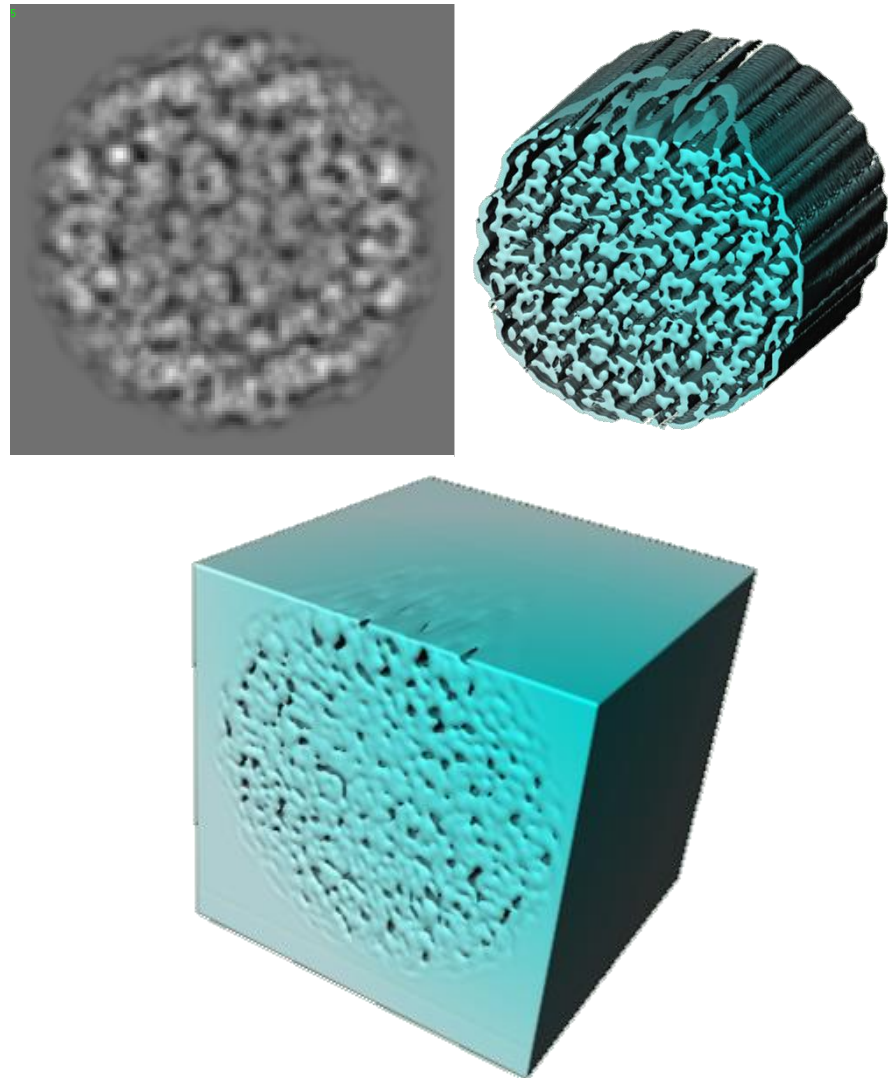
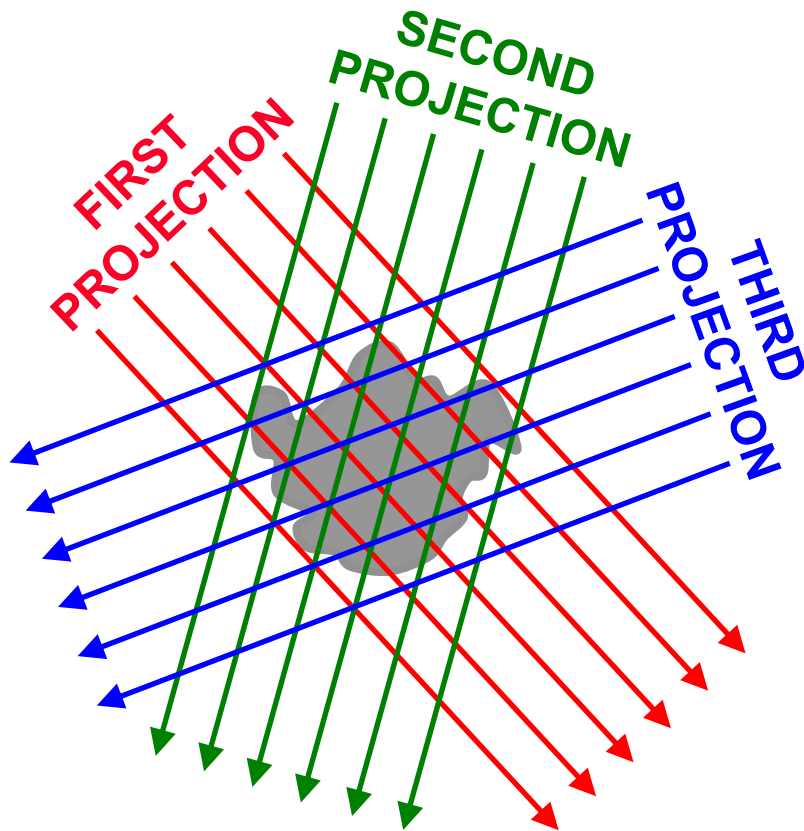
**Tilt pair (45deg/0deg); tilt axis**



**THE “cone”**

## II. Structure determination

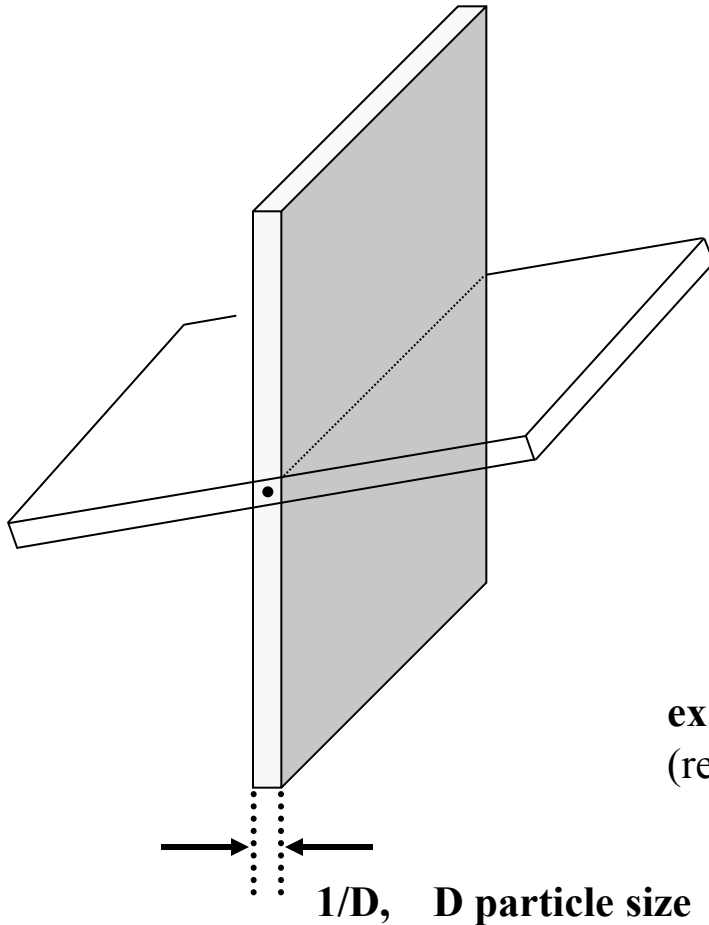
### - 3D reconstruction



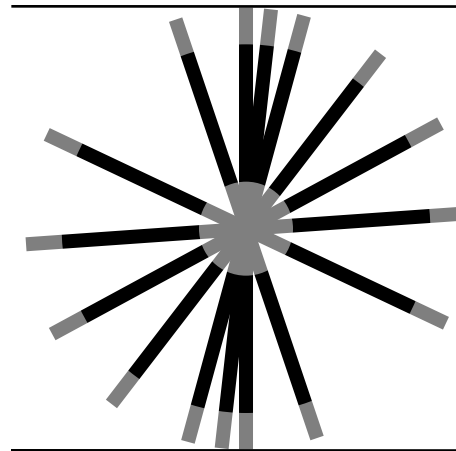
*drawn by  
I. Orlov*

## II. Structure determination

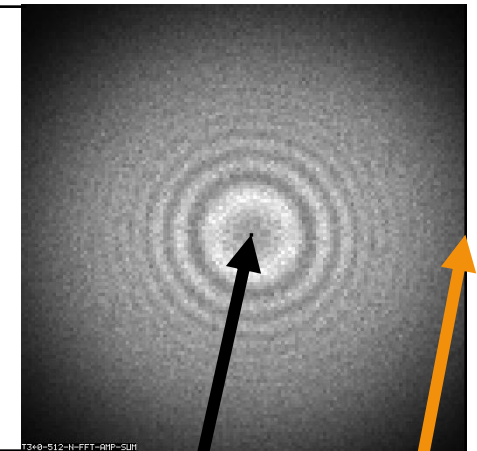
### - 3D reconstruction



exact, weighted filtered back-projection  
(represented 2D for simplification)



Reminder: powerspectrum:

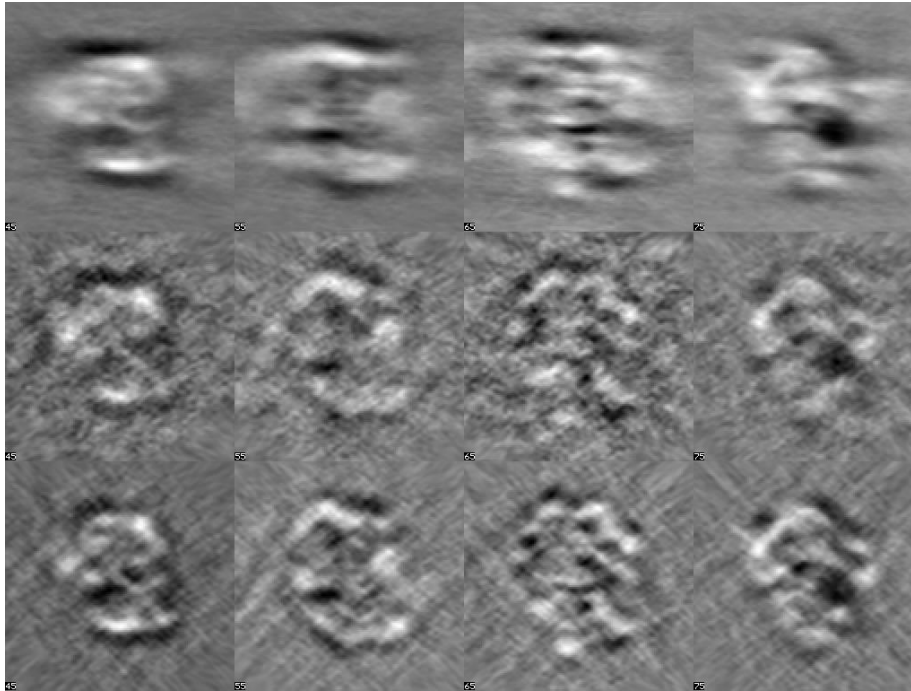


low resolution low frequency **high resolution high frequency**

*(representations of central sections in Fourier space)*

## II. Structure determination

### - 3D reconstruction

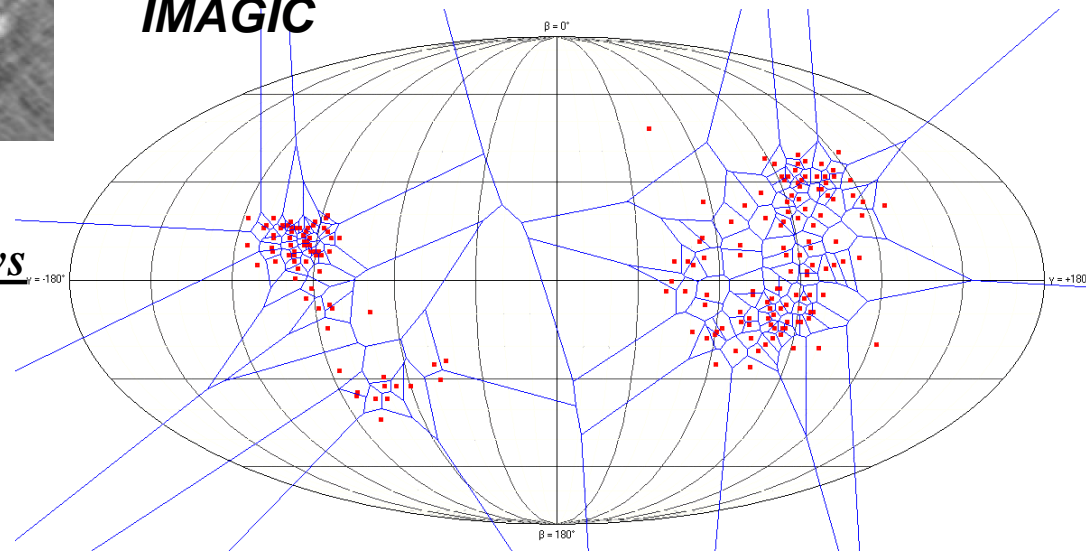


*non-weighted BACK-PROJECTION*

*Weighted BACK-PROJECTION*  
~~waited~~

*sections of 3D's calculated from 250  
class averages with strong preferential views*

**IMAGIC**

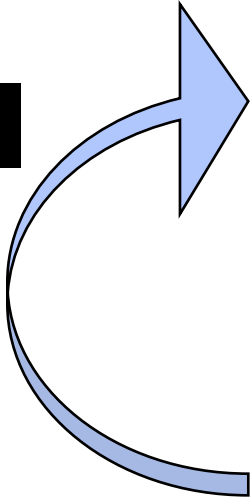


Orlov *et al.*, in prep. (2014).

## II. Structure determination

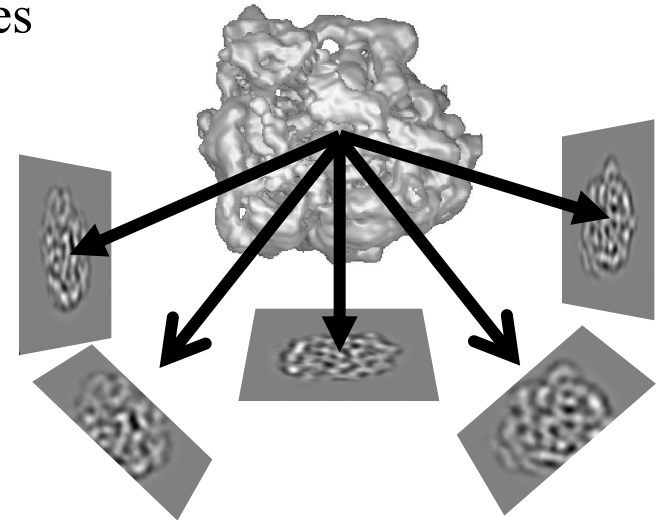
### - structure refinement

**iterations**



- centering/alignment
- variance analysis + classification
- angle assignment
- angular reconstitution → 3d-reconstruction
- reprojections = new references

- improve quality of angle assignment
- improve quality of particle alignment

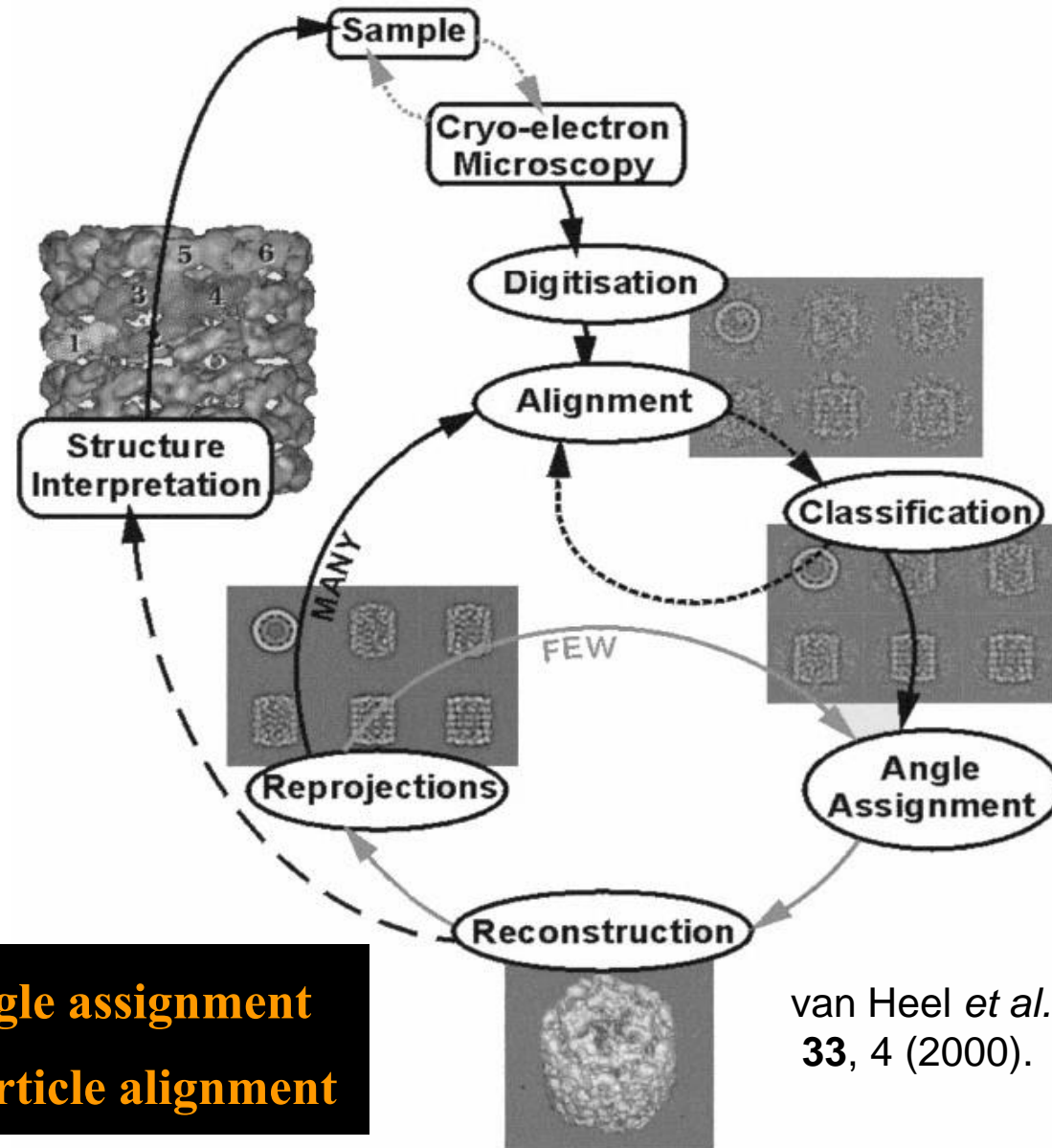


**equally distributed  
forward-projections  
(re-projections)**



## II. Structure determination

### - overview



van Heel *et al.*, Quart. Rev. Biophys.  
33, 4 (2000).

- improve quality of angle assignment
- improve quality of particle alignment



## II. Structure determination

### - resolution assessment

Particle data set

split into 2 halves



3D

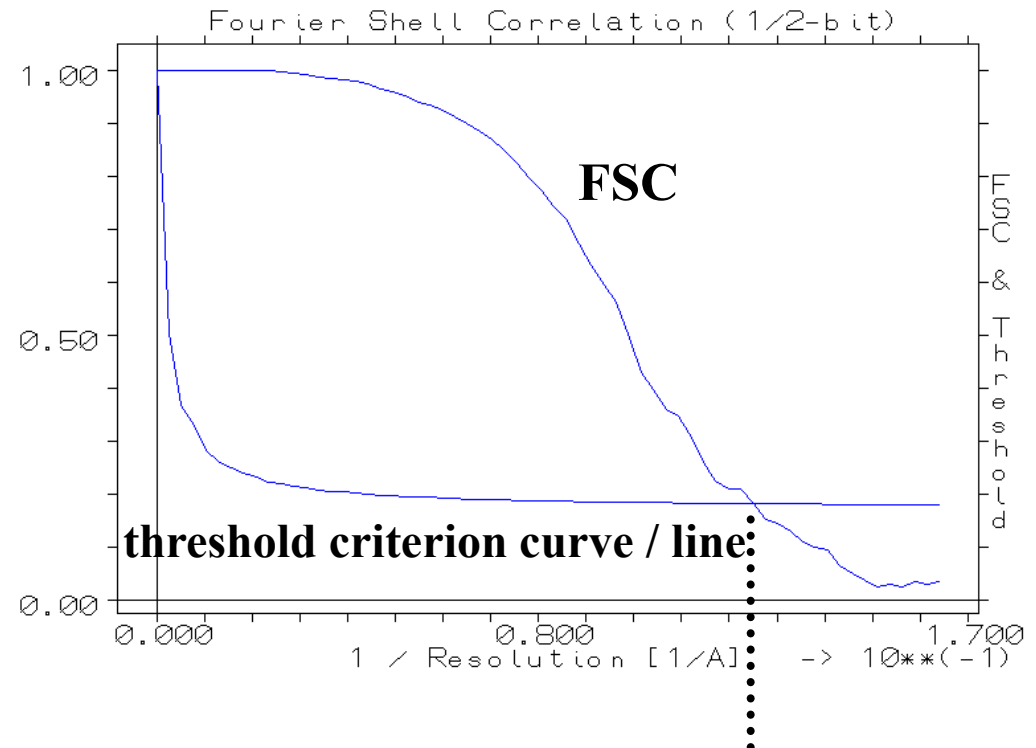


3D

calculate cross-correlation

by shells in Fourier space

### Fourier Shell Correlation (FSC)



$$1 / 1.25 \times 1 / 10 \text{ Å} = 8 \text{ Å}$$

**Keep in mind: resolution is what you can resolve in the 3D map!**

## II. Structure determination

- map interpretation ; fitting of crystal or NMR structures

**Fitting procedures:**

- manual fitting (e.g. O, A. Jones, Acta Cryst. (1991))
- real space fitting
- reciprocal space fitting

**1) global search**

**2) refinement**

e.g. torsion-angle molecular dynamics

- fit complete structures, domains, factors;

**Usually backbone is enough.**

**rigid body or flexible fitting**

- use full maps or difference maps

**Be careful with local minima and over-fitting!**

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Cryo Electron Microscopy

**A Titan Krios cryo electron microscope** will be installed during 2013. This is the latest generation electron microscope with capacities for high-resolution data collection for both Cryo Electron Tomography (CET) experiments and Single Particle Analysis (SPA) with automated data collection. The electron source is a Field Emission Gun (FEG) that can be operated at 80keV, 100keV, 200keV or 300keV. An automatic loading mechanism allows mounting twelve grids at a time. This microscope is equipped with a CMOS (FEI FALCON 4K\*4K) high-sensitivity direct electron detector camera.

**The Tecnai F30 Polara** cryo electron microscope allows high-resolution SPA and CET automated data collection. Its FEG is usually tuned to 100keV, 200keV or 300keV. The grids are manually mounted 6 at a time. This microscope is equipped with 3 digital cameras: FEI CMOS 4K\*4K "FALCON", FEI CCD 4K\*4K "EAGLE" and a GATAN CCD 2K\*2K "ORIUS".

**The Tecnai F20** is equipped with a FEG operated at 100keV or 200keV. Grids are mounted one at a time using a Gatan side-entry cryo-holder. This microscope is equipped with one digital camera (GATAN CCD 2K\*2K "US10001"), it is used to collect data for cryo-SPA or room temperature electron tomography using a Fischion side-entry holder.

Cryo-EM grids are flash-frozen in a temperature and humidity controlled environment using a Vitrobot system (FEI).

Although all our microscopes are equipped with digital cameras, we can still record images on films and scan them with a high-resolution Heidelberg Druckmaschinen drum scanner ( 5 micron pixelsize)

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Jean-Francois.Menetret@igbmc.fr

OTHER PLATFORMS

Sample preparation

- Bacterial expression
- Baculovirus expression
- Yeast expression
- Purification
- Mammalian expression

Biophysical characterisation

- Analytical Ultra Centrifugation
- Calorimetry
- Thermal Shift Assay

Windows Taskbar: Internet Explorer, File Explorer, VLC, Firefox, Chrome, PowerPoint, PDF Reader

System Tray: EN, Network, Volume, 21:43

**Thank you for your attention**