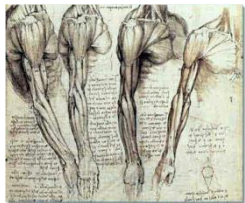


Que peut-on voir avec un microscope électronique à transmission ?

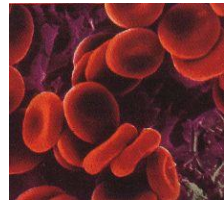
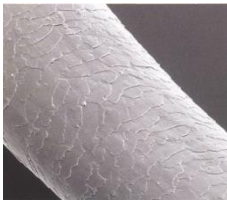
What can we see with a TEM ?

Origin of research in biology

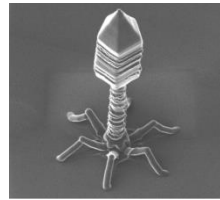
15th century



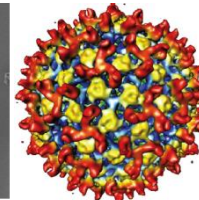
17th century



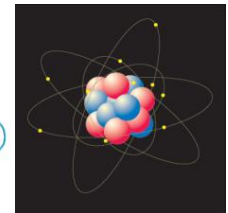
19th century



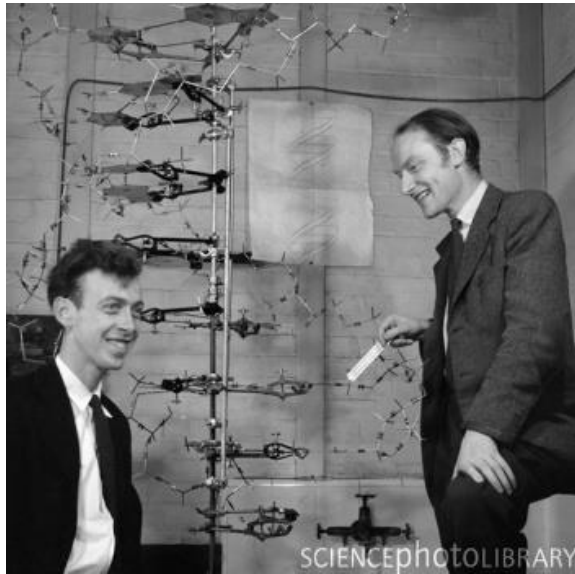
20th century



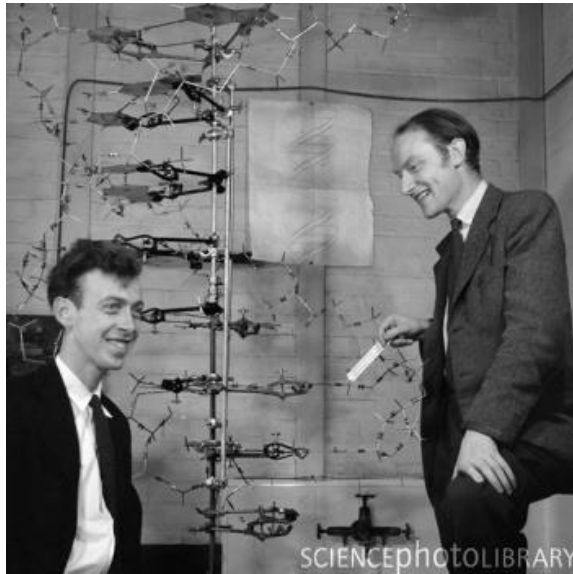
21st century



Structure/function studies



Structure/function studies



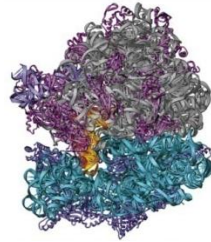
Watson/Crick (1953)



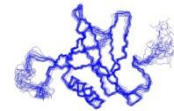
Perutz/Kendrew (1959)

Structural biology today

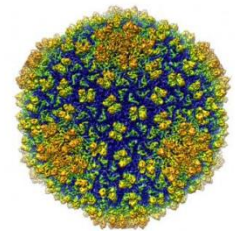
X-ray crystallography



Nuclear magnetic resonance (NMR)

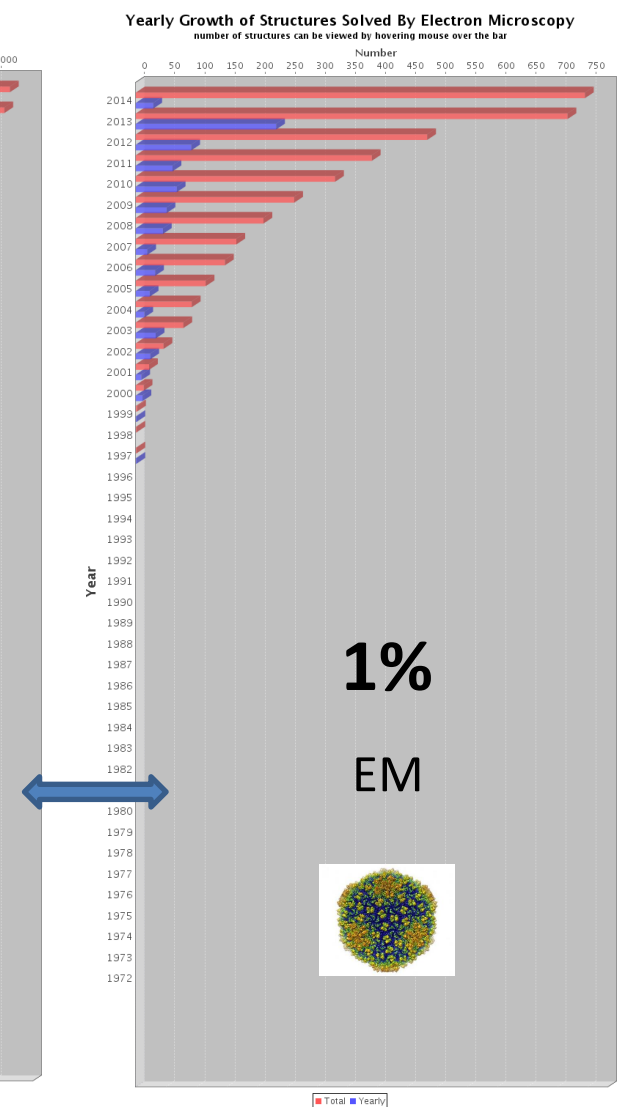
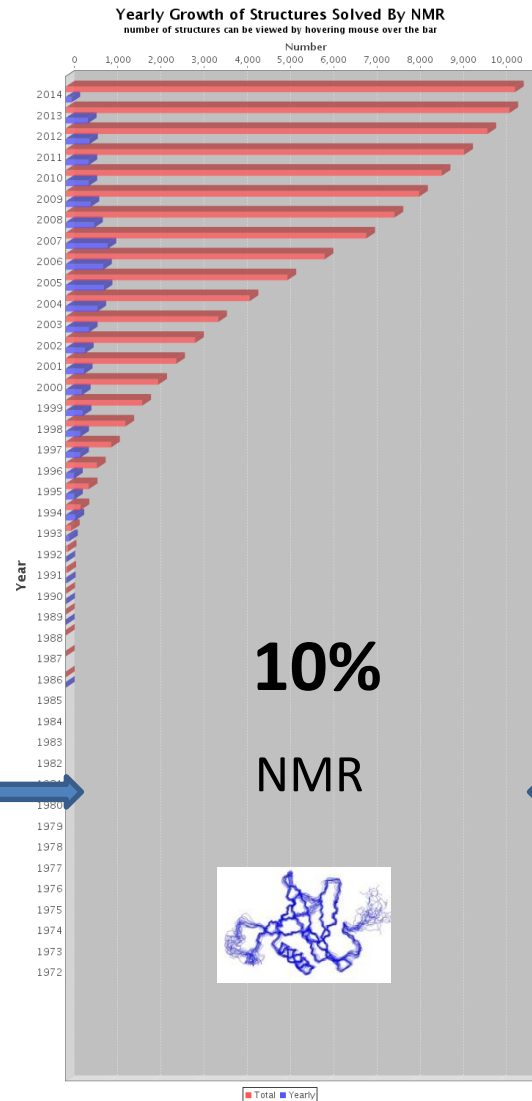
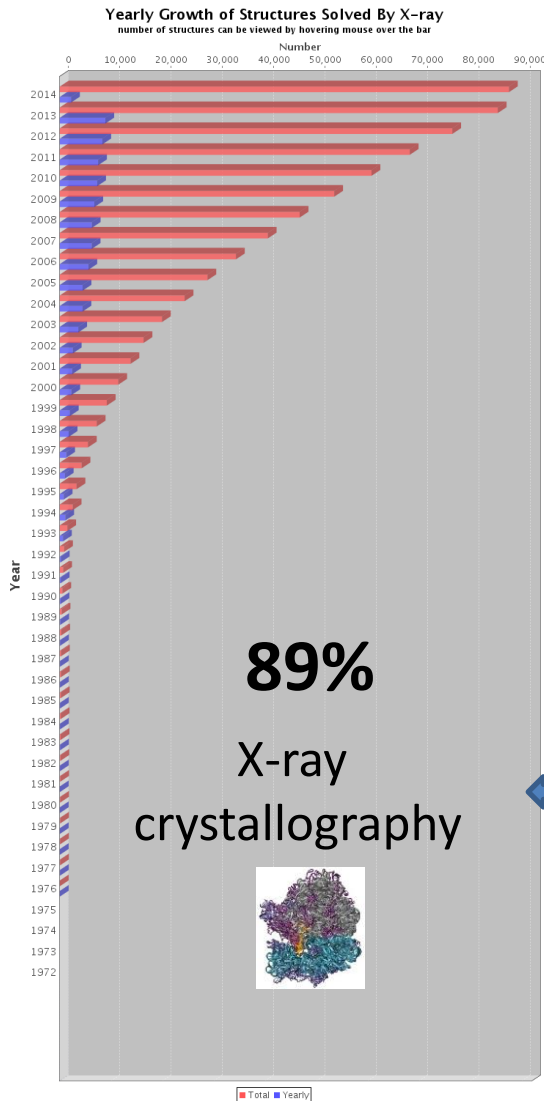


Electron microscopy (EM)

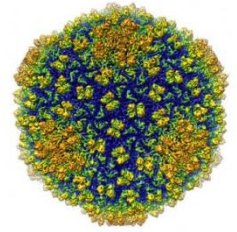


SAXS, ...

Structural biology statistics



Electron microscopy



- What type of biological material can we study (cell, protein, atom) ?
- Difference optical microscopy vs electronic microscopy ?
- Which microscopy ? For what for?
- Composition of an electron microscope
- What structural informations can you get?

Size of objects

E. coli

Mitochondria
of mammalian
cell

Ribosome

Nucleus of
mammalian
cell

Globular
monomeric
protein

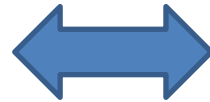
Human oocyte

S. cerevisiae

Mammalian
cell

What is the resolution that you can achieve ?

- To summarize, it's the smallest detail that you can observe.



X-ray crystallography, NMR ?

Wavelength of light

Rayleigh criterion:

$$d = 0,61 \lambda / n \sin(\alpha)$$

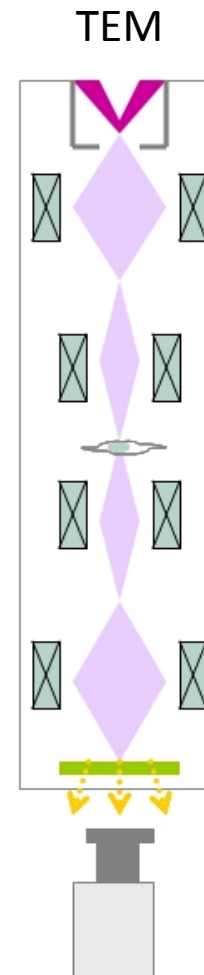
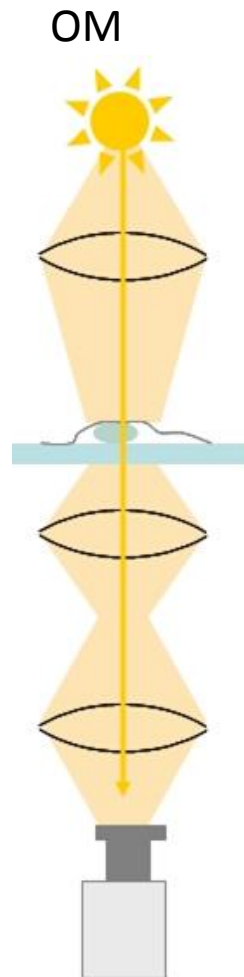
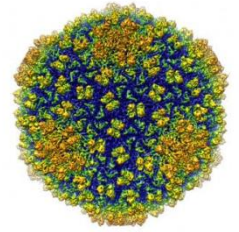
Numerical aperture
of the lens

distance

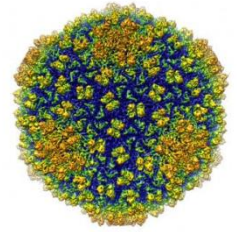
Refraction index of studied matter

To simplify: $d \approx 0.5 \lambda$

Optical vs electron microscope



Particules sources



Photon

Mass

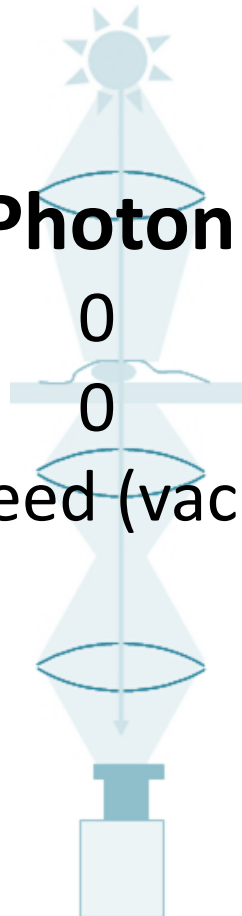
0

Charge

0

Speed

light speed (vacuum)

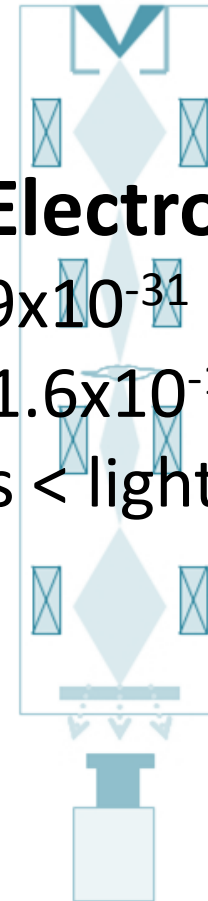


Electron

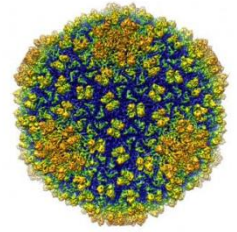
9×10^{-31} kg

-1.6×10^{-19} C

$0 \leq s < \text{light speed}$



Wave-particle duality

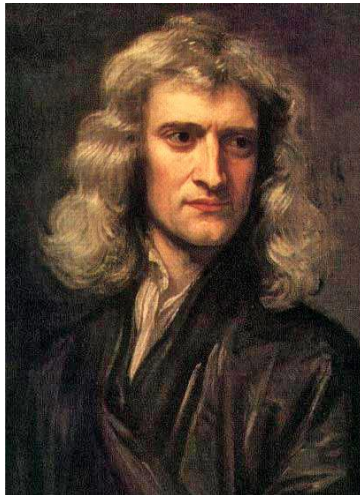


Relation Acceleration tension - wavelength

$$E_c = \frac{1}{2} m v^2$$

$$\lambda = \frac{h}{m v}$$

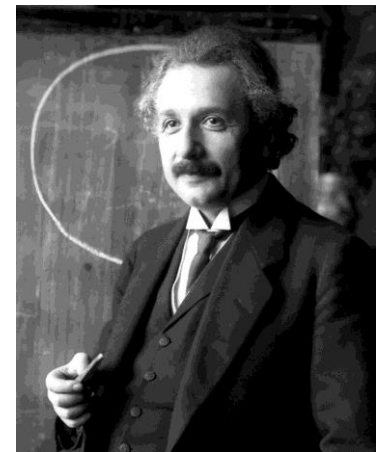
$$m = \frac{m_0}{\sqrt{1 - \frac{v^2}{c^2}}}$$



Isaac Newton

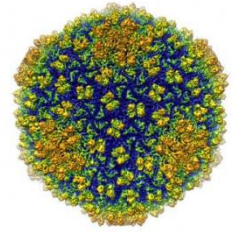


Louis de Broglie



Albert Einstein

Wave-particle duality

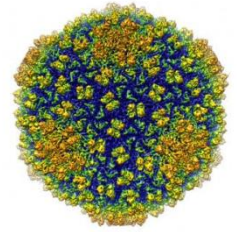


Relation Acceleration voltage - wavelength

$$E_c = \frac{1}{2} m v^2 \quad \lambda = \frac{h}{m v} \quad m = \frac{m_0}{\sqrt{1 - \frac{v^2}{c^2}}}$$

$$\lambda = \frac{1.23}{\sqrt{V + 10^{-6} V^2}} \text{ nm}$$

Wave-particle duality



Relation Acceleration voltage - wavelength

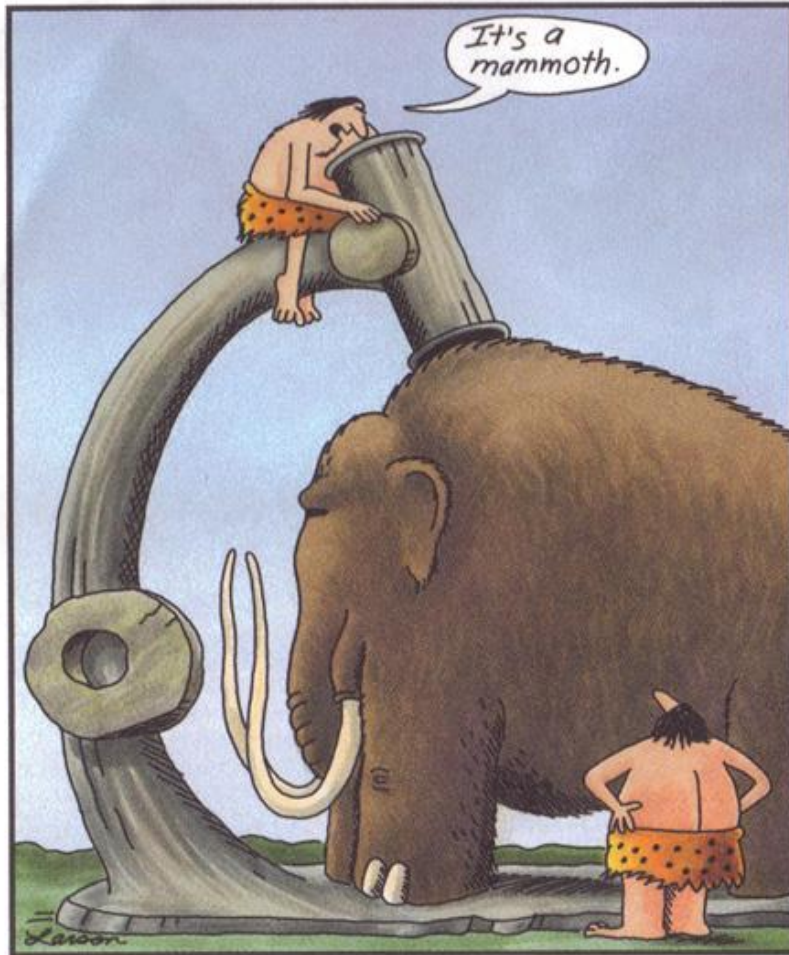
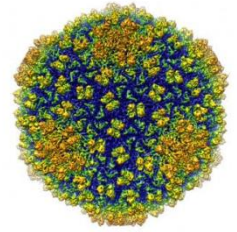
V	λ (nm)	Theoretical resolution (nm)
10,000	0.01223	0.00611
50,000	0.00536	0.00268
100,000	0.00370	0.00185
1,000,000	0.00086	0.00043

80-300 kV



Electron microscopy

History

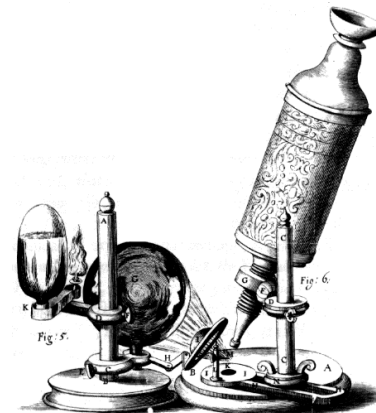


Early microscope

1600-1700

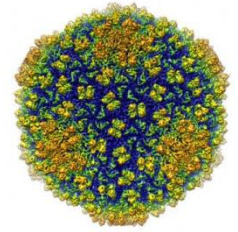


Van Leeuwenhoek



Electron microscopy

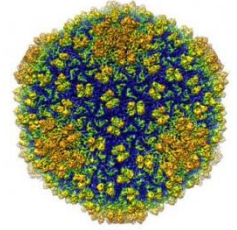
History



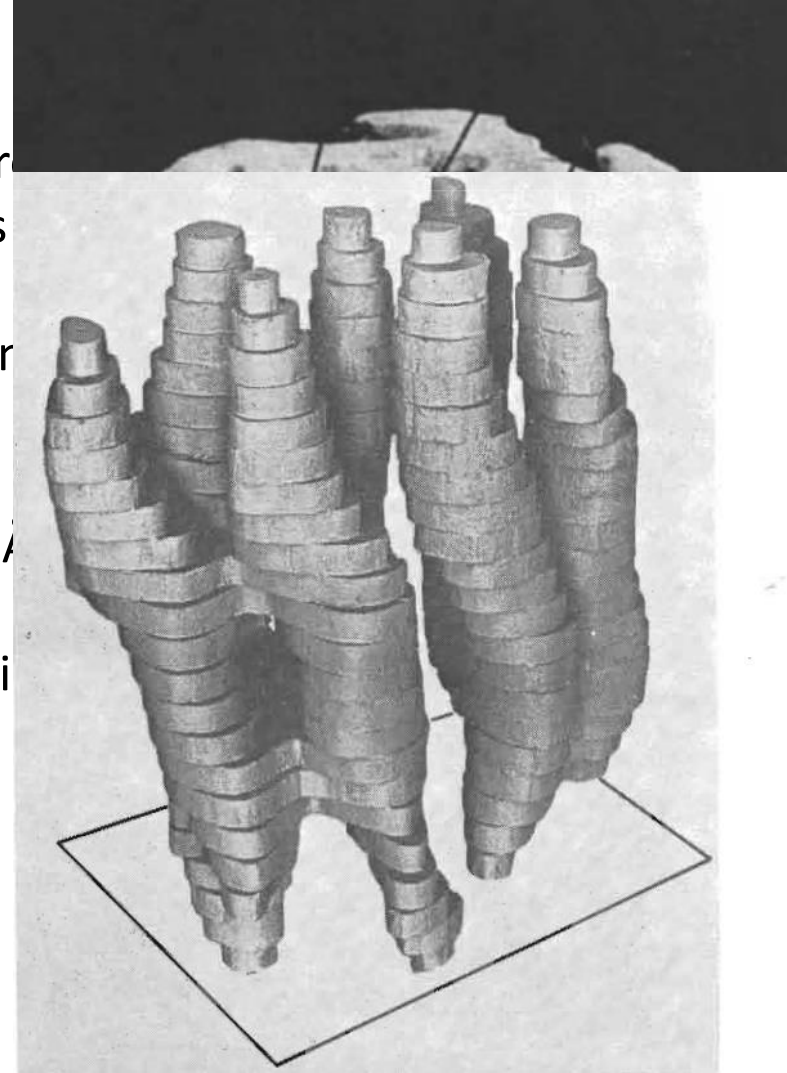
DATES	NAMES	HIGHLIGHTS
1897	J.J. Thomson	Electron discovery
1924	L. de Broglie	Identification of electron wavelength in movement
1926	H. Busch	Characterization of lens effects on magnetic and electric fields with electrons
1929	E. Ruska	Ph.D on magnetic lenses
1931	Knoll & Ruska	Building the first electron microscope
1934	Driest & Muller	Resolution better than optical microscope
1938	von Borries & Ruska	First microscope (Siemens) - 10 nm resolution
1940	RCA	Commercial microscope - 2.4 nm resolution
1945		resolution better than 1.0 nm

Electron microscopy

History

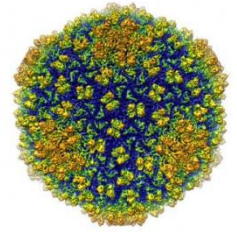


DATES	NAMES	Highlights
1960-1970	France/Japon	High voltage micro material sciences
1968	de Rosier et Klug	3D reconstruction from EM images
1975	Unwin and Henderson	3D structure at 7 Å
1981	Dubochet et al	Sample preparati



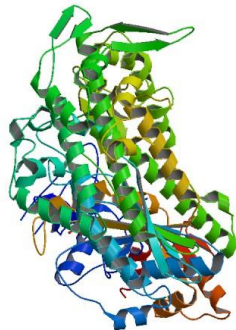
EM in the 1990-2000s

For what for?

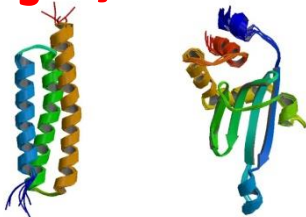


To combine structural informations from other methods (X-ray crystallography, NMR, SAXS, MS...) to link the structure and function of a protein (or a macromolecular complex)

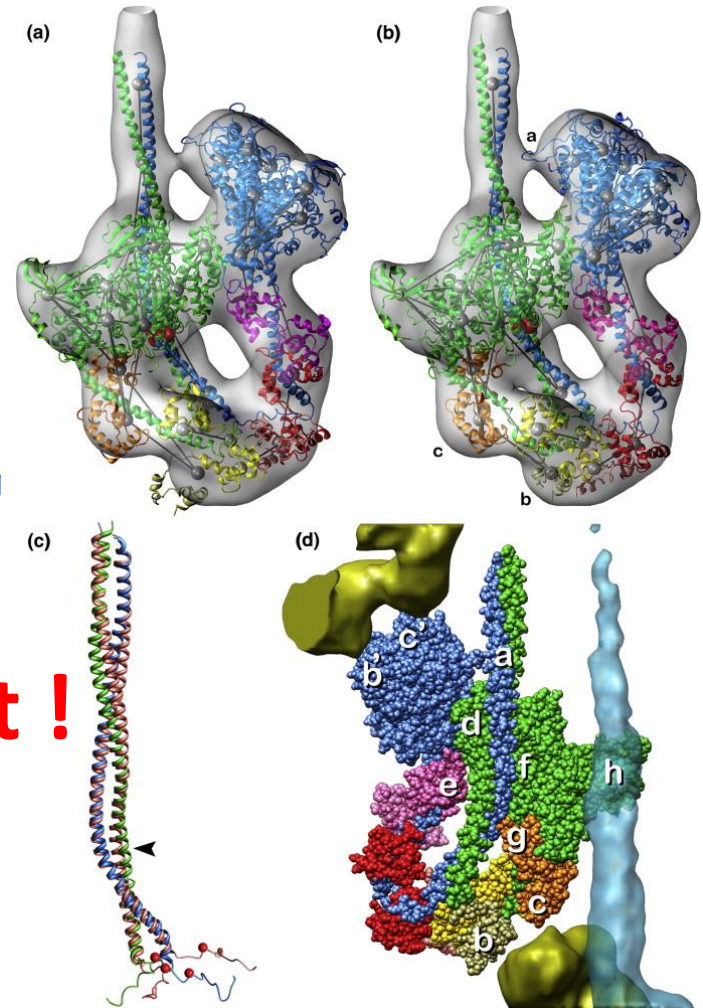
X-ray
crystallography



NMR

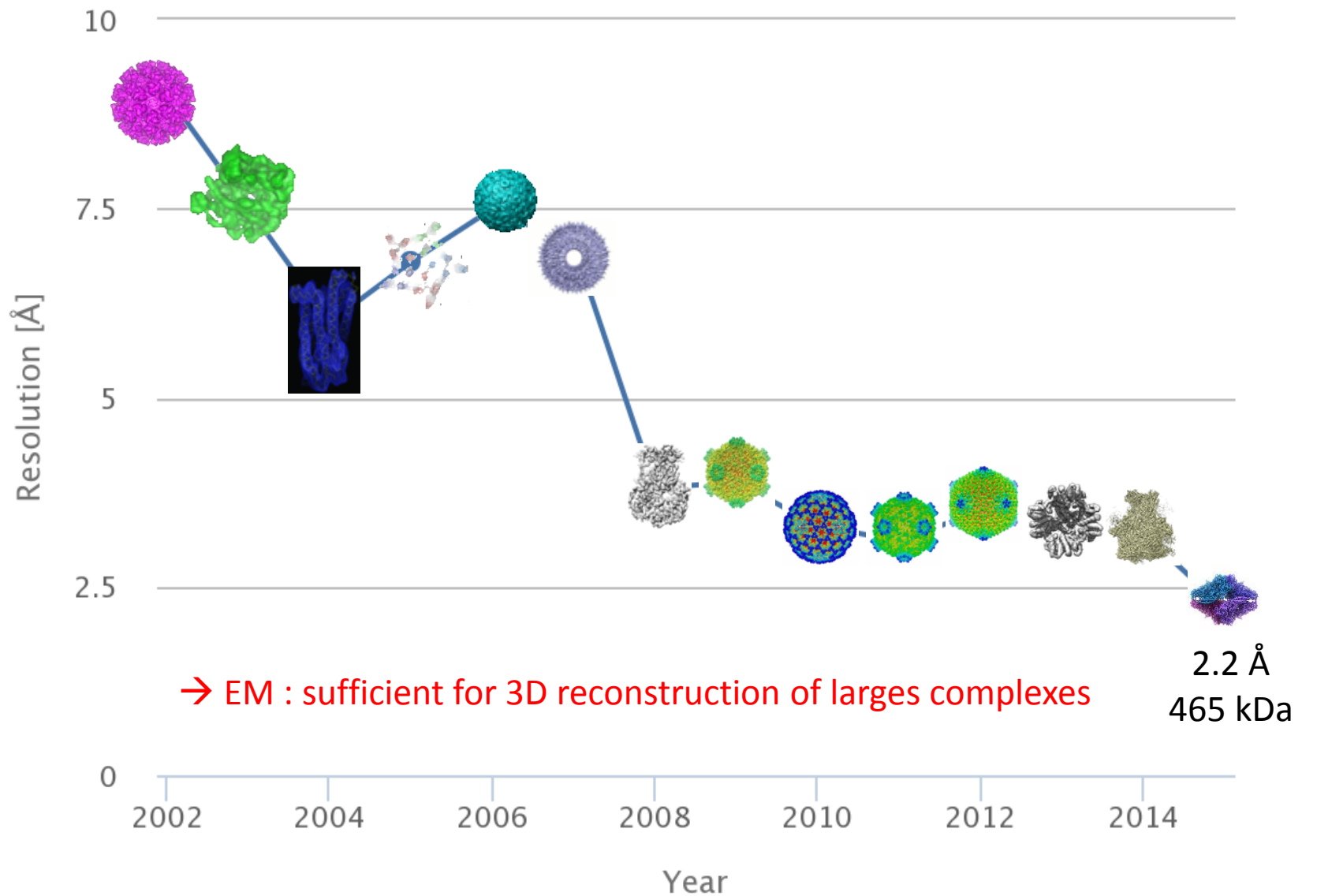


Mais ça, c'était avant !



Alamo et al, 2008

Resolution trends



→ EM : sufficient for 3D reconstruction of larges complexes

2.2 Å
465 kDa

EM in april 2015

Structure of the human 80S ribosome.

Khatter H, Myasnikov AG, Natchiar SK, Klaholz BP.
Nature. 2015 Apr 30;520(7549):640-5.

The structure of the human mitochondrial ribosome.

Amunts A, Brown A, Toots J, Scheres SH, Ramakrishnan V.
Science. 2015 Apr 3;348(6230):95-8.

The complete structure of the 55S mammalian mitochondrial ribosome.

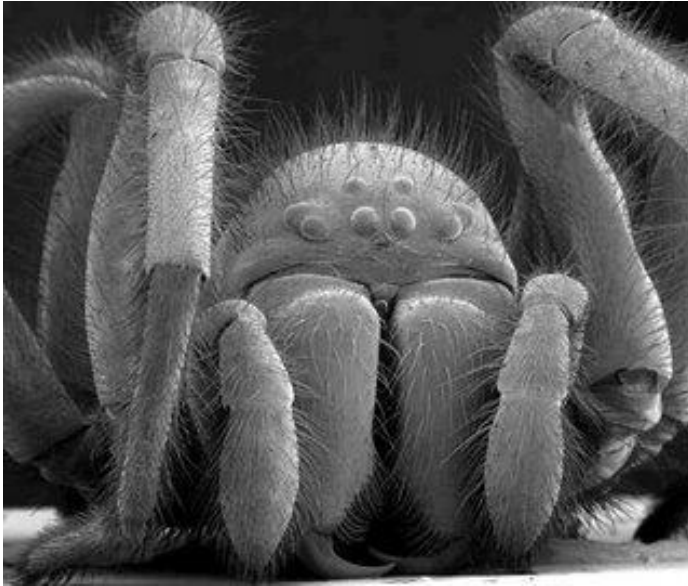
Greber BJ, Bieri P, Leibundgut M, Leitner A, Aebersold R, Boehringer D, Ban N.
Science. 2015 Apr 17;348(6232):303-8.

Structure of the E. coli ribosome-EF-Tu complex at <3 Å resolution by Cs-corrected cryo-EM.

Fischer N, Neumann P, Konevega AL, Bock LV, Ficner R, Rodnina MV, Stark H.
Nature. 2015 Apr 23;520(7548):567-70.

Electron microscopy types

Scanning electron
microscopy (SEM)



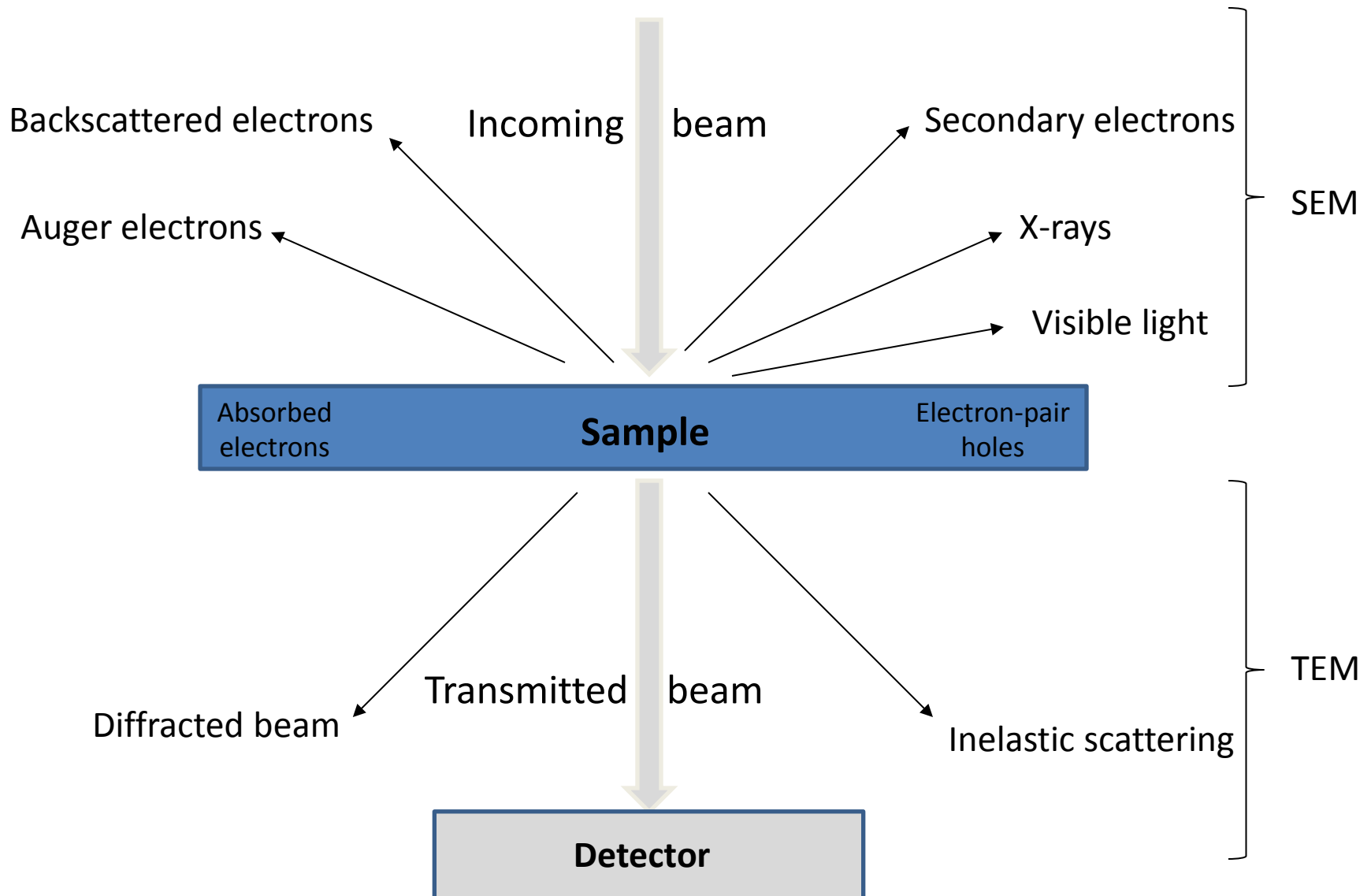
Transmission electron
Microscopy (TEM)



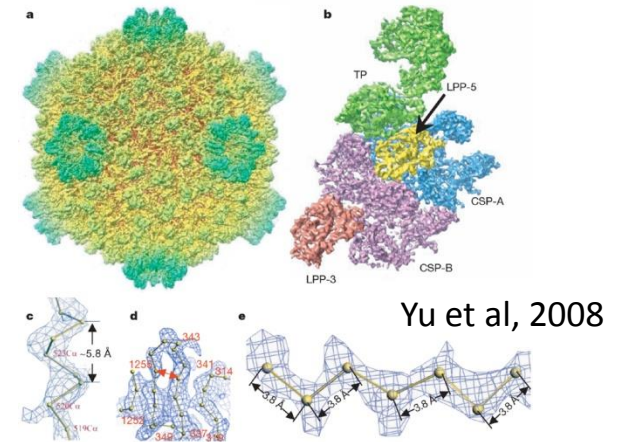
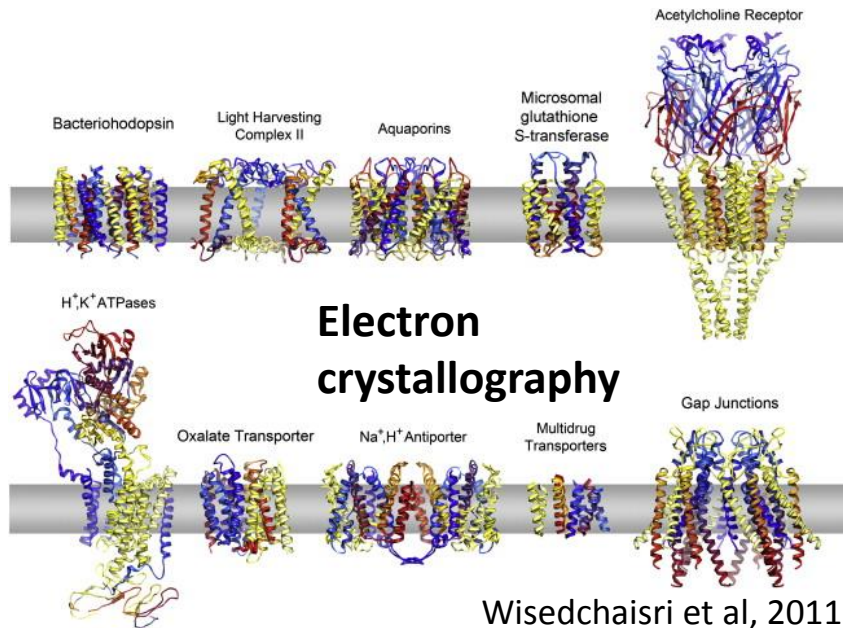
≠ Atomic force microscopy (AFM)



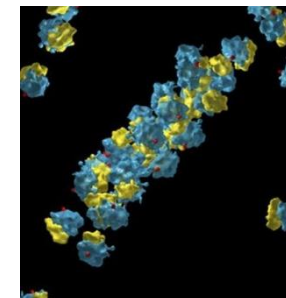
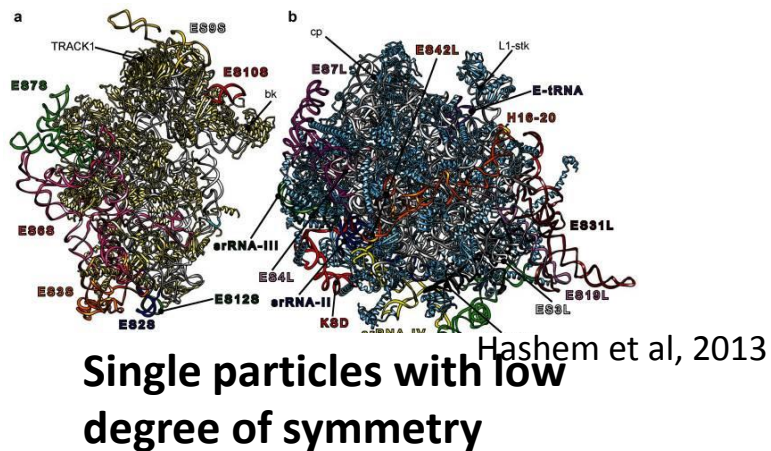
How is an image formed?



Fields in transmission electron microscopy



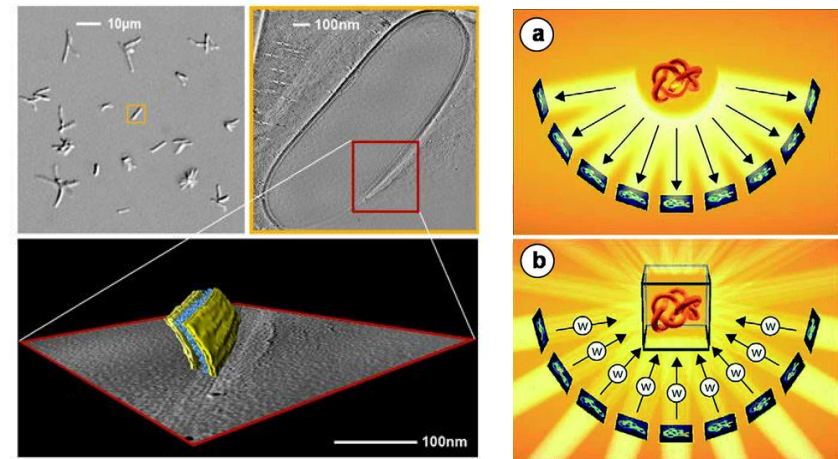
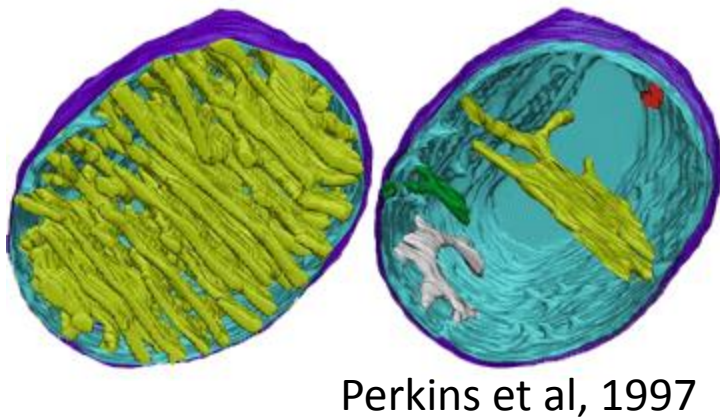
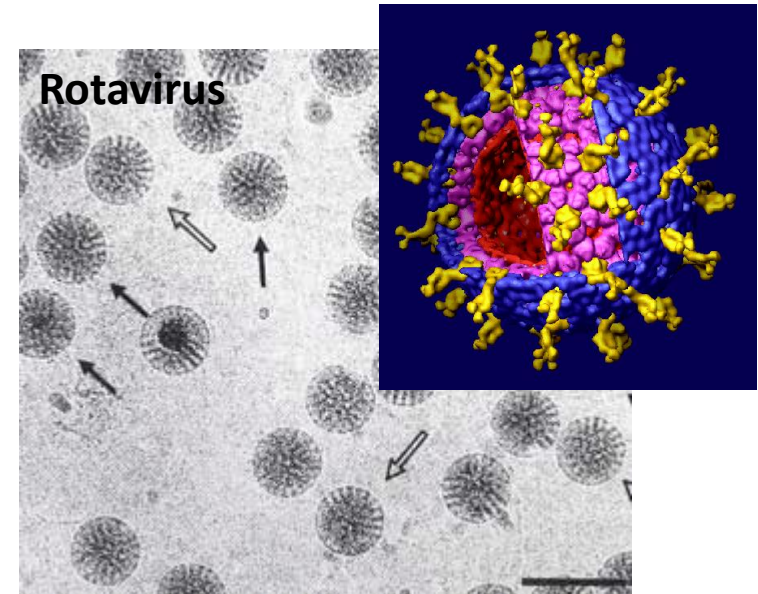
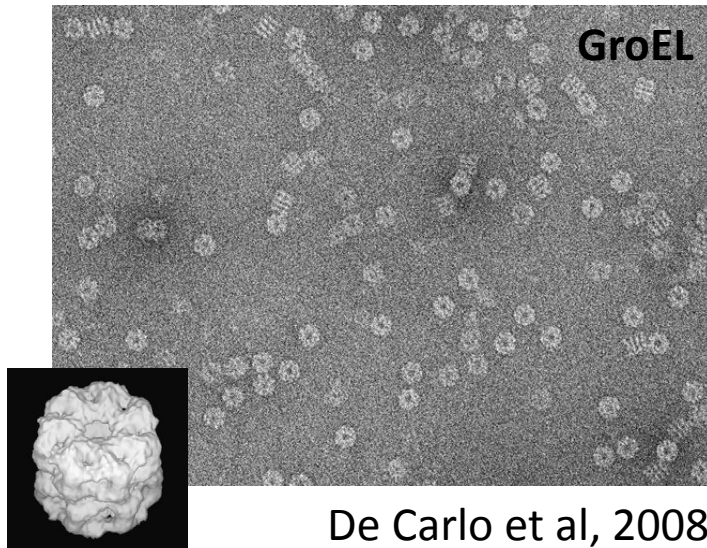
Single particles with high degree of symmetry



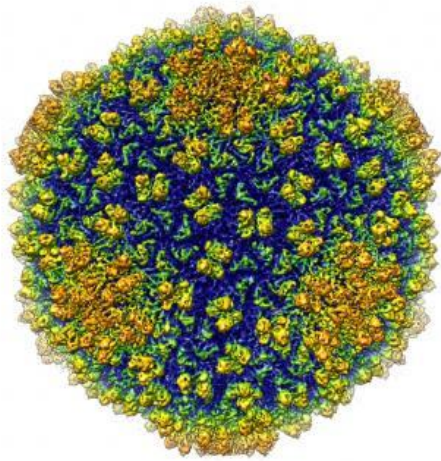
Brandt et al, 2010

Tomography

What can we see with a TEM?



Structure of a protein complex obtained by electron microscopy

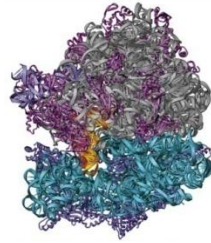


Advantages :

- 3D structure of the protein complex
- (very) large complexes (protein → cell)
- No crystal needed !
- Small amount of material

Quantity needed for an experiment

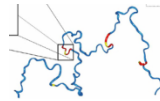
X-ray crystallography



$\sim 200 \mu\text{M}$

$\sim 100 \mu\text{L}$

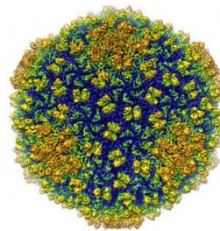
NMR



$\sim 200 \mu\text{M}$

$\sim 200 \mu\text{L}$

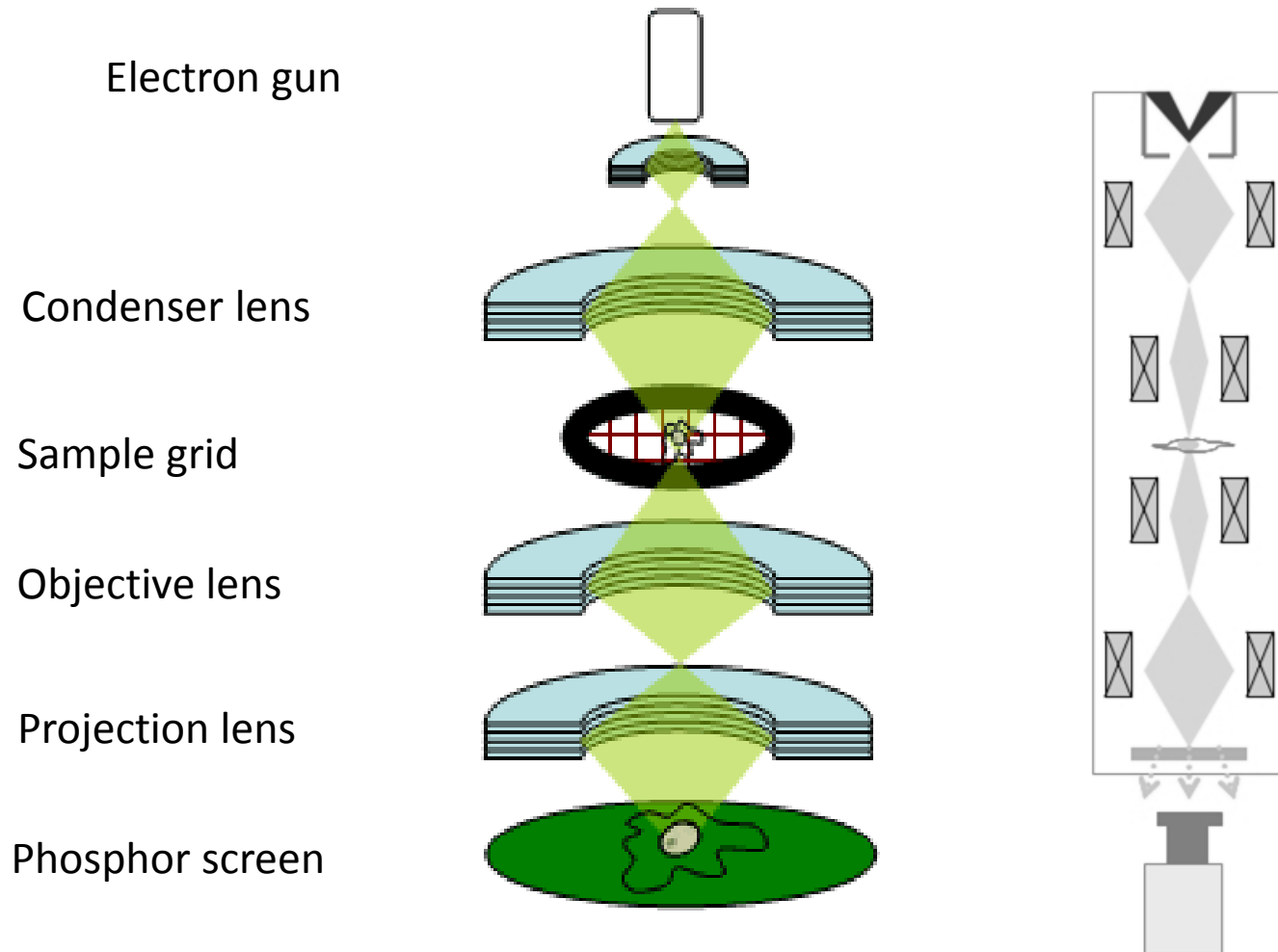
Electron microscopy



$\sim 0.2 \mu\text{M}$

$\sim 50 \mu\text{L}$

Composition of a TEM



Microscope drawbacks

And solutions...

Stability:

- Mechanical vibrations
- Acoustic vibrations
- Magnetic fields
- Electron source
- Lens current

How does a modern TEM look like ?

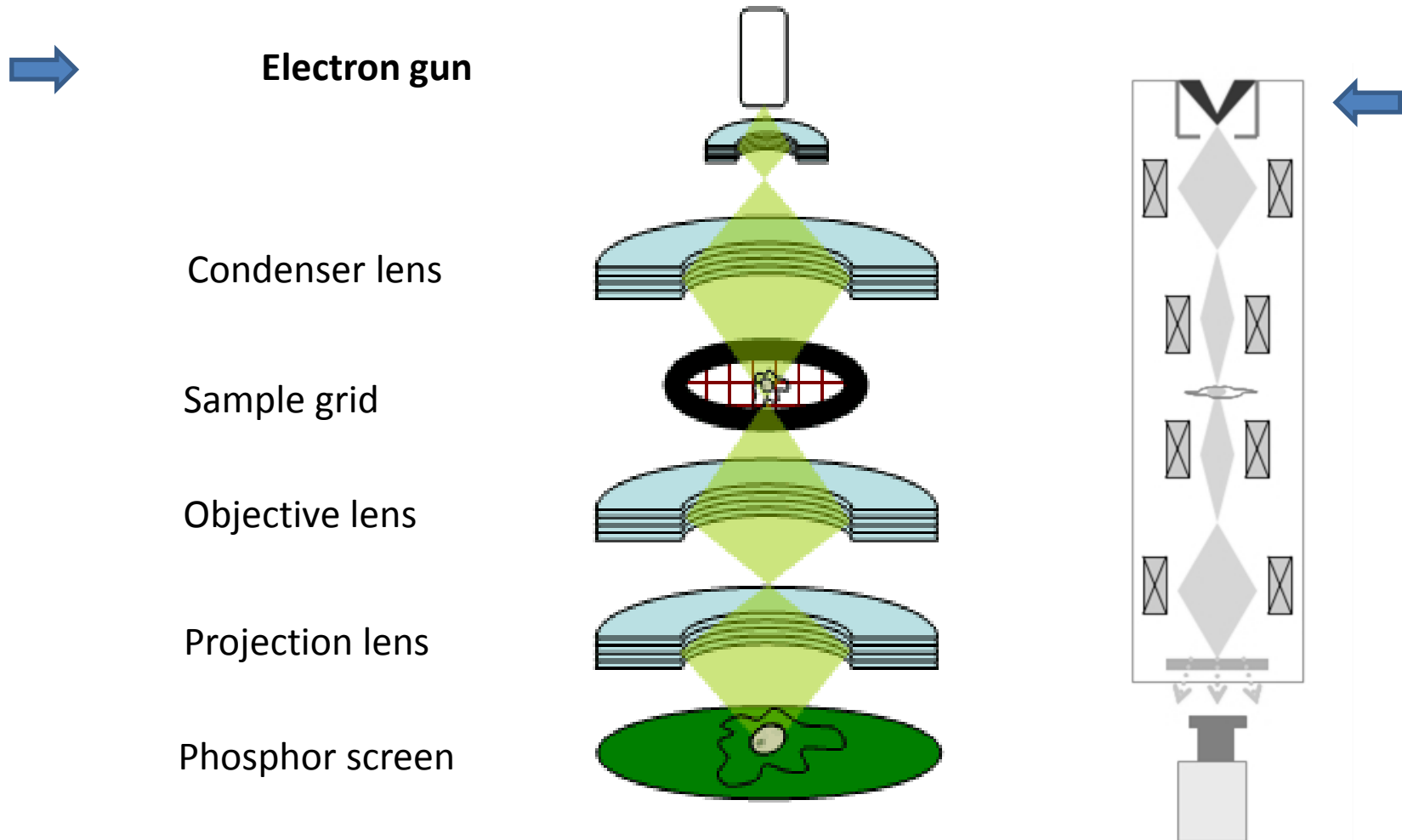


JEOL -3200 FSC (Japan)

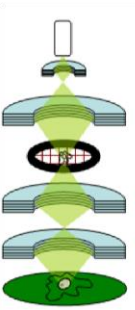


FEI-Titan Krios (USA)

Composition of a TEM



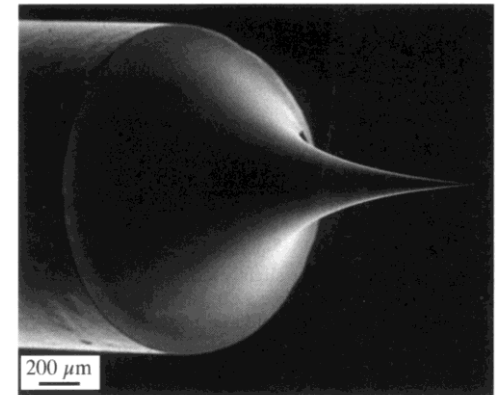
Source of electrons



- Thermoionic gun



- Field emission gun (FEG)



Field emission gun (FEG)

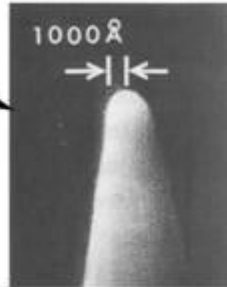
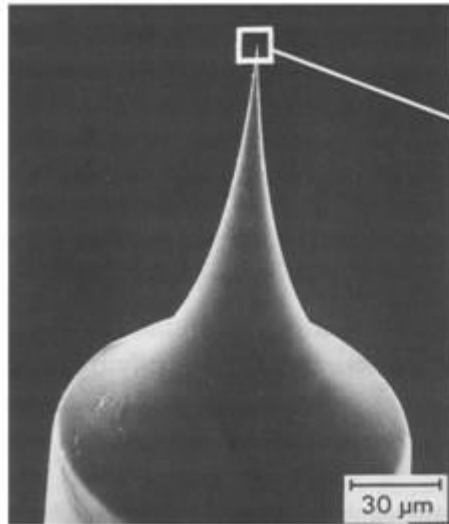
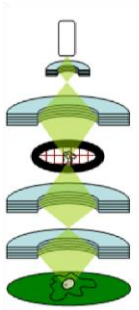
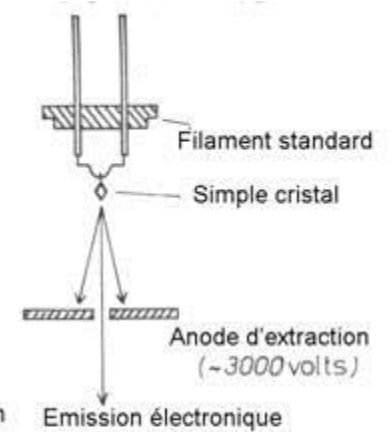
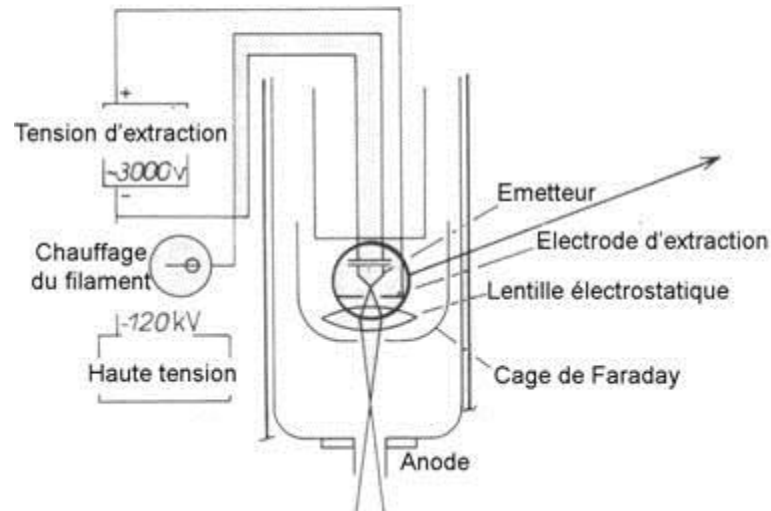


Image agrandie



Composition of a TEM

Electron gun

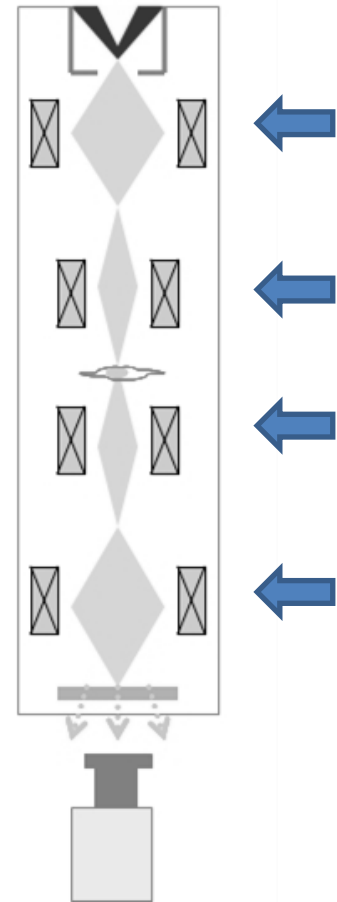
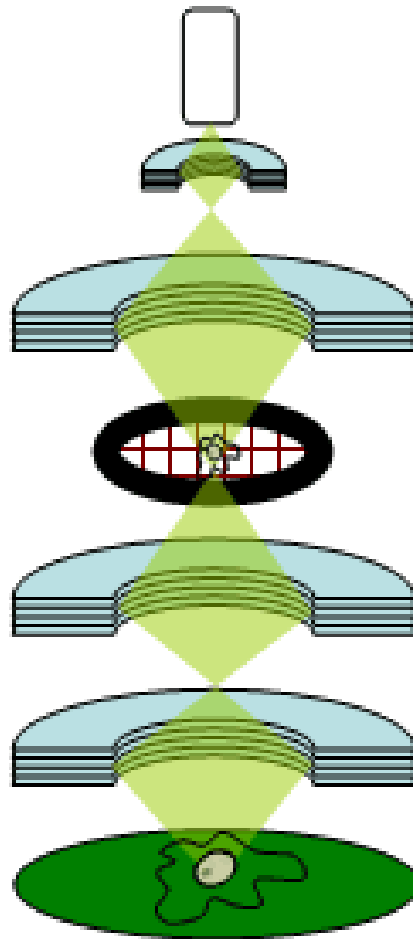
→ **Condenser lens**

Sample grid

→ **Objective lens**

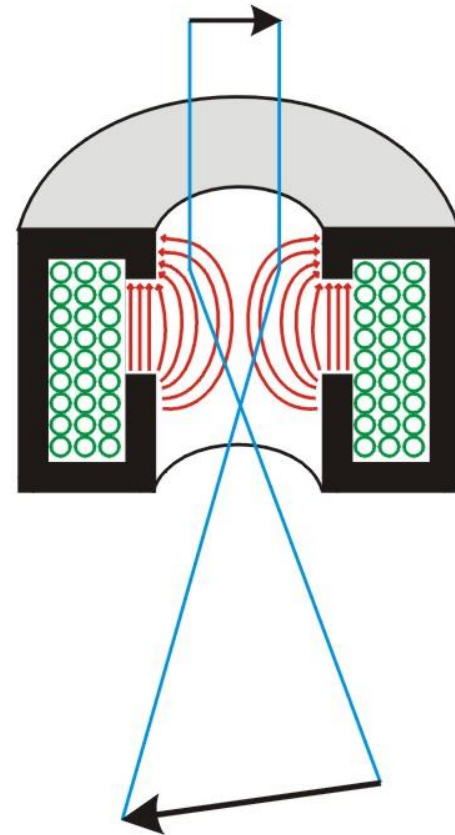
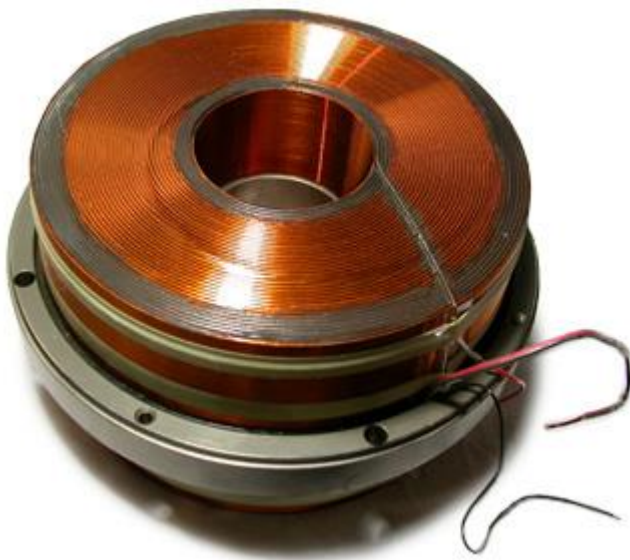
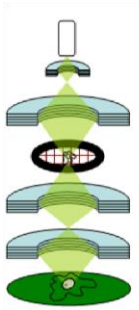
→ **Projection lens**

Phosphor screen





Electromagnetic lens



Microscope drawbacks

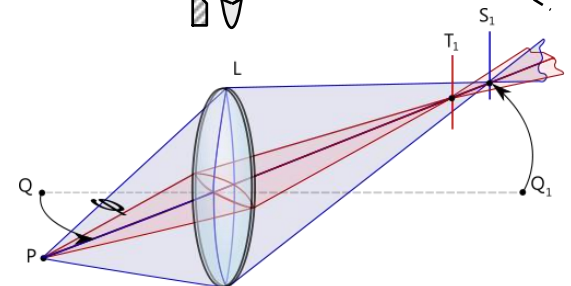
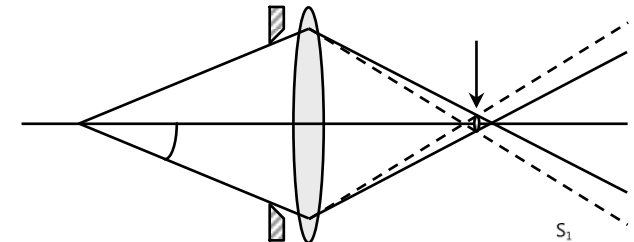
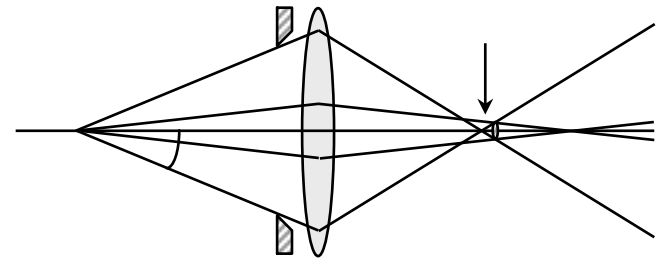
And solutions...

Stability:

- Mechanical vibrations
- Acoustic vibrations
- Magnetic fields
- Electron source
- Lens current

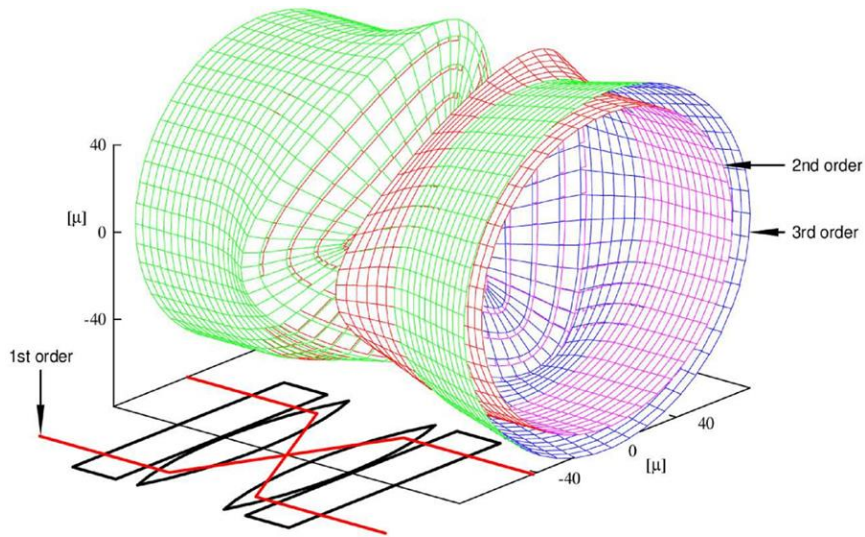
Lens:

- Spherical aberration
- Chromatic aberration
- Astigmatism

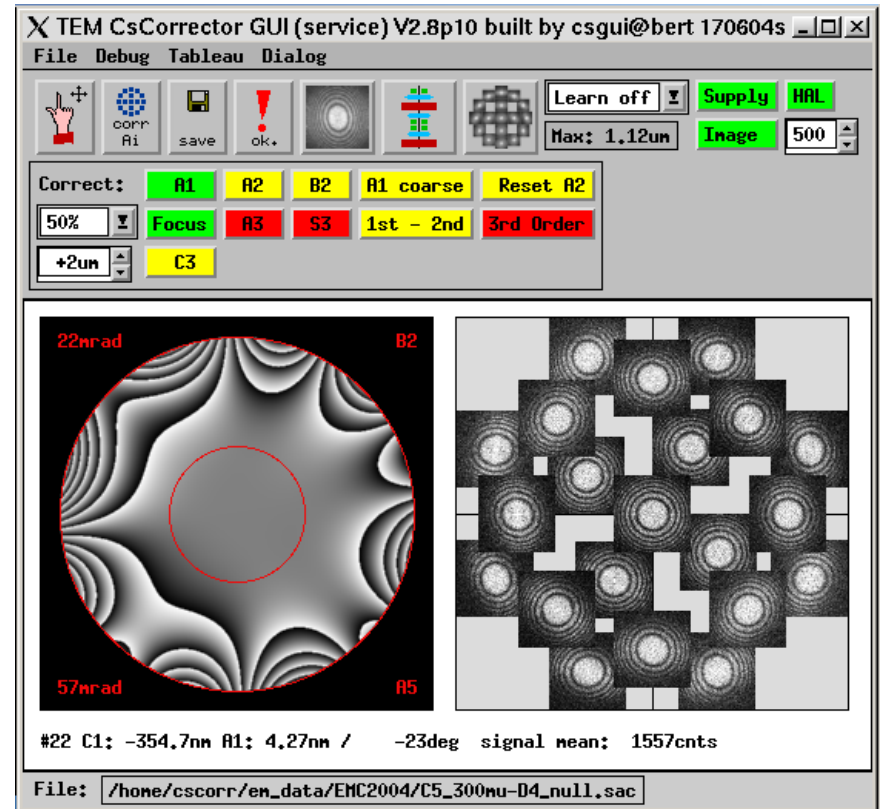


Lens spherical aberration

Correction



<http://salve-project.de>



<http://www.ceos-gmbh.de/English/products/residualsCEXCOR.html>

Composition of a TEM

Electron gun

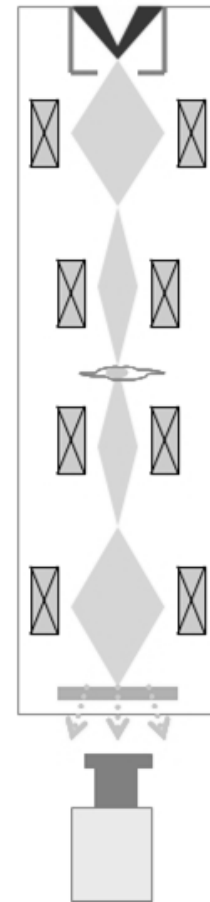
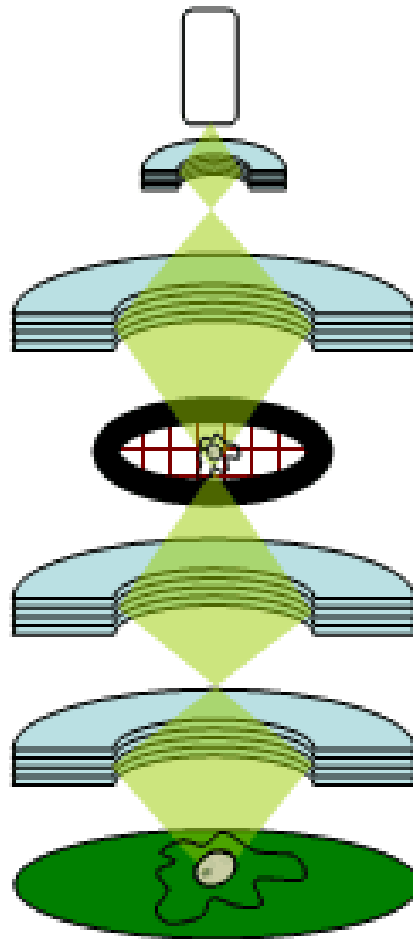
Condenser lens

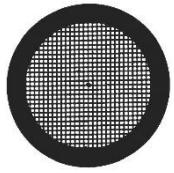
Sample grid

Objective lens

Projection lens

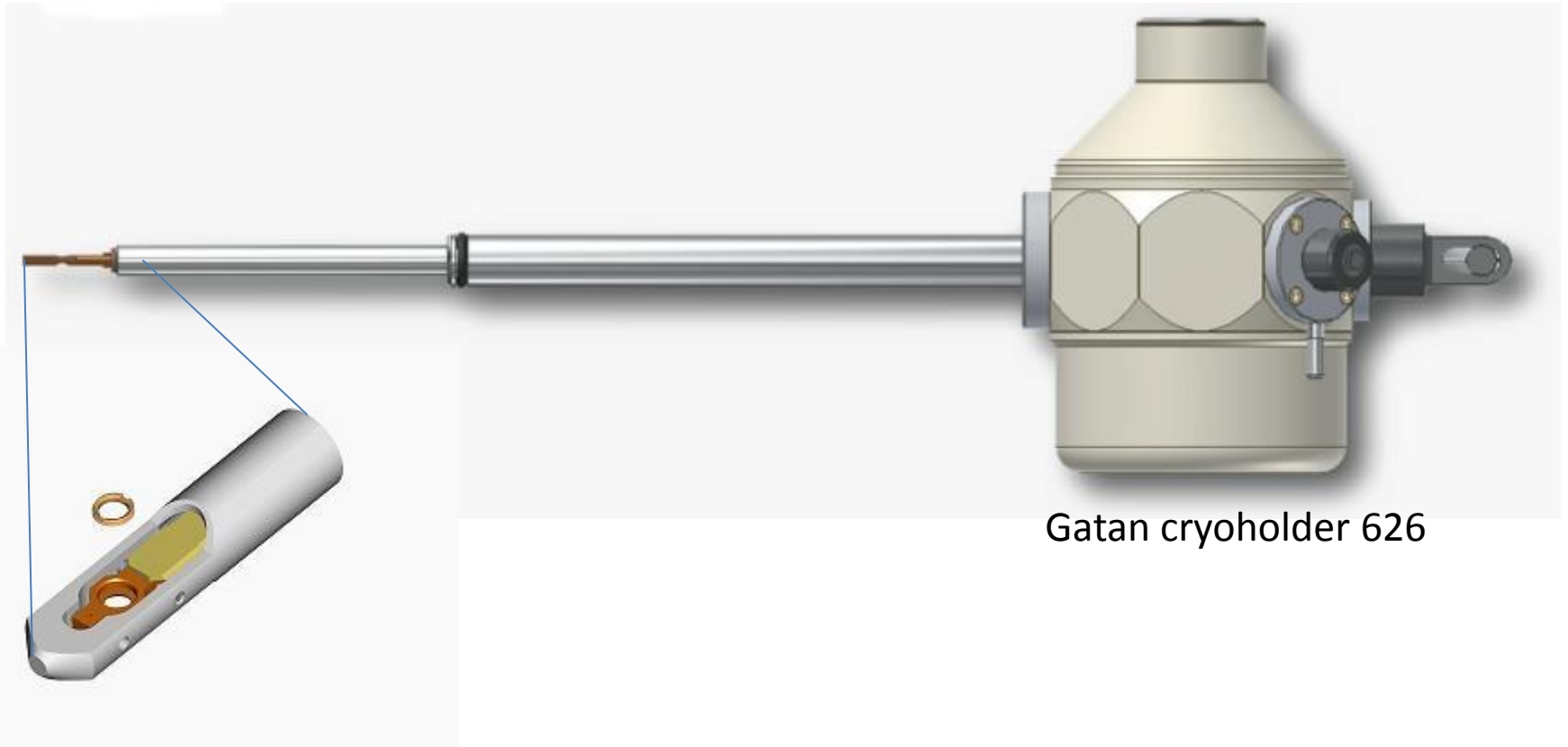
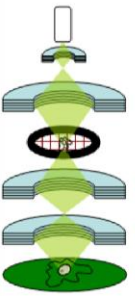
Phosphor screen





3 mm

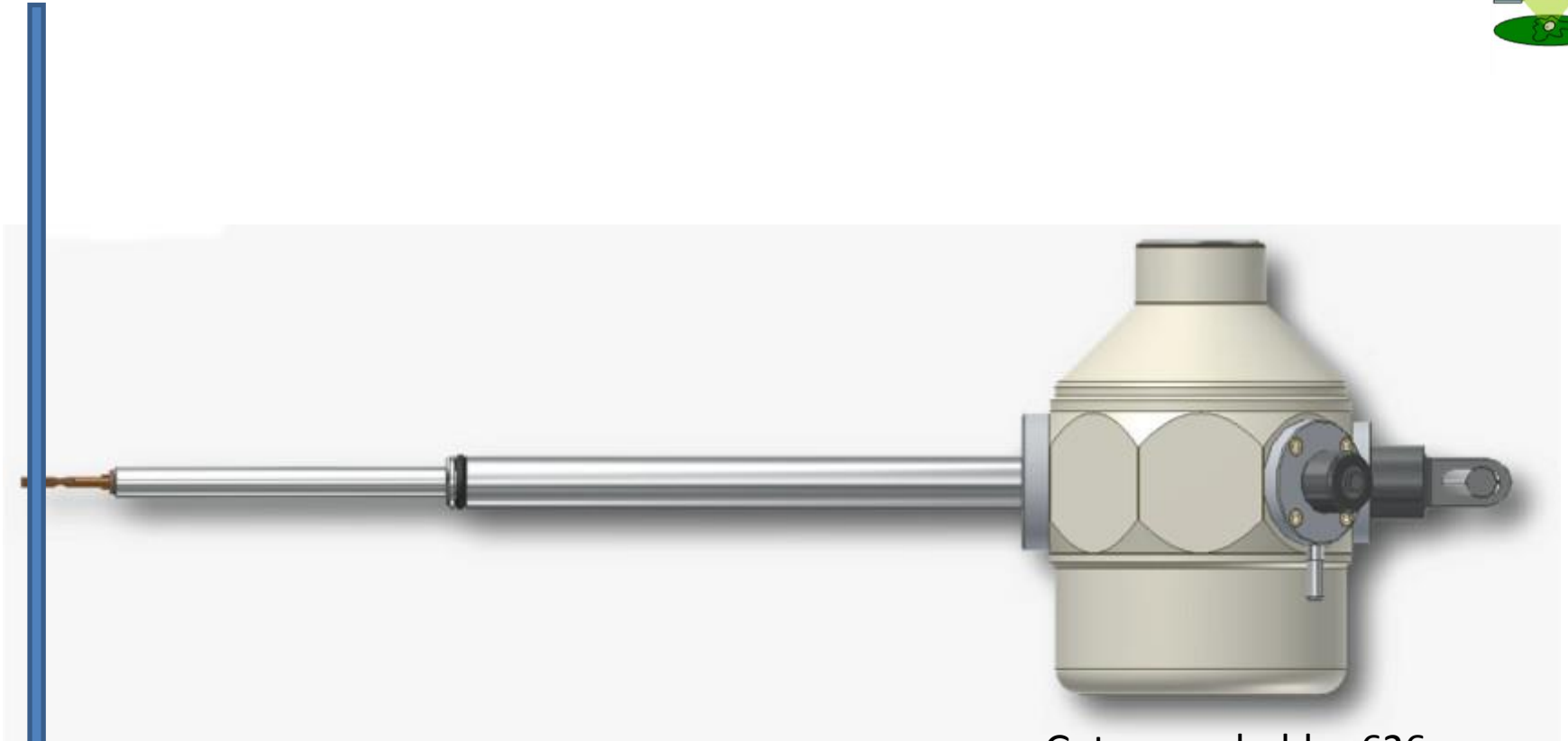
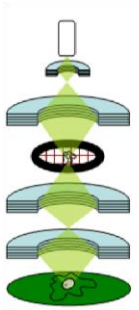
Sample holder



Gatan cryoholder 626



Sample holder



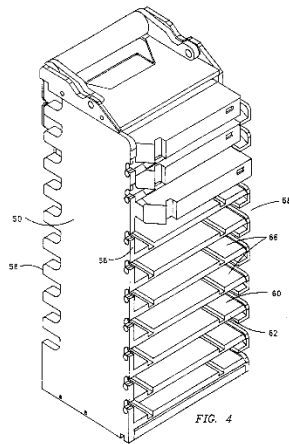
Gatan cryoholder 626



Inside the microscope

Outside the microscope

Autoloader (FEI)



Composition of a TEM

Electron gun

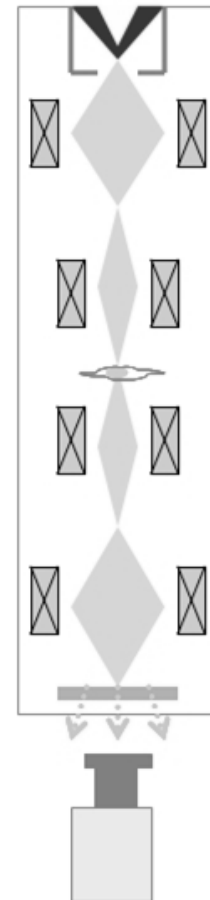
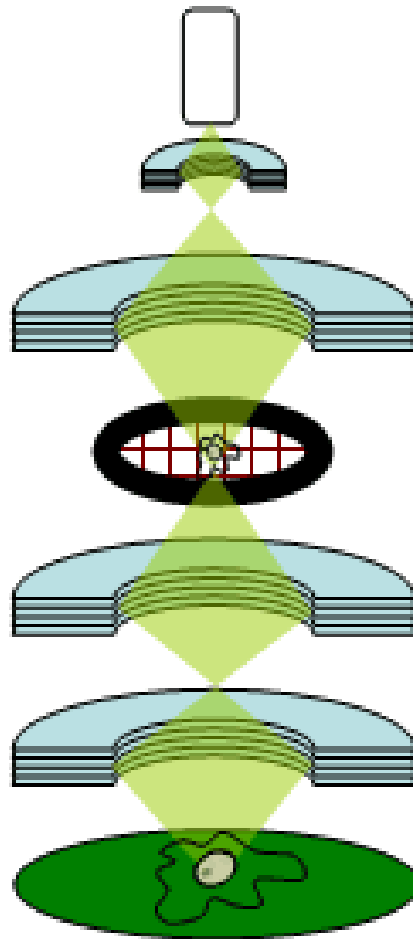
Condenser lens

Sample grid

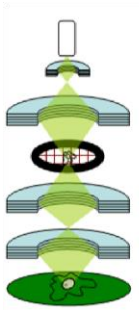
Objective lens

Projection lens

Phosphor screen



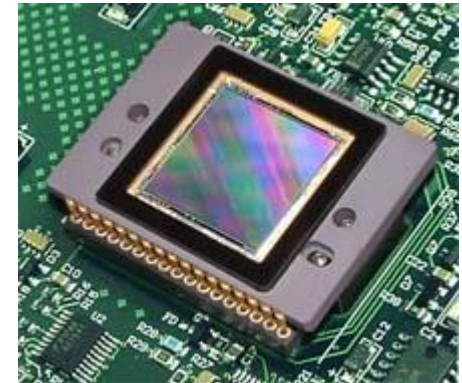
Visualization



Screen



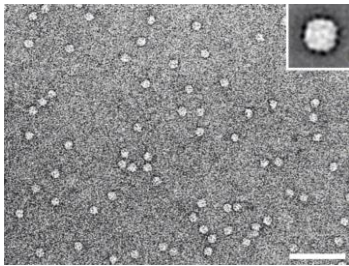
Negatives



Digital detector

Digitalization of images

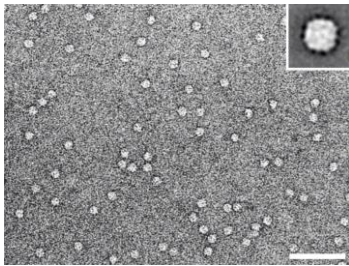
Film sensitive
to electrons



Pixel size = $8\text{ }\mu\text{m}$
1 image / s

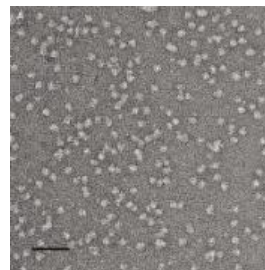
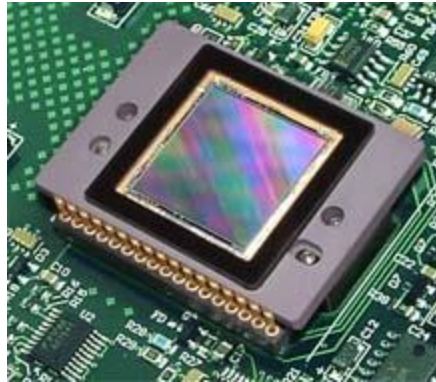
Digitalization of images

Film sensitive
to electrons



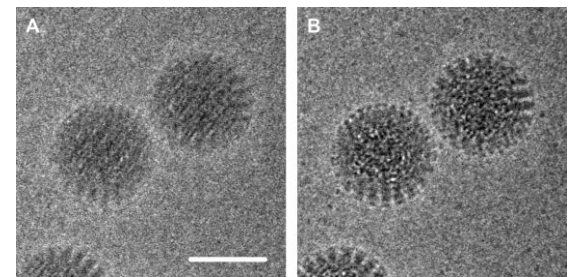
Pixel size = $8\ \mu\text{m}$
1 image / s

CCD



$14\ \mu\text{m}$
1 image / s

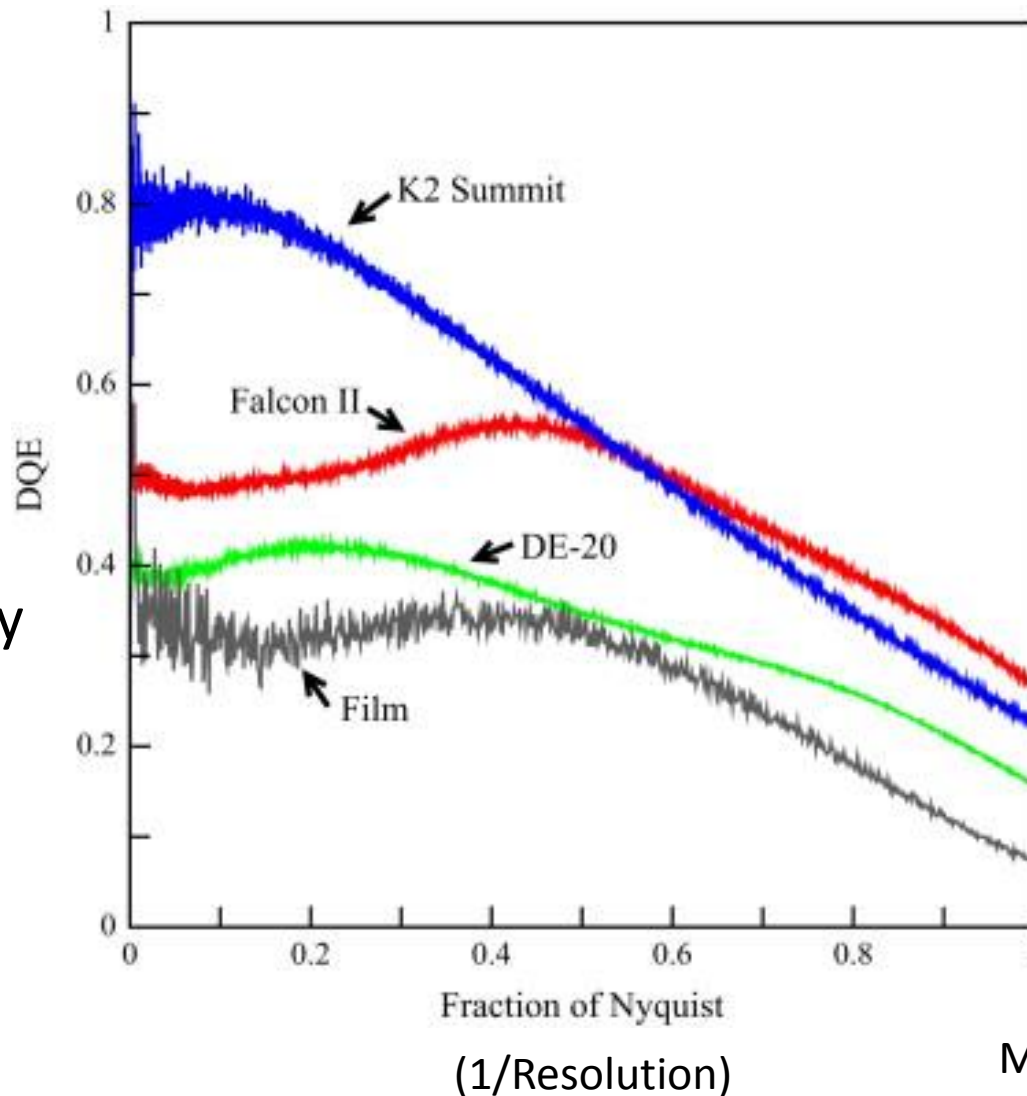
DED



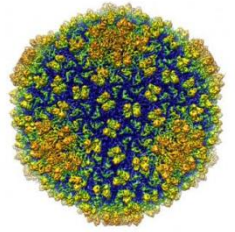
$5\ \mu\text{m}$
20 images / s

Sensitivity Film vs Digital detector

Detective
quantum
efficiency
 $\approx$ Sensitivity



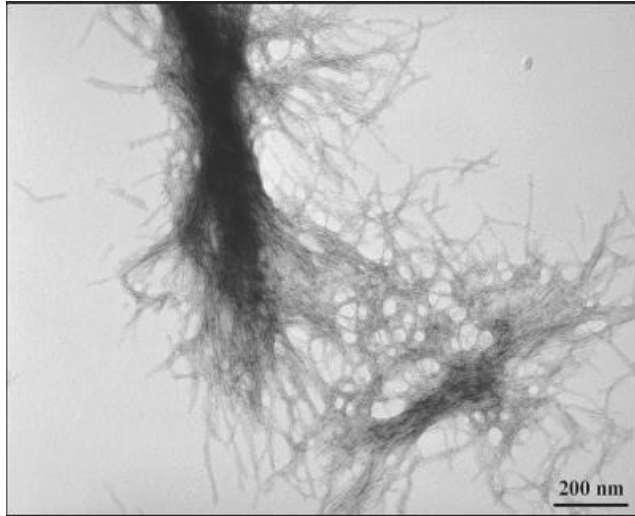
Electron microscopy



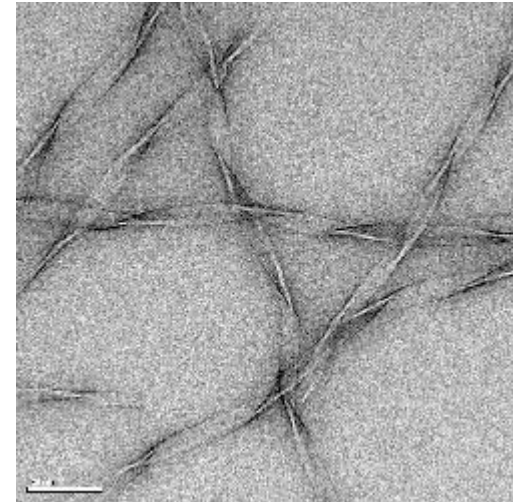
What structural informations
can you get ?

Qualitative analysis

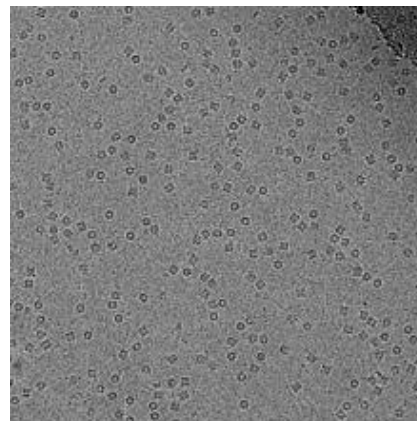
TEM



Agreggation



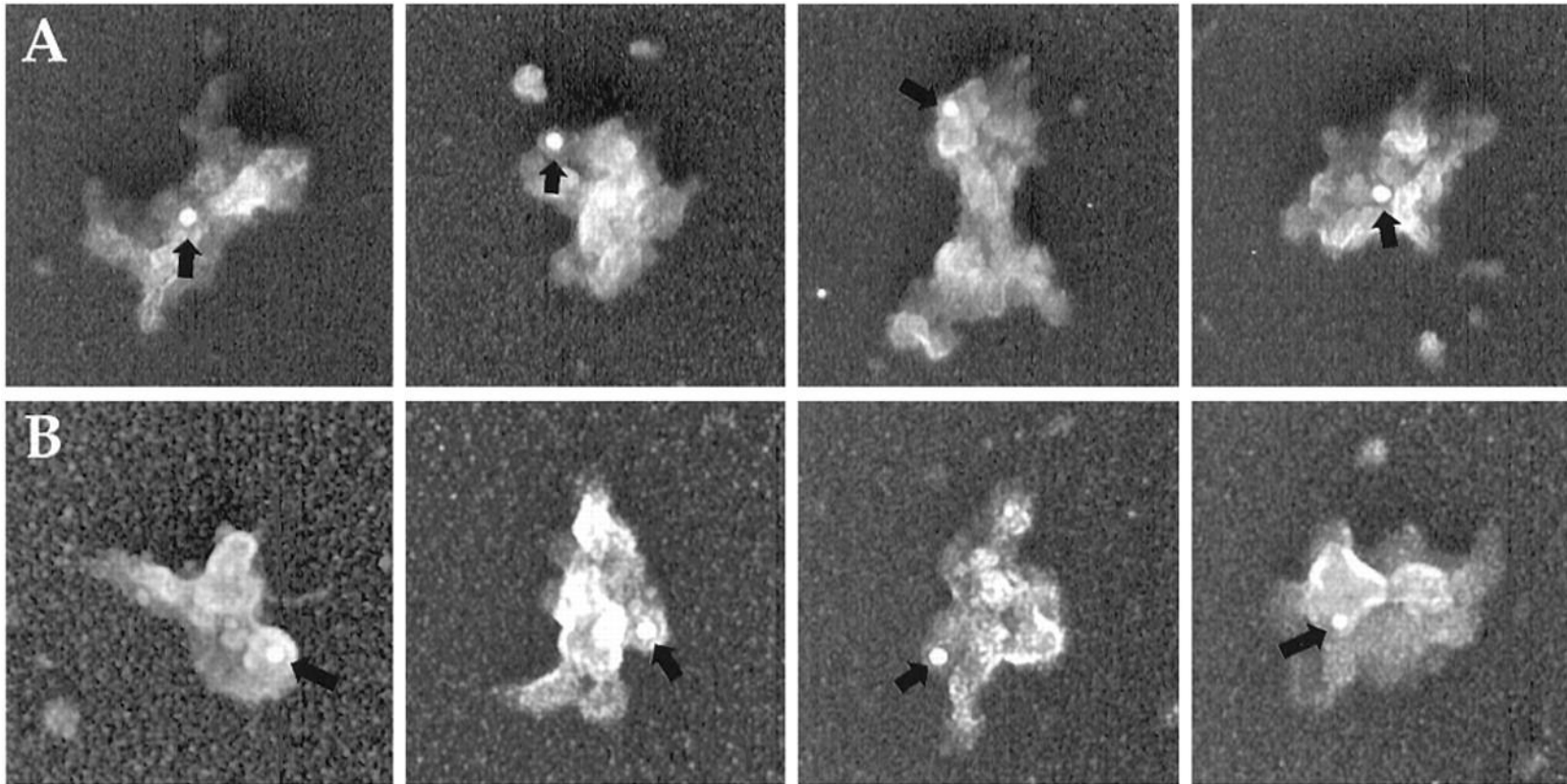
Filaments



Soluble protein

Localize a protein in a complex

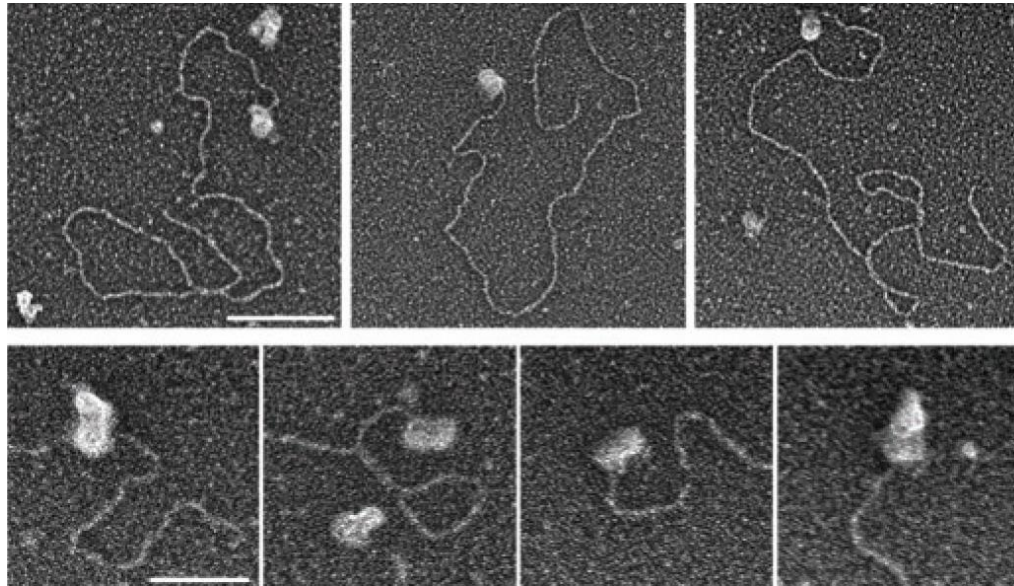
Human spliceosome (SEM)



Zhou et al. PNAS 2002

DNA-proteins interactions

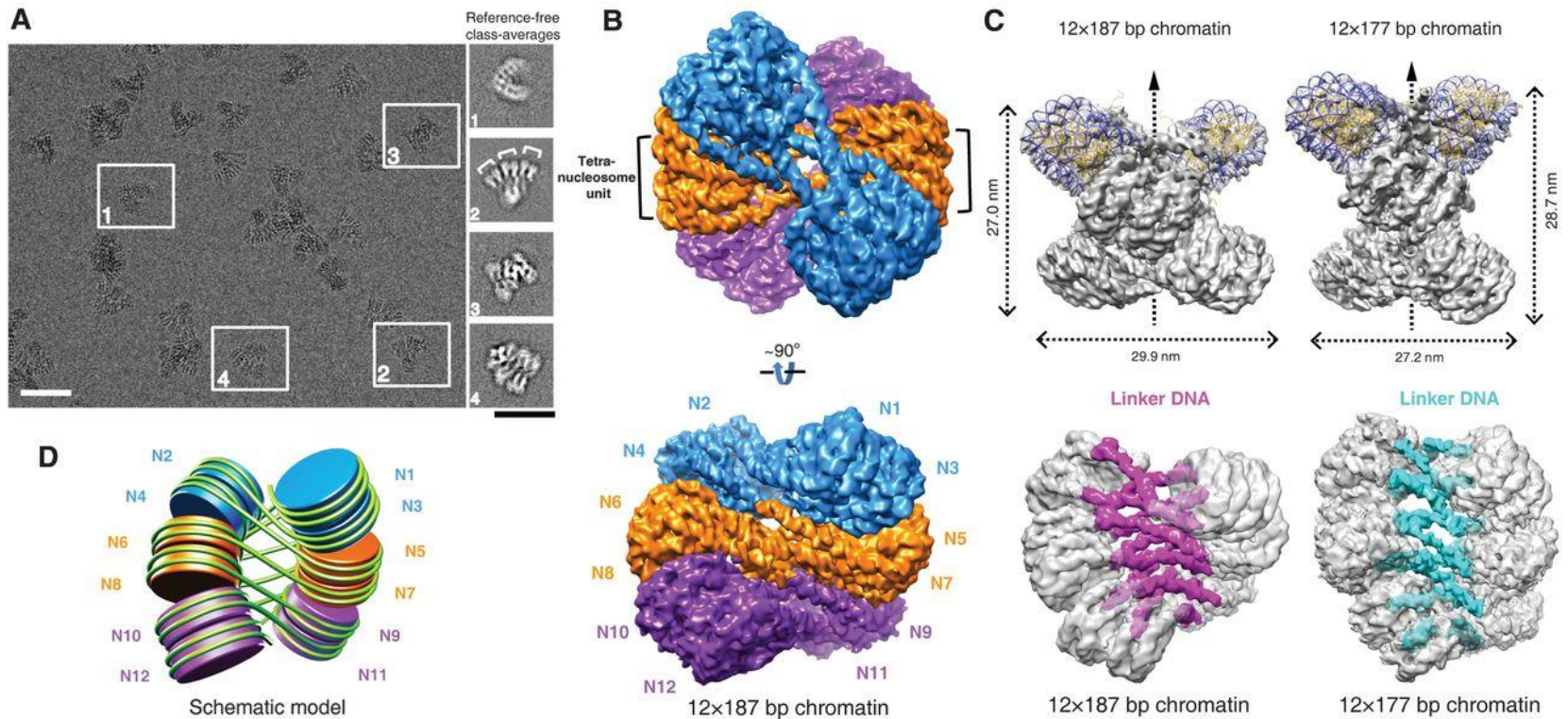
SEM



Thorslund et al, 2010

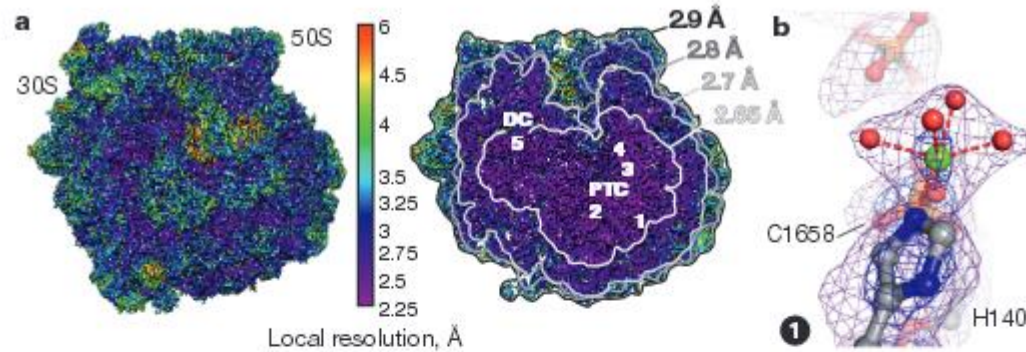
DNA-proteins interactions

TEM



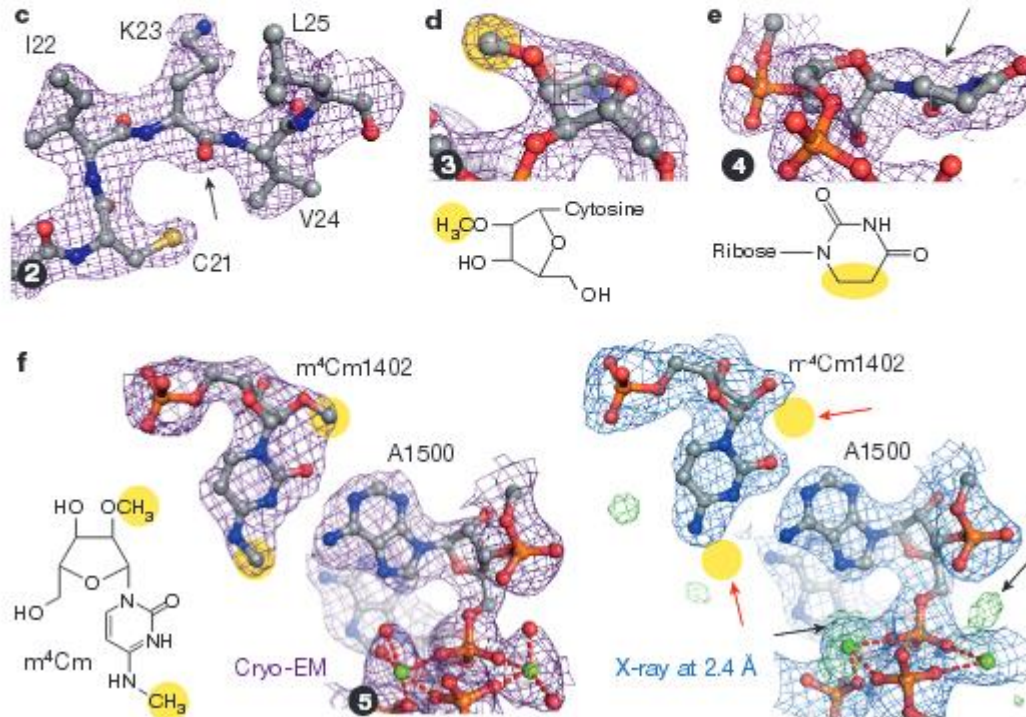
Song et al, 2014

RNA-proteins interactions



TEM

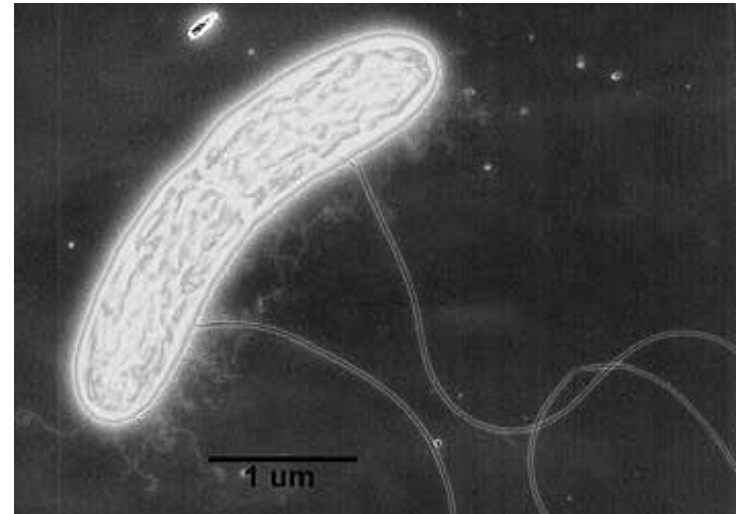
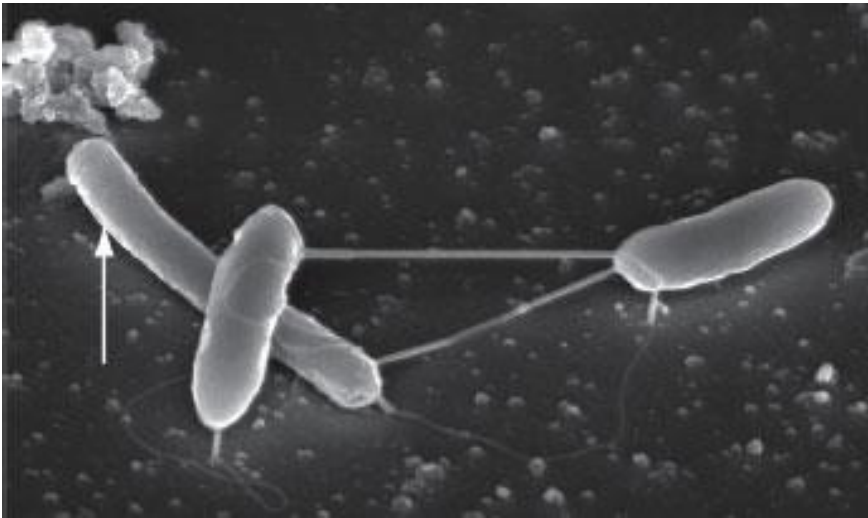
< 3 Å résolution



Fischer et al, Nature, 2015

Nanowires

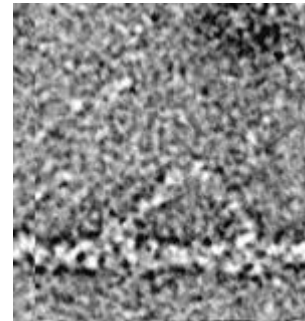
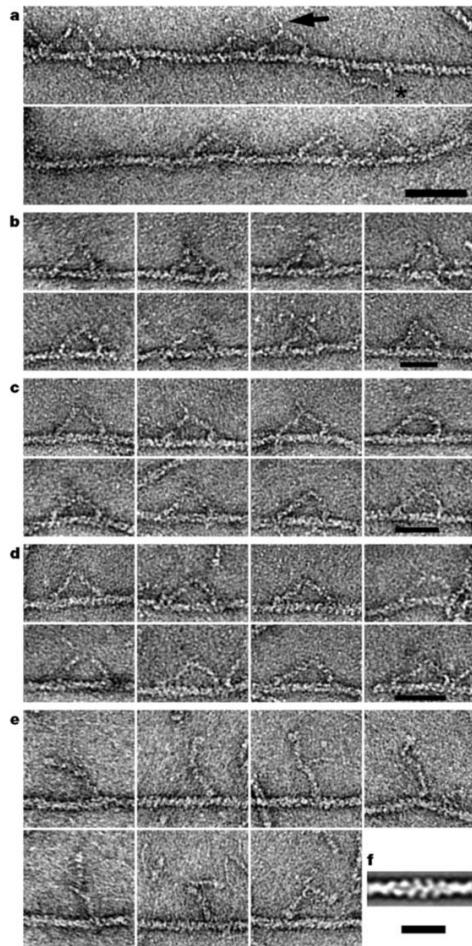
SEM & TEM



Geobacter metallireducens

Conformational changes

TEM

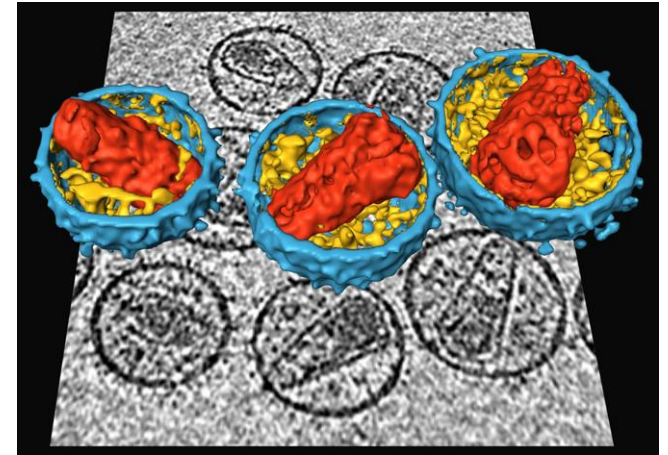
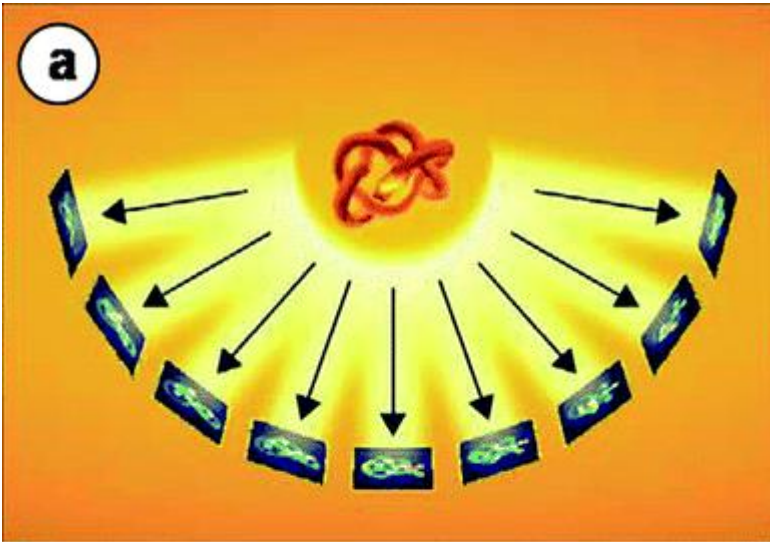


The Muscle Group, Leeds 2000

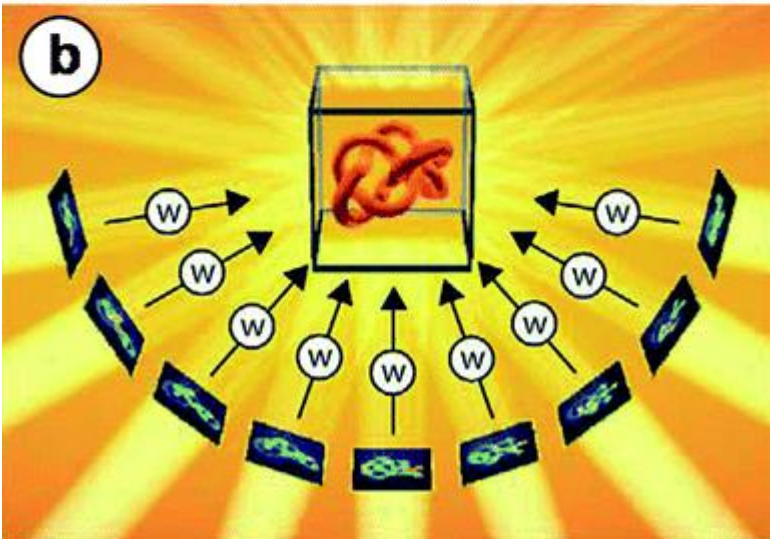
Walker et al, Nature, 2000

Tomography

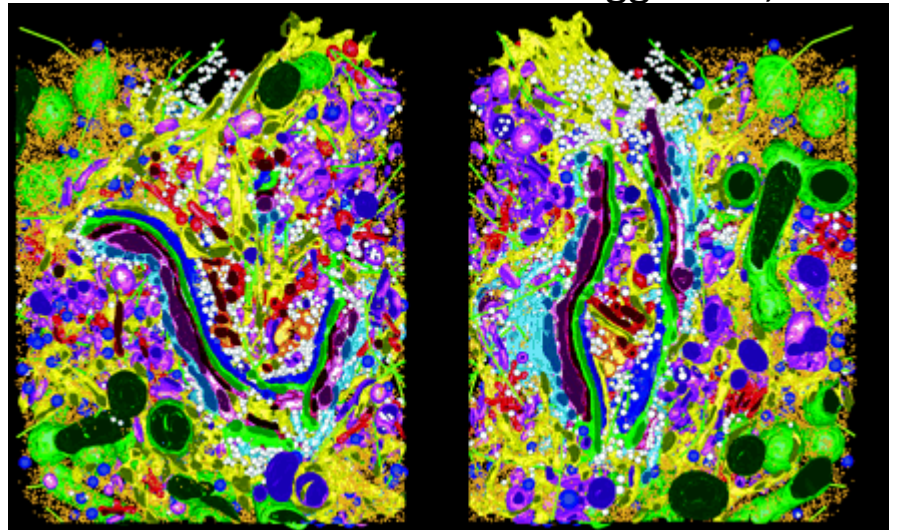
TEM



Briggs et al, 2006

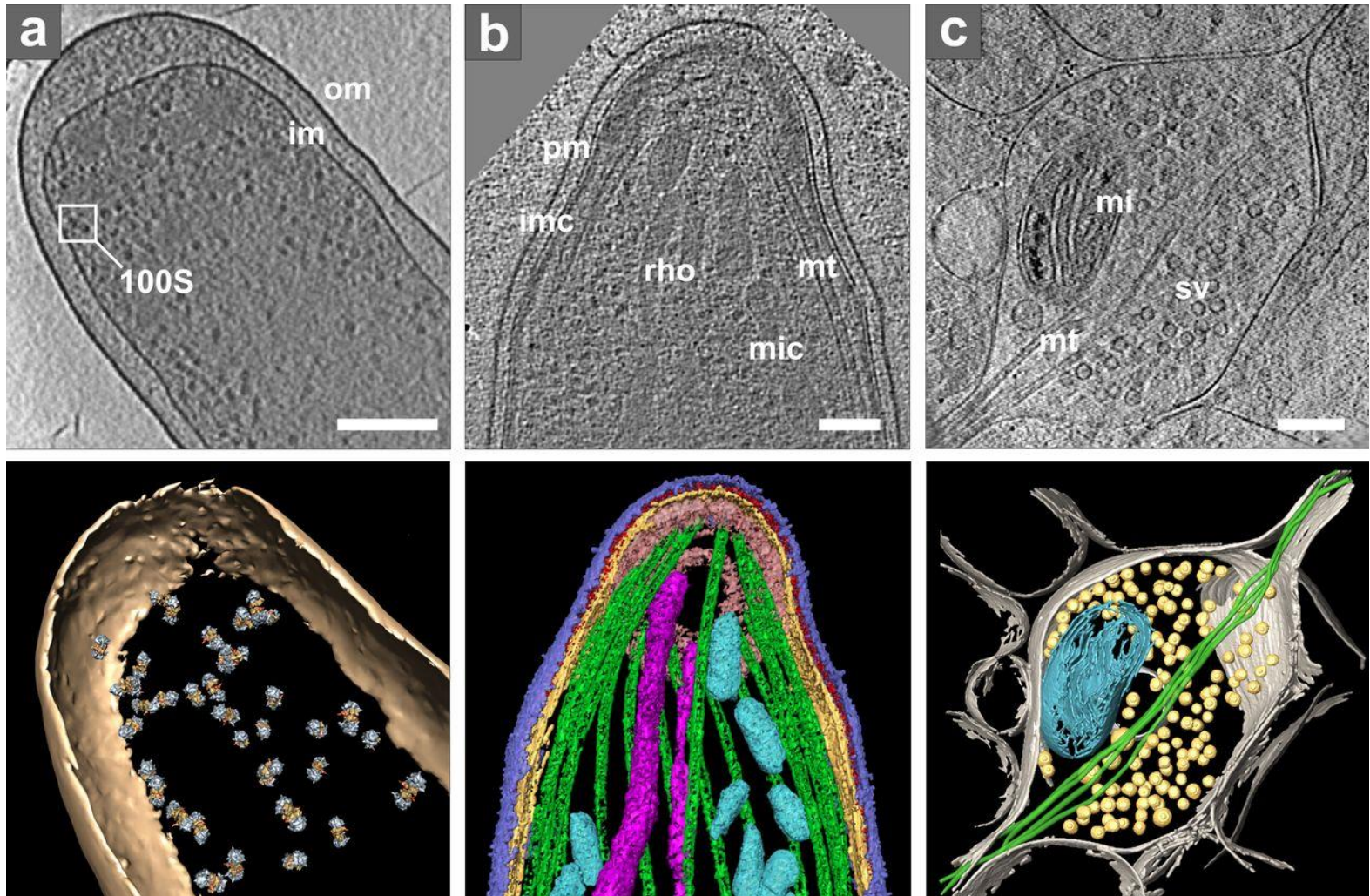


Baumeister, 2004



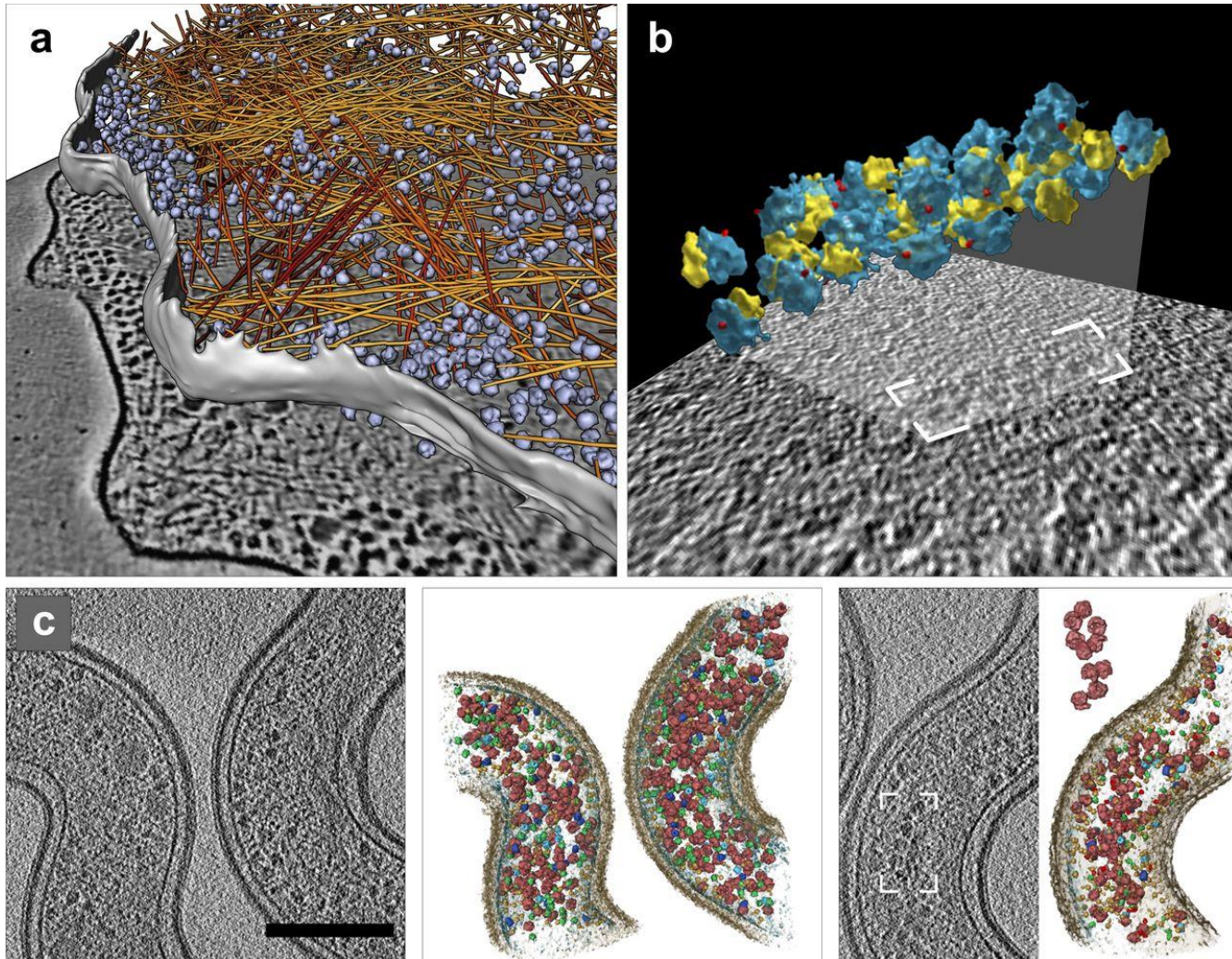
Marsh et al, 2001

Cryo-ET on whole cell or sections



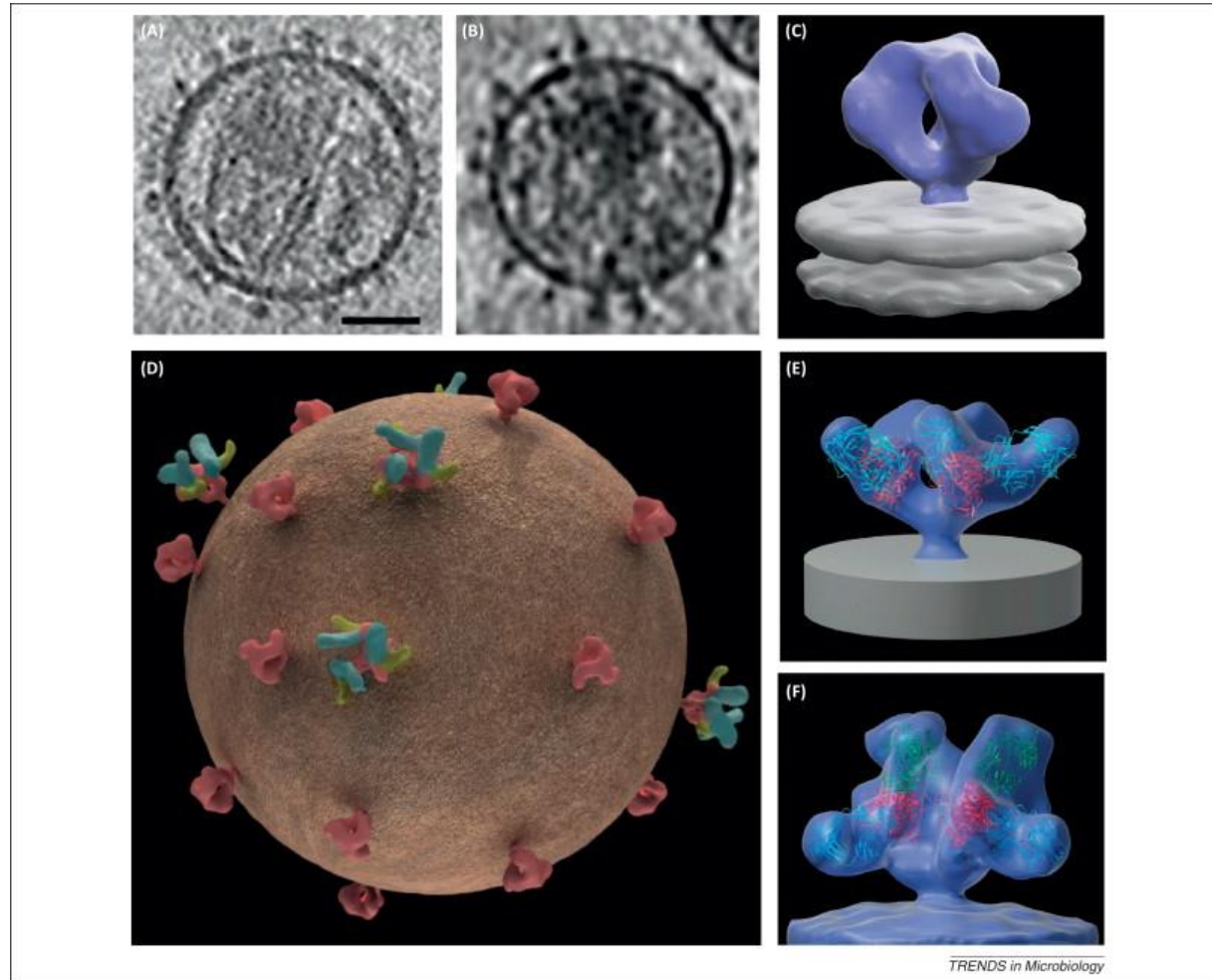
Lučić et al, 2013

Segmentation – Template matching



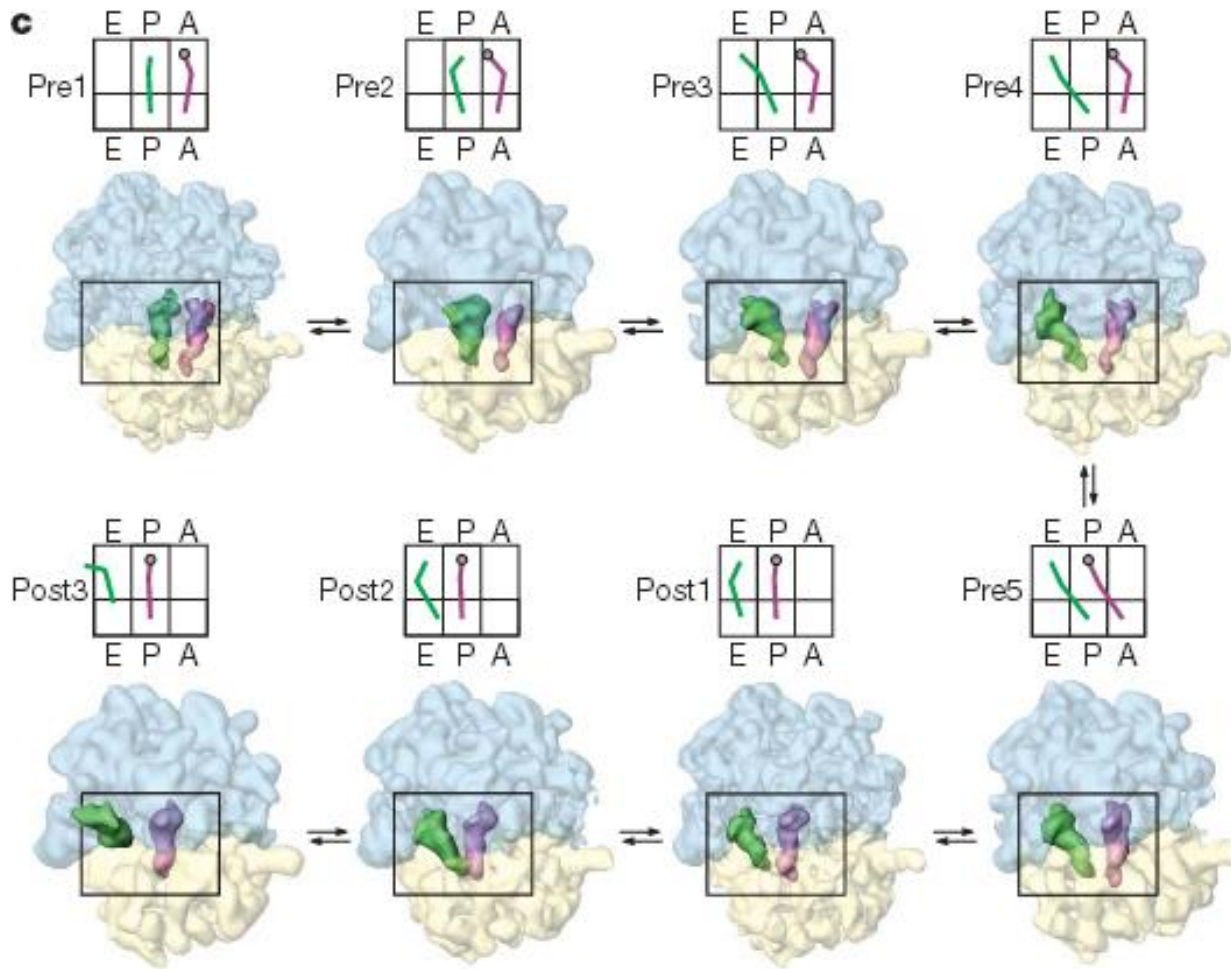
Lučić et al, 2013

Sub-tomogram averaging



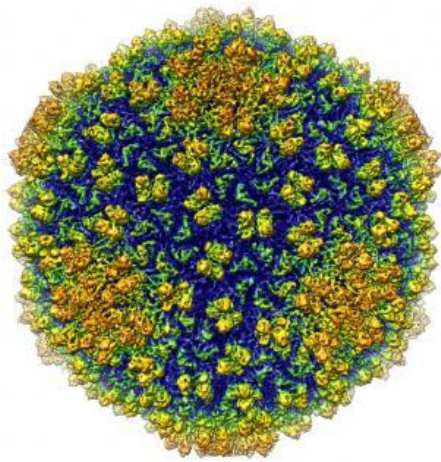
Earl et al, 2013

Dynamic of a mechanism



Fischer et al, 2010

Structure of a protein complex obtained by electron microscopy



Advantages :

- 3D structure of the protein complex
- (very) large complexes
- No crystal needed !
- Small amount of material

Drawbacks :

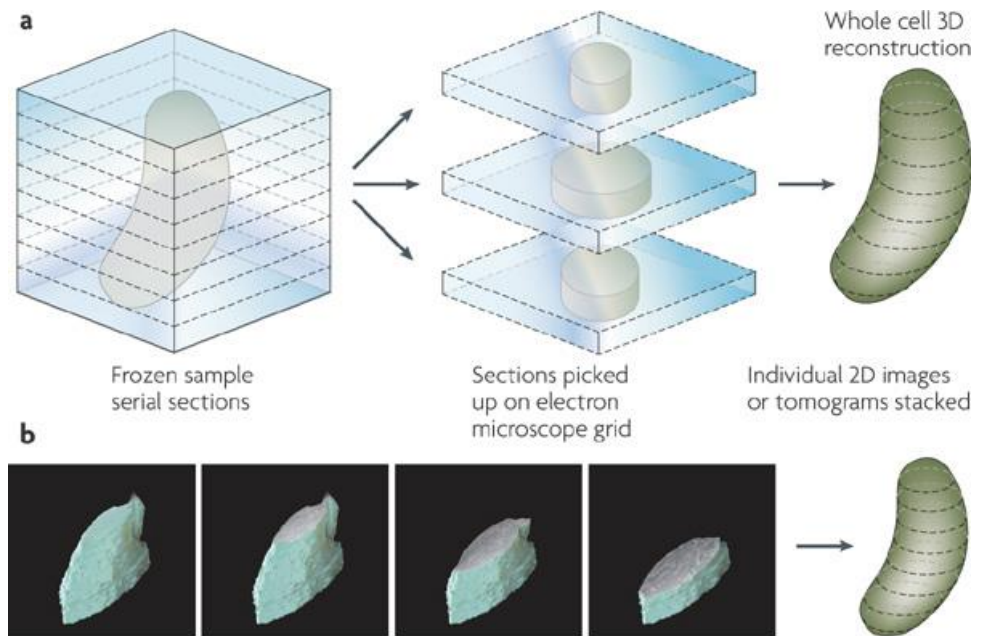
- Size limit (>100-300 kDa)
- High computing power
- Signal/noise ratio very low
- Dose /contrast compromise

Sample preparation

Negative stain



Frozen hydrated



Nature Reviews | Microbiology

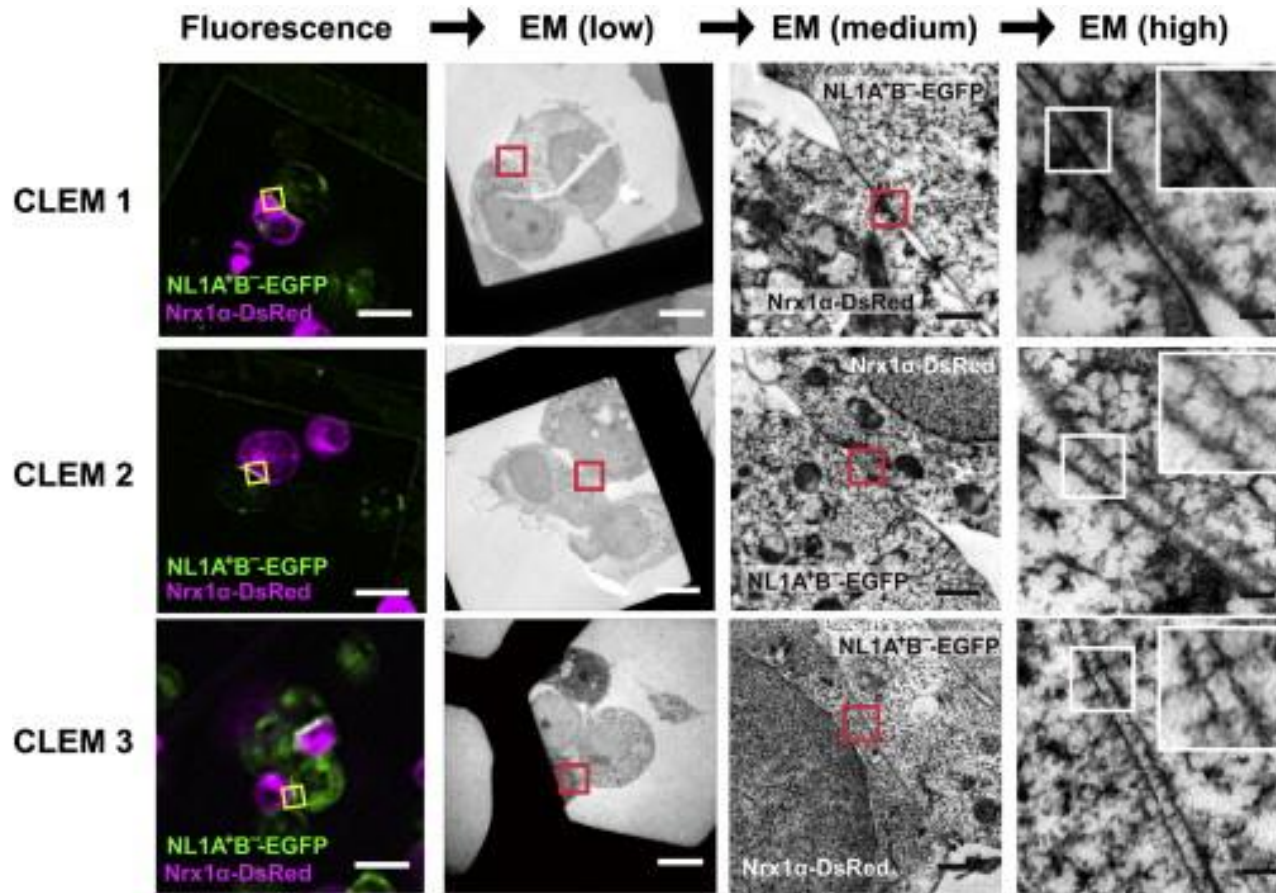
Durand et al, 2013

Data processing

2D → 3D →

CLEM

Correlative light-electron microscopy



Tanaka et al, Cell Reports, 2012

Questions ?