

# Tutorial

## I - MOLECULAR REPLACEMENT PHASING

Resolution of the structure of hen lysozyme from diffraction of Hen lysozyme crystal and a coordinates file of Bovine lysozyme :

### Hen lysozyme sequence :

```
>sp|P00698|LYSC_CHICK Lysozyme C OS=Gallus gallus GN=LYZ PE=1 SV=1  
MRSLLILVLCFLPLAALGKVFGRCLEAAAMKRHGLDNYRGYSLGNWVCAAKFESNFNTQA  
TNRNTDGSTDYGILOQINSRWWCNDGRTPGSRNLCNIPCSALLSSDITASVNCACKIVSDG  
NGMNAWVAWRNRCKGTDVQAWIRGCRL
```

### Bovine lysozyme sequence :

```
>sp|P04421|LYSC_BOVIN Lysozyme C OS=Bos taurus GN=LYZ1 PE=1 SV=2  
MKALVILGFLFLSVAVQGKVFERCELARTLKKLGLDGYKGVSLANWLCLTKWESSIONTKA  
TNYNPSSESTDYGIFQINSKWWCNDGKTPNAVDGCHVSCRELMENDIAKAVACAKHIVSE  
QGITAWVAWKSHCRDHDVSSYVEGCTL
```

### Sequence associated to PDB coordinates file of Bovine lysozyme:

```
>2Z2F:A|PDBID|CHAIN|SEQUENCE  
KVFERCELARTLKKLGLDGYKGVSLANWLCLTKWESSIONTKA  
TNYNPSSESTDYGIFQINSKWWCNDGKTPNAVDGCHVSCRELMENDIAKAVACAKHIVSE  
QGITAWVAWKSHCRDHDVSSYVEGCTL
```

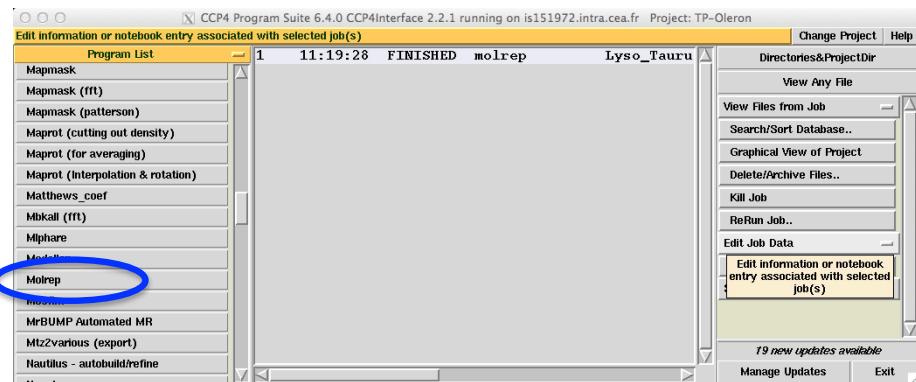
### Hen and bovine lysozymes sequence alignment :

```
>z2z2f_A Lysozyme C-2; stomach lysozyme, 1,4-beta-N-acetylmuramidase C, bacteriolytic  
enzyme, hydrolase; 1.50A {Bos taurus} SCOP: d.2.1.2  
Length = 129 Score = 154 bits (389), Expect = 1e-38, Method: Composition-based  
stats. Identities = 71/130 (54%), Positives = 96/130 (73%), Gaps = 2/130 (1%)
```

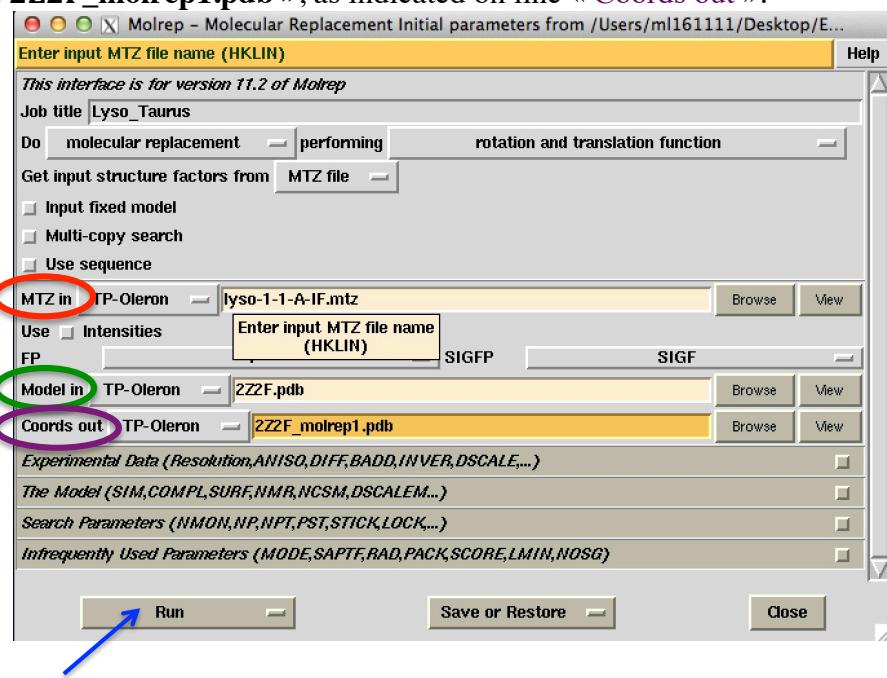
```
Query: 19 KVFGRCLEAAAMKRHGLDNYRGYSLGNWVCAAKFESNFNTQATNRNTDG-STDYGILQIN 77  
KVF RCELA +K+ GLD Y+G SL NW+C K+ES++NT+ATN N STDYGI QIN  
Sbjct: 1 KVFERCELARTLKKLGLDGYKGVSLANWLCLTKWESSIONTKATNYNPSSESTDYGIFQIN 60  
  
Query: 78 SRWWCNDGRTPGSRNLCNIPCSALLSSDITASVNCACKIVSDGNGMNAWVAWRNRCKTD 137  
S+WWCNDG+TP + + C++ CS L+ +DI +V CAK IVS+ G+ AWVAW++ C+ D  
Sbjct: 61 SKWWCNDGKTPNAVDGCHVSCRELMENDIAKAVACAKHIVSE-QGITAWVAWKSHCRDHD 119  
  
Query: 138 VQAWIRGCRL 147  
V +++ GC L  
Sbjct: 120 VSSYVEGCTL 129
```

You will now calculate the phase of the observed amplitudes  $F_{\text{obs}}(\text{hkl})$ , by the molecular replacement method using the software **MOLREP** from CCP4i package.

The interface of the software opens by clicking on **Molrep** :

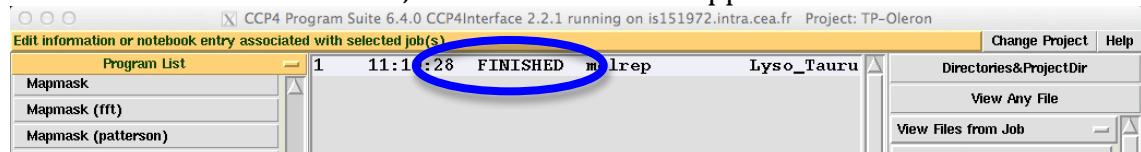


The amplitudes and their associated errors will be extracted from the file « lyso-1-1-A-IF.mtz » that you must enter on line « **MTZ in** ». The phase information will be calculated from the PDB file "2Z2F.pdb", line « **Model In** ». The « solution » PDB file will be named « **2Z2F\_molrep1.pdb** », as indicated on line « **Coords out** ».

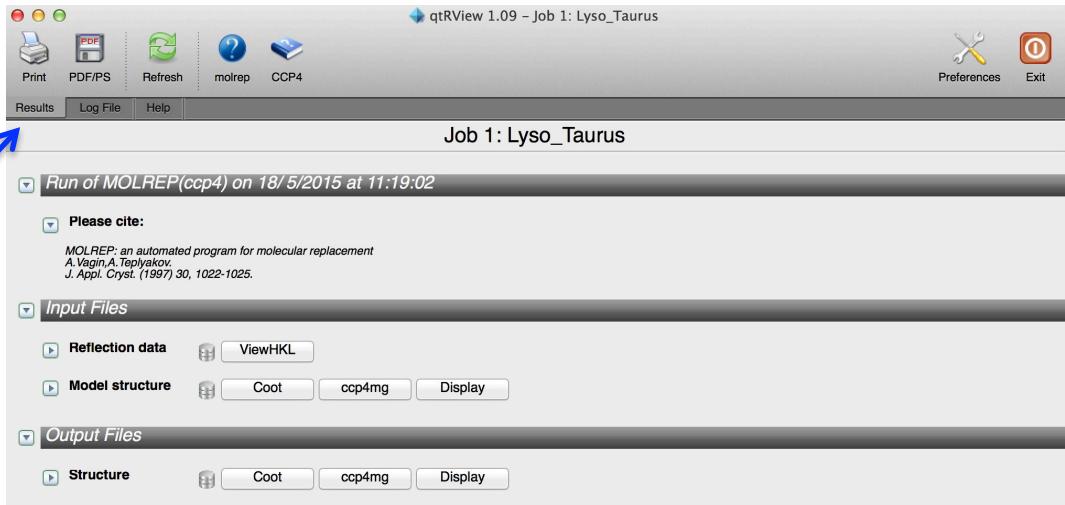


Press **Run** to start the calculation.

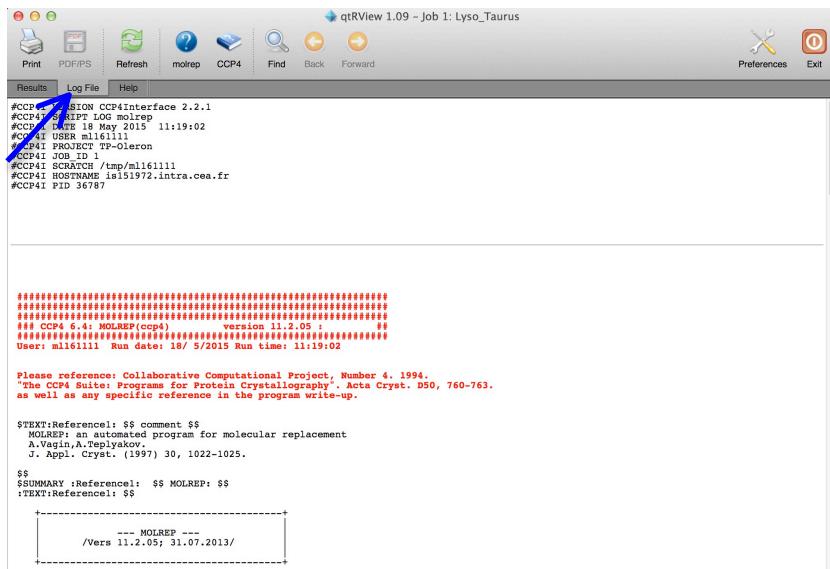
When the calculation is done, the word FINISHED appears in the CCP4i Gui.



You can open the output file by double click on the job line  
Two tabs are going to allow you to analyze the results of your calculation :



The Results section allows you to open the input and output with COOT, ccp4mg or Display.



The Log file contains the information regarding the calculation itself.

## Questions

1. Which parameters will allow to determine the correctness of **MOLREP** solution ? / *Quels paramètres vont permettre de déterminer si la solution trouvée par **MOLREP** est correcte ?*
  2. What is the reliability factor Rfac of the solution found by MOLREP ? / *Quel est le facteur de confiance Rfact de la solution trouvée par **MOLREP** ?*
  3. What contrast has the solution found by MOLREP ? / *Quel contraste a la solution trouvée par **MOLREP** ?*
  4. Open the output file structure with **COOT**, and visualize the solution. What can tell you that the solution is probably correct (or wrong) ? *Ouvrez le fichier de sortie avec **COOT** et visualisez la solution. Qu'est-ce qui peut vous indiquer que la solution est coorecte (ou fausse) ?*

## **II - REFINEMENT OF MOLECULAR REPLACEMENT SOLUTION WITH REFMAC**

### **1) Before running the refinement with Refmac ...**

Check that a **free Rfactor flag** (FreeRflag) has been set in your mtz data file.

If you have followed the XDS data processing tutorial, the FreeRflag has already been added to the mtz file (look at the uniquemtz.out log file).

If you have used imosflm, you should add the FreeRflag as follow:

- Check whether the FreeRflag is present in the mtz file:

type in the terminal window:

```
mtzdump hkl lyso-1-1-A-IF.mtz
```

```
go
```

if FreeRflag is absent, type:

```
freerflag hkl lyso-1-1-A-IF.mtz hklout lyso-1-1-A-IFfree.mtz << eof
```

```
freerfrac 0.05
```

```
end
```

```
eof
```

and check again:

```
mtzdump hkl lyso-1-1-A-IFfree.mtz
```

```
go
```

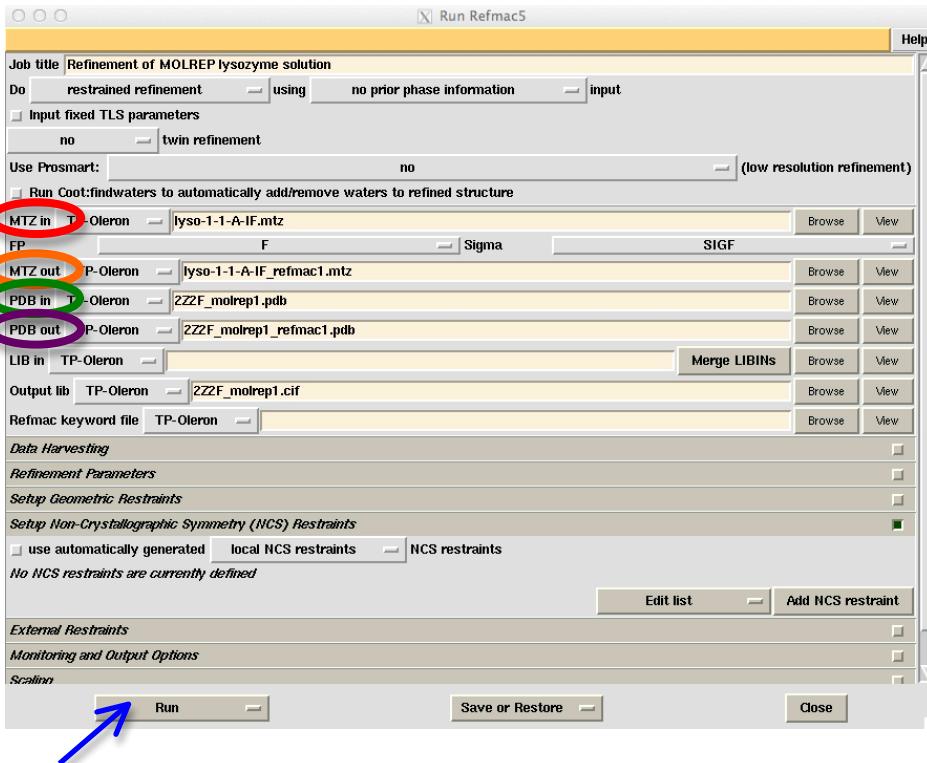
if the FreeRflag is there now, lyso-1-1-A-IFfree.mtz is the file to use for the next steps.

### **2) Refinement with Refmac**

Based on molecular replacement solution, you can initiate REFMAC :



As previously, the amplitudes and their associated errors will be extracted from the file « lyso-1-1-A-IF.mtz » that you must enter on line « **MTZ in** ». The phase information will be directly calculated from the MOLREP solution PDB file "2Z2F\_molrep1.pdb", line « **Model In** ». After refinement, the refined model PDB file will be « **2Z2F\_molrep1\_refmac1.pdb** », as indicated on line « **PDB out** ». The amplitudes, and electron density map coefficient will be in the binary file « **lyso-1-1-A-IF\_refmac1.mtz** », as indicated on line **MTZ out**.



Press **Run** to start the calculation.

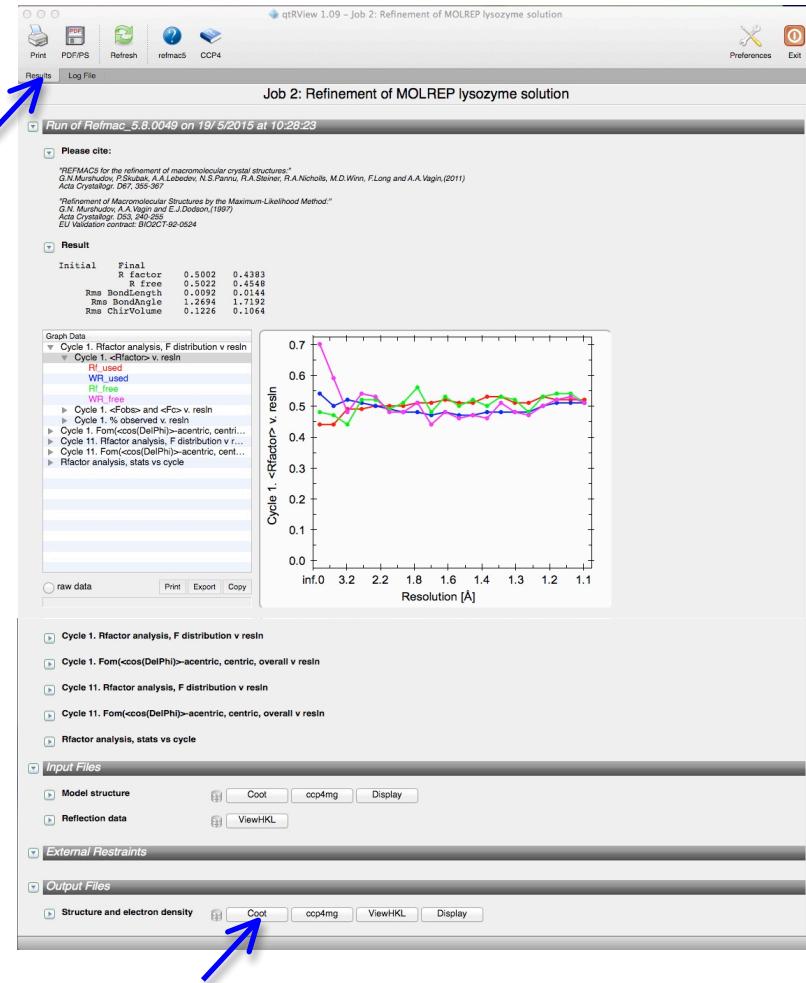
The REFMAC job will be RUNNING first, then FINISHED :

Program List	Date	Status	Program	Project
ProDry	2 10:28:2	RUNNING	refmac5	Refinement
Profess	1 18 May 15	FINISHED	molrep	Lyso_Tauru
ProSMART				

Program List	Date	Status	Program	Project
ProDry	2 10:28:5	FINISHED	refmac5	Refinement
Profess	1 18 May 15	FINISHED	molrep	Lyso_Tauru

As previously, you can open the output file by double click on the job line. There is also one Results section and one Log File section in the output file.



Press Coot to open the output files (pdb and mtz files) in coot.

```

qtRView 1.09 - Job 2: Refinement of MOLREP lysozyme solution
Print PDFPS Refresh refmac5 CCP4 Find Back Forward Preferences Exit

Results Log File
Job 2: Refinement of MOLREP lysozyme solution

Run of Refmac_5.8.0049 on 19/5/2015 at 10:28:23

Please cite:
REFMAC5 for the refinement of macromolecular crystal structures
G.N.Murshudov, P.Skubak, A.A.Lebedev, N.S.Pannier, R.A.Steiner, R.A.Nicholls, M.D.Winn, F.Long and A.A.Vagin (2011)
Acta Crystallogr. D67, 353-367

Refinement of Macromolecular Structures by the Maximum-Likelihood Method
G.N.Murshudov, P.Skubak and E.J.Dodson (1997)
Acta Crystallogr. D53, 240-255
EU Validation contract: BIO2CT-92-0524

Result
Initial Final
Initial R factor 0.5002 0.4383
R free 0.5022 0.4548
Rms BondLength 0.0392 0.0441
Rms BondAngle 1.2694 1.7192
Rms ChirVolume 0.1226 0.1064

Graph Data
Cycle 1. Rfactor analysis, F distribution v resin
Cycle 1. <Rfactor> v resin
RF used
WR used
WR free
WRI free
Cycle 1. Fobs-><cos(DelPhi)>-acentric, centri...
Cycle 11. Rfactor analysis, F distribution v resin
Cycle 11. Fom(<cos(DelPhi)>-acentric, centri...
Rfactor analysis, stats vs cycle

Cycle 1. Rfactor analysis, F distribution v resin
Cycle 1. Fom(<cos(DelPhi)>-acentric, overall v resin)
Cycle 11. Rfactor analysis, F distribution v resin
Cycle 11. Fom(<cos(DelPhi)>-acentric, centric, overall v resin)
Rfactor analysis, stats vs cycle

Input Files
Model structure Coot ccp4mg Display
Reflection data ViewHKL

External Restraints

Output Files
Structure and electron density Coot ccp4mg ViewHKL Display

```

## Questions

- What parameters inform on the refinement quality?
- What appears in the Coot window?
- Based on the lysozyme sequences alignment, can you correct the refined structure in Coot?

### 3) Model building with Coot

#### The minimum to know about Coot:

Coot is fairly “intuitive”: most of the necessary action can be selected from the icons on the right icon bar of the Coot window: just put the mouse arrow on the icon to know what it does.

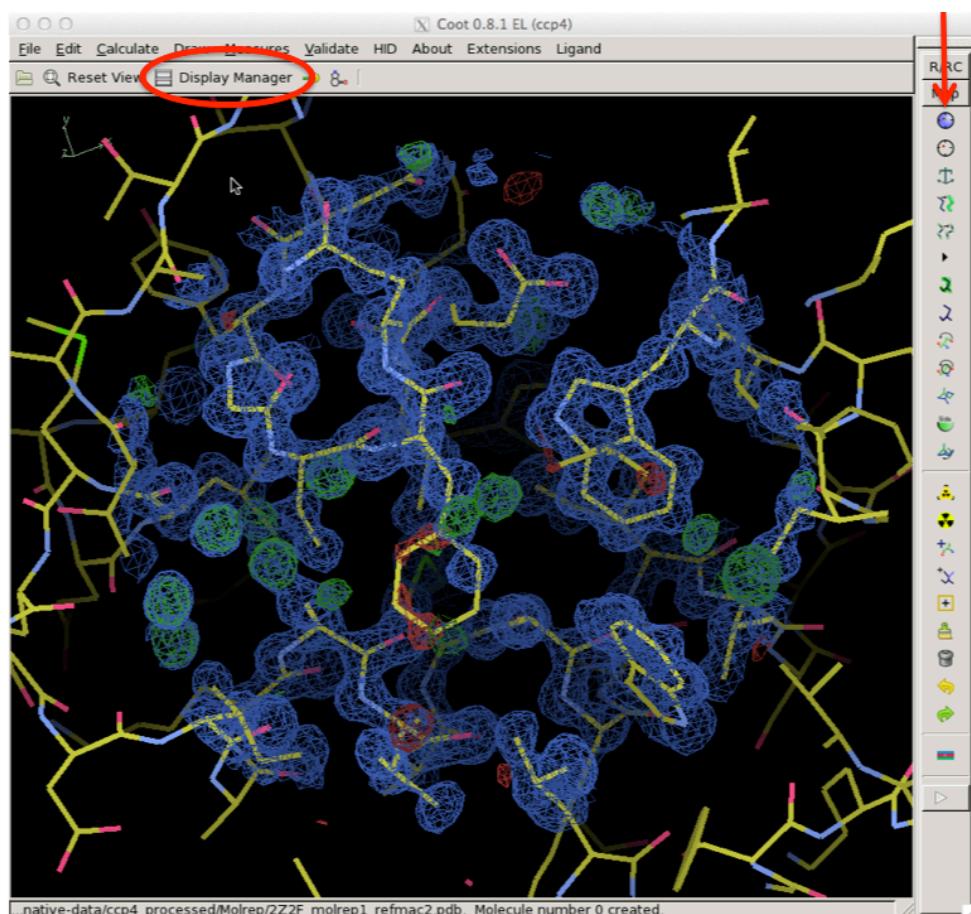
#### Before to start model building

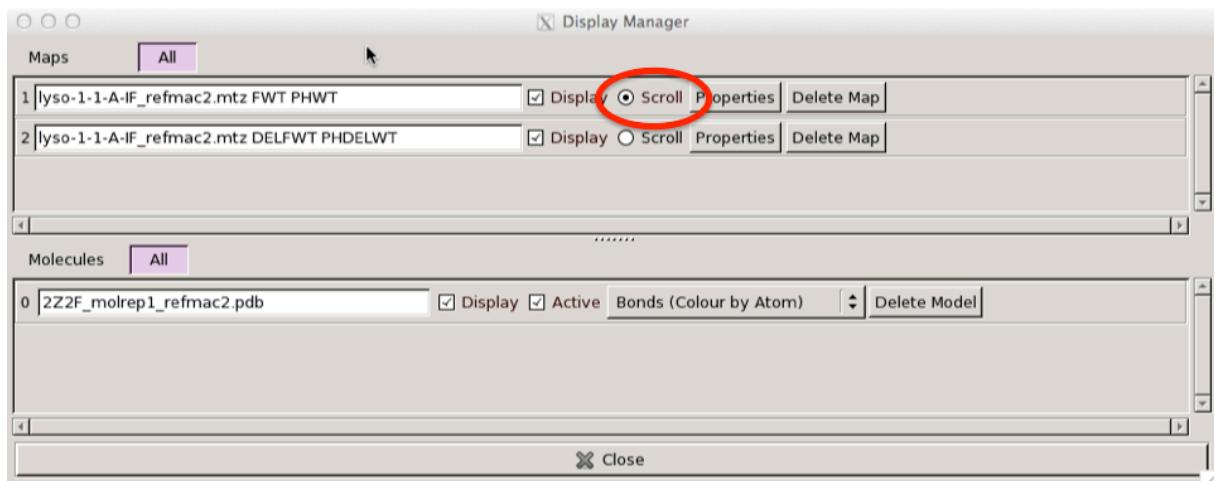
Check out the contouring level of electron density maps:

{2Fobs – Fcalc} maps (FWT, PHWT) should be contoured at 1 rms

{Fobs – Fcalc} residual maps (DELFWT, PHDELWT) should be contoured at +3 or -3 rms

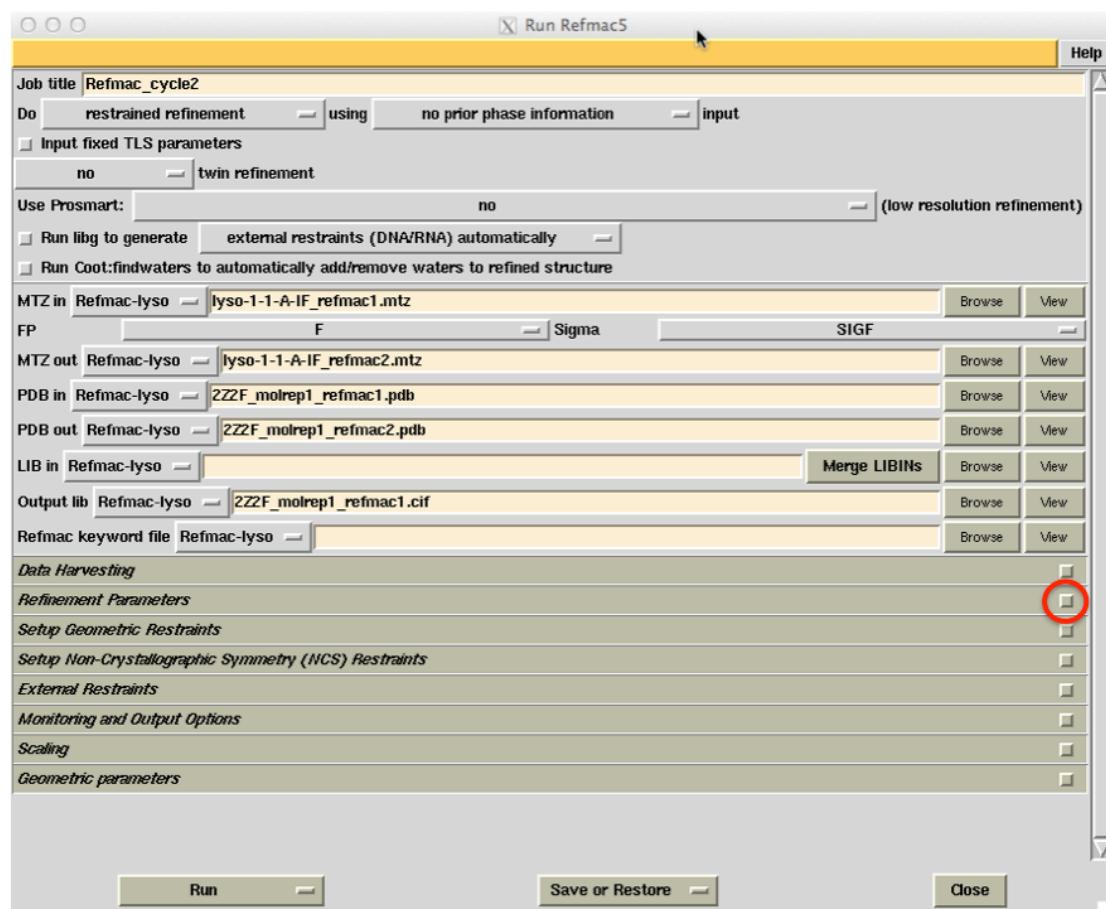
Clic on “Display Manager” and select the scroll button corresponding to the desired map; the use the middle wheel of your mouse to increase or decrease the level.

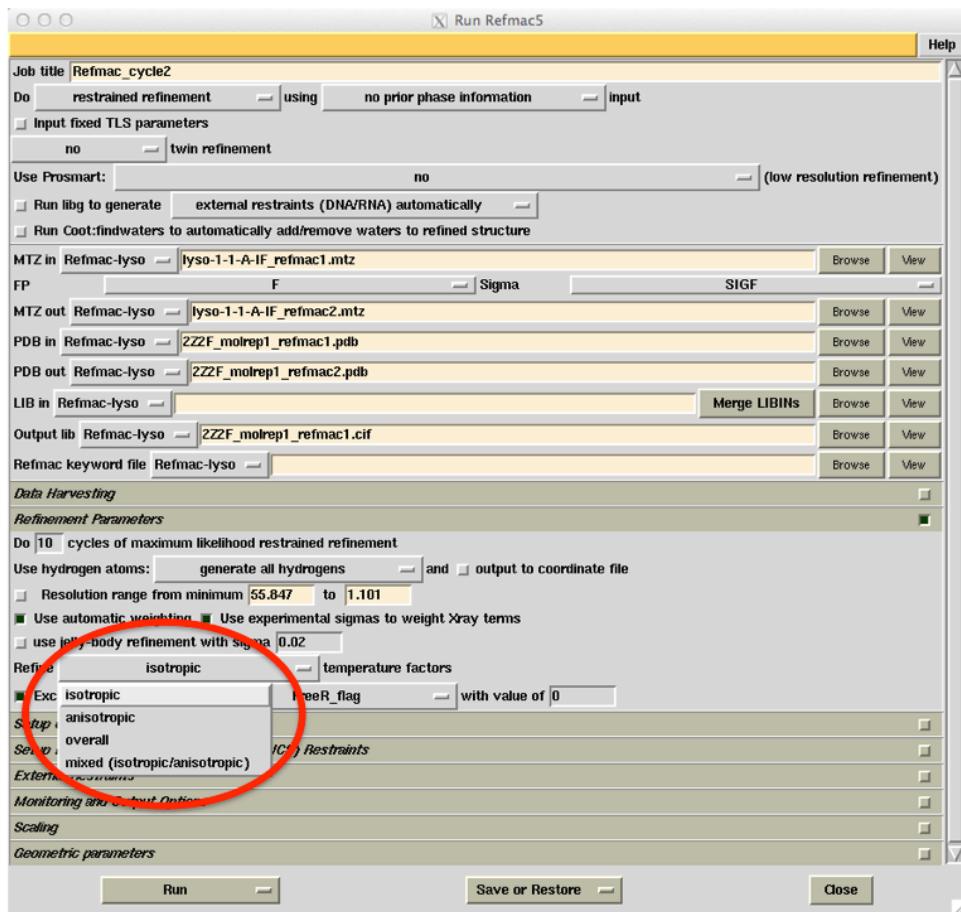




- Once you have made the desired structural change in coot, you can save the new coordinates of the model ("file" menu) and run refmac again.
- You do cycle coot and refmac steps until you have interpreted all the significant electron density, with a good fit between your model and the electron density.

At some point (but you might not reach it during the practical), you may need to look at the refinement parameters submenu in Refmac:





You may have to modify:

- the number of cycles: put more than 10 if your Rfactor is not stable after 10 cycles
- the B factor (temperature factor) : isotropic (default, one B factor per atom), overall (one B factor for the entire model, recommended for low resolution data, i.e.  $d_{\max} > 3.2 \text{ \AA}$ ), anisotropic B factor (6 parameters per atom, model the anisotropy of atom motion, useful for high resolution data, i.e.  $d_{\max} < 1.3 \text{ \AA}$ )